Inhibitory Effect of *Kochujang* Extracts on Chemically Induced Mutagenesis

So-Ja Kim, Keun-Ok Jung* and Kun-Young Park*

Department of Nutrition Catholic Sangii College, Andong 760-711, Korea *Department of Food Science and Nutrition, Pusan National University, Pusan 609-735, Korea

Abstract

Antimutagenic effects of 5 kinds of kochujang (Korean red pepper soybean paste) samples compared with doenjang (Korean soy paste) were studied using the Ames test with Salmonella typhimurium TA100 and the SOS chromotest with E. coli PQ37. The antimutagenic effects of methanol extracts from red pepper powder and meju (fermented soybean) powder, the major ingredients of the kochujang, were also evaluated for the mutagenicity of aflatoxin B_I (AFB_I) in the Ames assay. The methanol extracts from the kochujang samples showed lower antimutagenicities than those of doenjang against AFB_I and N-methyl-N'-nitro-N-nitrosoguanidine (MNNG) in the Ames assay. Traditional kochujang I and II exhibited strong antimutagenic activity against AFB_I and MNNG. The traditional kochujang III also effectively reduced the mutagenicity induced by MNNG. The antimutagenic effects of the kochujang samples against MNNG were also observed in the SOS chromotest system with the same fashions as shown in the Ames mutagenicity test. The methanol extracts from meju powder had the strongest inhibitory effects on mutagenicity induced by AFB_I, however, those from red pepper powder showed lower inhibition rate than kochujang. These results suggest that traditional kochujang exhibit higher antimutagenic activity than the commercial variety, and that meju powder seems to be one of the major antimutagenic components in kochujang.

Key words: kochujang, Ames test, SOS chromotest, antimutagenicity

INTRODUCTION

Kochujang (red pepper soybean paste) is a traditional Korean fermented food, which has been eaten with doenjang (soy paste) for a long time in Korea. It has played an important role in providing the specific taste and flavor in foods. Various tastes of kochujang have originated from the fermentation of raw materials such as soybean, starch sources and red pepper powder (1–5). The compound primarily responsible for the pungency of kochujang is capsaicin in red pepper powder. Kawada et al. (6,7) reported that capsaicin enhanced lipid and energy metabolism. In humans, Lim et al. (8) found that the ingestion of red pepper powder stimulated carbohydrate oxidation at rest and during exercise.

Kochujang can be classified into two groups, traditional kochujang using meju (by the conventional method) and commercial one (by the convenient method) using koji or bacterial enzyme. Generally traditional kochujang is made of glutinous rice, meju (naturally fermented soy paste), red pepper powder and salt, which is fermented by enzymatic reaction of bacteria or yeast. The malt is an optional ingredient that may be used to saccharify glutinous rice. Meju makes the difference between traditional kochujang and the commercial variety. In commercial kochujang, saccharogenic amylase or koji from glutinous rice which had already been incubated with Aspergillus oryzae is added. Differences in quality of kochujang is due to the ratio of raw material, fermentation time, and meshing method. Also

it varies with different localities and manufacturers.

Kochujang is fermented with red pepper powder and meju etc. that are known to have antimutagenic and the anticancer properties (9–16). Several studies indicated that red pepper powder revealed antimutagenic activity(9–11). It was reported that some components of soybean, especially trypsin inhibitor, isoflavones, saponin and phytic acid etc., showed anticancer effects (12–16). There was a report that meju which was fermented soybean cake showed antimutagenic effect against AFB₁ (17).

In this study, to determine the antimutagenic effects of methanol extracts from 5 varieties of *kochujang* (3 traditional and 2 commercial product), compared with *doenjang*, the Ames test and the SOS chromotest were carried out. The effects of methanol extracts from red pepper powder and *meju* powder, the major ingredients of *kochujang*, were also evaluated on AFB₁—mediated mutagenicity in the Ames assay.

MATERIALS AND METHODS

Samples

Traditional kochujang I, II, doenjang, red pepper powder and meju were obtained from Sunchang traditional kochujang village (Dookeobi Food Co., Sunchang, Choenbuk). Traditional kochujang III were obtained from Andong. Commercial kochujang I (chungjungwon from Miwon, Co., Sunchang, Cheonbuk) and Commercial kochujang II (Hapyo, Co., Chungnam)

^{*}Corresponding author. E-mail: kunypark@hyowon.pusan.ac.kr

were purchased from local market in pusan. Korea.

Kochujang and doenjang samples were freeze dried and powdered, and meju was powdered. 20-folds of methanol was added to the powdered samples and extracted by shaking with 3 times. The methanol extracts were evaporated using a vacuum evaporator, concentrated, then dissolved in dimethyl sulfoxide (DMSO, Sigma Chemical Co., USA).

Mutagens

AFB₁ was purchased from Sigma Chemical Co., St. Louis, Mo. (USA) and dissolved in DMSO. MNNG was obtained from Aldrich Chemical Co., Milwaukee, WI (USA) and dissolved in distilled water.

Ames mutagenicity test

The Salmonella typhimurium TA100 bacterial strain, a histidine requiring mutant was provided by Dr. B. N. Ames, Univ. of California, Berkerly, CA, USA and was maintained as described by Marons and Ames (18). The genotype of the tester stain was checked routinely for their histidine requirement, deep rough (rfa) character, UV sensitivity (uvr B mutation) and the presence of R factor.

S9 mixture to activate the indirect mutagen, AFB₁, was also prepared by the method of Maron and Ames (18). The mutagenicity test (19,20) was carried out by a modified plate incorporation test (liquid preincubation of the organism with the test compound). In the preincubation test, 0.5 ml of S9 mixture (or 0.5 ml of phosphate buffer for direct mutagen, MNNG) was distributed into sterilized capped tubes in an ice bath and then 0.1 ml of test strain cultured overnight $(1-2 \times 10^9 \text{ cells/ml})$, 0.1 ml of test compound (50 µl of mutagens and 50 µl of methanol extracts) were added. The tubes were vortexed gently and preincubated at 37°C for 20 min. 2 ml of the top agar supplemented with L-histidine and D-biotin kept at 45°C were added to each tube and vortexed for 3 seconds. The resulting entire mixture was overlaid on the minimal agar plate. The plates were inverted and incubated at 37°C for 48 hrs and then the revertant bacterial colonies on each plate were counted. Dose response tests (18) of the mutagens on the tester strain were carried out to determine the regions revealing mutagenicity induced by the mutagens. Toxicity tests for the different levels of the methanol extracts were also carried out. The methanol extracts used for antimutagenicity test did not show any toxicity on the test strain.

SOS Chromotest

The modified method of Quillardet and Hofnung (21), and Baik and Ham (22) was employed. 50 μ l of frozen stock of E. coli PQ37 was added to 5 ml/L medium and incubated in shaking water bath at 37°C overnight, then it was inoculated to the 5 ml/L medium and incubated for 2 hrs at 37°C until absorbance at 660 nm reached 0.3 \sim 0.4, in the active culture. The obtained active culture was diluted 10 fold with L medium. 100 μ l of the diluted culture was distributed to the 2 series in the wells of 96 well plate. 20 μ l of methanol extracts that was treated with mutagen (10 μ l methanol extracts +10 μ l mutagen)

were added, and then the SOS response was induced at 37° C for 90 min. $100\,\mu$ l of ONPG (o-nitrophenyl- β -D-galactopyranoside) and $100\,\mu$ l of PNPP (p-nitrophenyl phosphate disodium) were added to each set of the wells to determine the activities of β -galactosidase (β -G) and akaline phosphatase (A-P), respectively.

After the color development for 30 min., $100\,\mu$ l of $1.5\,\mathrm{M}$ Na₂CO₃ and $50\,\mu$ l of $1\,\mathrm{M}$ HCl were added to stop the color developments of β -G and A-P, respectively. After $5\,\mathrm{min.}$, $50\,\mu$ l of $2\,\mathrm{M}$ Tris buffer was added to the A-P assay system to neutralize the HCl and then the absorbance of each well was read on Elyza reader at $420\,\mathrm{nm.}$ Units of enzyme activities were calculated using the formula outlined by Miller (23).

$$Eu = \frac{1000 \times OD_{420}}{t(min)}$$

Statistical analysis

Statistical analysis was performed by analysis of variance. Significant differences between treatment means were determined by using Duncan's multiple range test (24).

RESULTS AND DISCUSSION

The methanol extracts from various kinds of *kochujang* and *doenjang* showed antimutagenic activity against AFB₁ in *Salmonella typhimurium* TA100 (Table 1). *Doenjang* (Korean fermented soy paste) has already demonstrated its strong antimutagenic and anticancer activities (25–34). Compared to the *doenjang* sample, extracts from *kochujang* exhibited lower antimutagenicities. However, AFB₁ induced mutagenesis was especially inhibited by traditional *kochujang* II which inhibited

Table 1. Effect of methanol extracts from various kinds of kochujang and doenjang on the mutagenicity induced by aflatoxin B₁ (AFB₁, 2.0 µg/plate) in Salmonella typhimurium TA100

Treatment	Revertants/plate (level of sample, mg/plate)			
	1.25	2.5		
Spontaneous	92± 1 ⁶⁾			
Control	1245 ± 32^{a}			
Traditional kochujang I 1)	$739 \pm 23^{d} (44)^{7}$	$640 \pm 15^{\circ}$ (52)		
Traditional kochujang II ²⁾	711 ± 15^{d} (46)	$561 \pm 17^{\circ} (59)$		
Traditional kochujang III ³⁾	$874 \pm 17^{\circ}$ (32)	$774 \pm 22^{\circ}$ (41)		
Commercial kochujang I 4)	918 ± 14^{c} (28)	$826 \pm 11^{\circ} (36)$		
Commercial kochujang II ⁵⁾	1025 ± 24^{b} (19)	917 ± 16^{b} (28)		
Doenjang	$655 \pm 16^{\circ}$ (51)	$485 \pm 28^{\circ}$ (66)		

¹⁾Sunchang traditional *kochujang* prepared with glutenous rice powder

²⁰Sunchang traditional *kochujang* prepared with glutenous rice powder and malt

³⁾Andong traditional kochujang

⁴⁾Sunchang chungjungwon

⁵⁾Haepyo

⁶⁾The values are means of 3 replicates±SD

The values in parentheses are the inhibition rates (%)

a-'Means with the different letters beside data are significantly different at the p<0.01 level of significance as determined by Duncan's multiple range test.

the mutagenicity of AFB₁ by 59% at 2.5 mg/plate concentration. The inhibition rates for AFB₁ were $41 \sim 59\%$ for the traditional kochujang samples while the inhibition rates of commercial kochujang extracts were 28~36% on the 2.5 mg/plate. Among the traditional kochujang samples. Sunchang traditional kochujang had higher antimutagenicity than the Andong variety. Jung et al. (35) reported that kochujang had an inhibitory effect similar to doeniang on the mutagenicity induced by AFB₁ in Salmonella typhimurium TA98 and homemade kochujang had higher antimutagenicity than commercial kochujang. Therefore, the traditional kochujang has higher antimutagenicity than commercial kochujang, even though the methanol extract of kochujang is less effective than that of doenjang. It is thought that the differences in the activity between the traditional kochuiang and commercial variety depended on the ratio of raw material, fermentation time and meshing method. Table 2 showed the antimutagenicity of the methanol extracts of kochujang samples and doenjang against MNNG. The methanol extracts from traditional kochujang I and II exhibited weaker antimutagenic effects against direct mutagen, MNNG, than against indirect mutagen, AFB₁. However, on the 2.5 mg/plate, both of them inhibited the mutagenicity of MNNG by above 40%.

Table 2. Effect of methanol extracts from various kinds of *kochu-jang* and *doenjang* on the mutagenicity induced by N-methyl-N'-nitro-N-nitrosoguanidine (MNNG, 0.3 µg/plate) in *Salmonella typhi-murium* TA100

Treatment	Revertants/plate (level of sample, mg/plate)			
	1.25	2.5		
Spontaneous	105± 7 ⁶⁾			
Control	$1333 \pm 18^{\circ}$			
Traditional kochujang I 1)	$923 \pm 25^{cd} (33)^{7)}$	$807 \pm 26^{\text{de}}$ (43)		
Traditional kochujang II ²⁾	915 ± 28^{cd} (34)	$767 \pm 17^{\circ}$ (46)		
Traditional kochujang III ³⁾	$954 \pm 15^{\circ}$ (31)	833 ± 17^{d} (41)		
Commercial kochujang I 41	$1052\pm21^{\rm b}$ (23)	$924 \pm 19^{\circ}$ (33)		
Commercial kochujang II ⁵⁾	1095 ± 21^{b} (19)	$973 \pm 12^{b} (29)$		
Doenjang	896 ± 15^{d} (36)	672±35 ^f (54)		

^{1~7)}The explanation is the same as shown in Table 1.

It is known that NaCl in foods plays a cocarcinogenic role in the presence of MNNG (10, 36-37). Although kochujang contained high level (10~15%) of NaCl, its extracts inhibited the mutagenicity of MNNG. These results suggest that kochujang samples have antimutagenic effects on both AFB₁ and MNNG. In the SOS chromotest, the kochujang extracts also revealed antimutagenic effects on the direct mutagen, MNNG (Table 3). 43~45% of SOS response induced by MNNG were blocked by adding 100 µg/assay of traditional kochujang samples but commercial kochulang extracts showed 27~33% inhibition rate (p<0.01). Patterns of antimutagenic effects were almost the same as shown in the Ames mutagenicity test. From the above studies, the methanol extracts of traditional kochujang concluded to reveal higher antimutagenic activities than those of commercial variety against AFB1 and MNNG in Ames test, and MNNG in SOS chromotest

It is reported that the quality of *kochujang* is influenced by several factors such as ratio of raw material, fermentation time and meshing methods (3). Among the ingredients of *kochujang*, red pepper powder and *meju* (fermented soybean cake) powder that are known for their antimutagenic and anticancer activities (9–11,17) were evaluated on AFB₁-mediated mutagenicity as compared with *kochujang* in the Ames assay (Table 4). At the 2.5 mg/plate concentration, the methanol extract of *meju* powder exhibited 83% antimutagenicity against AFB₁, while that of red pepper powder showed 29% antimutagenicity. *Meju* pow-

Table 4. Effect of methanol extracts from *kochujang*, red pepper powder and meju on the mutagenicity induced by aflatoxin B_1 (AFB₁, 2.0 μ g/plate) in *Salmonella typhimurium* TA100

Treatment (mg/plate)		Revertants/plate	Inhibition rate (%)	
Spon		107± 6		
Control		1246±23		
kochujang	1.25	713±17	47	
	2.5	642 ± 15	53	
Red pepper powder	1.25	918 ± 45	29	
	2.5	1027 ± 21	18	
Meiu powder	1.25	414 ± 23	73	
V	2.5	295 ± 23	83	

¹⁾The values are means of 3 replicates ±SD

Table 3. SOS response of methanol extracts(100 µg/assay) from various kinds of *kochujang* and *doenjang* against N-methyl-N'-nitro-N-nitrosoguanidine (MNNG, 40 ng/assay) in *E. coli* PQ37

Treatment	β-Glactosidase(β)		Alkaline phosphatase(p)		(0) //e)	SOS induction	Inhibition rate
	OD_{420}	Unit	OD420	Unit	(β)/(ρ)	factor	(%)
Spon	0.467 ± 0.003^{6}	15.6	0.415±0.010	13.8	1.13	1.00	
Control	1.093 ± 0.048	36.4	0.049 ± 0.004	14.0	2.60^{a}	2.30	
Traditional kochujang I 1)	0.815±0.016	27.2	0.418±0.003	13.9	1.95°	1.73	44
Traditional kochujang II ²⁾	0.804 ± 0.010	26.8	0.414 ± 0.005	13.8	1.94°	1.72	45
Traditional kochujang III ³⁾	0.832 ± 0.009	27.7	0.422 ± 0.070	14.1	$1.97^{\rm c}$	1.74	43
Commercial kochujang I 4)	0.889 ± 0.010	29.6	0.423 ± 0.007	14.1	$2.10^{\rm b}$	1.86	33
Commercial kochujang II ⁵⁾	0.926 ± 0.016	30.9	0.421 ± 0.003	14.0	$2.20^{\rm b}$	1.95	27
Doenjang	0.725 ± 0.010	24.2	0.424 ± 0.006	14.1	1.70^{d}	1.50	62

The explanation is the same as shown in Table 1.

³ Means with the different letters beside data are significantly different at the p<0.01 level of significance as determined by Duncan's multiple range test.

a^{-d}Means with the different letters beside data are significantly different at the p<0.01 level of significance as determined by Duncan's multiple range test.</p>

der exerted stronger antimutagenicity than *kochujang* and red pepper powder. The above data suggest that the high antimutagenicity of *kochujang* probably results from some end products produced by the action of microorganisms during fermentation of the soybean or soybean itself. There was the report that *meju* showed an antimutagenic effect on AFB₁ (17). Thus, the amount of *meju* in *kochujang* can be one of the important factors that inhibit mutagen-induced mutagenesis.

In conclusion, the methanol extract of *kochujang* is less effective than that of *doenjang* in antimutagenic activity and traditional *kochujang* has higher antimutagenic activity than commercial *kochujang* according to the Ames test and the SOS chromotest. Further study is needed to identify the components and mechanisms for the differences of activity between traditional *kochujang* and commercial *kochujang*.

REFERNCES

- Moon, T. W. and Kim, Z. U.: Some chemical and physical characteristics and acceptability of kochujang from various starch sources. J. Kor. Agric. Chem. Soc., 31, 387 (1988)
- Woo, D. H. and Kim, Z. U.: Characteristics of improved kochuiang. J. Kor. Agric. Chem. Soc., 33, 161 (1990)
- Lee, T. S., Chun, M. S., Choi, J. Y. and Noh, B. S.: Changes of free sugars and free amino acids in kochujang with different mashing method. Food Biotechnol., 2, 102 (1993)
- Lee, H. Y., Park, K. H., Min, B. Y., Kim, J. P. and Chung, D. H.: Studies on the change of composition of sweet potato kochujang during fermentation. Kor. J. Food Sci. Technol., 10, 331 (1978)
- Chun, M. S., Lee, T. S. and Noh, B. S.: The changes in capsaicin, dihydrocapsaicin and cansanthin in *kochujang* with different mashing methods. *Food Biotechnol.*, 3, 104 (1994)
- Kawada, T., Hagihara, K. and Iwai, K.: Effects of capsaicin on lipid metabolism in rats fed a high fat diet. J. Nutr., 116, 1272 (1986)
- Kawada, T., Watanabe, T., Takakashi, T., Tanaka, T. and Iwai, K.: Capsaicin-induced beta-adrenergic action on energy metabolism in rats: influence of capsaicin on oxygen consumption, the respiratory quotient, and substrates utilization. *Proc. Soc. Exp. Biol. Med.*, 186, 250 (1986)
- Lim, K., Yoshioka, M., Kikuzato, S., Kiyonaga, A., Tanaka, H., Shindo, M. and Suzuki, M.: Dietary red pepper ingestion increases carbohydrate oxidation at rest and during exercise in runners. Med. Sci. Sports Exerc., 29, 355 (1997)
- Kim, S. H., Park, K. Y. and Suh, M. J.: Inhibitory effect of aflatoxin B₁ mediated mutagenicity by red pepper powder in Salmonella assay system. J. Korean Soc. Food Nutr., 20, 156 (1991)
- Kim, S. H.: Comutagenic and antimutagenic effects of kimchi components. Ph.D. thesis, Pusan National University (1991)
- Kweon, Y. M., Rhee, S. H. and Park, K. Y.: Antimutagenic effects of juice from peppers in Salmonella assay system. J. Korean Soc. Food Nutr., 24(3), 440 (1995)
- Jing, Y. and Waxman, S.: Structural requirements for differentiation-induction and growth-inhibition of mouse erythroleukemia cells by isoflavones. *Anticancer Res.*, 15, 1147 (1995)
- Rao, A. V. and Sung, M. K.: Saponins as anticarcinogens. J. Nutrition, 125, 717S (1995)
- Shamsuddin, A. M.: Inositol phosphates have novel anticancer function. I. Nutrition. 125, 7258 (1995)
- Yavelow, J., Finlay, T. H., Kennedy, A. R. and Troll, W.: Bowman-Birk soybean protease inhibitor as an anticarcinogen. Cancer Res.

- (Suppl.), 43, 2454S (1983)
- Weed, H. G., McGandy, R. B. and Kennedy, A. R.: Protection against dimethylhydrazine-induced adenomatous tumors of the mouse colon by the dietary addition of an extract soybeans containing the Bowman-Birk soybean protease inhibitor. *Carcino*genesis, 6, 1239 (1985)
- Shin, S. H., Jhee, E. C., Rapp, N. S., Hong, I. S., Chang, S. H. and Seel, D. J.: Mutagencity and antimutagenicity of Meju, Hot sauce and the Korean foods by Slmonella/mammalian-microsome test: Abstract The 5th Federation of Asian and Oceanian Biochemists, August, 13-18, Seoul, Korea, p. 301 (1989)
- Maron, D. M. and Ames, B. N.: Revised methods for the Salmonella mutagenicity test. Mutat. Res., 113, 1783 (1983)
- Matsushima, T., Sugimura, T., Nagao, M., Yahai, T., Shirai, A. and Sawamura, M.: Factors madulating mutagenicity in microbial test. In "Short-term test systems for detecting carcinogens" Norpoth, K. H. and Carner, R. G. (eds.), Springer, Berling, p. 273 (1980)
- Ames, B. N., McGann, J. and Yamasaki, E.: Method for detecting carcinogens and mutagens with the Salmonella/mammalianmicrosome mutagenicity test. Mutat. Res., 31, 347 (1975)
- Qukillardet, P. and Hofnung, M.: The SOS chromotest, a colorimetric bacterial assay for gentoxins. *Mutat. Res.*, 147, 65 (1985)
- Baik, C. W. and Ham, S. S.: Antimutagenic effects of browning products reacted with polypherol oxidase extracted from apple by using SOS chromotest. *Korean J. Food Sci. Technol.*, 22, 618 (1990)
- Miller, J. H.: Experiments in Molecular Genetics, Cold Spring Harbor Laboratory. CSH, New York (1985)
- Steel, R. G. D. and Torrie, J. H.: Principles and procedures of statistics. McGraw-Hill Kogakusha, Ltd., Tokyo, p. 187 (1980)
- Park, K. Y., Moon, S. H., Baik, H. S. and Cheigh, H. S.: Antimutagenic effect of doenjang (Korean fermented soy paste) toward aflatoxin. J. Korean Soc. Food Nutr., 19, 156 (1990)
- Park, K. Y., Moon, S. H. and Rhee, S. H.: Antimutagenic effect of Doenjang (Korean soy paste) - Inhibitory effect of doenjang stew and soup on the mutagenicity induced by aflatoxin B₁. Environ. Mut. Carcino., 14, 145 (1994)
- Park, K. Y. and Rhee, S. H.: The stability against aflatoxins, and antimutagenic and anticancer effects of doenjang. Symposium on status and development of quality of Korean traditional foods. Korean Soc. Food Sci. Tech., p. 109 (1995)
- Park, K. Y.: Destruction of aflatoxin during the manufacture of doenjang (Korean soy paste) by traditional method, and anticancer activities of the doenjang. The 1st International Symposium Functional and Physiological Activities of Korean Traditional Soybean Fermented Foods. p. 37 (1996)
- Park, K. Y.: The stability and anticancer effect of traditionally fermented doenjang. Proceeding in Korean traditional soybean fermented foods. Oct. 15, p. 37 (1996)
- Park, K. Y., Moon, S. H., Cheigh, H. S. and Baik, H. S.: Antimutagenic effects of doenjang (Korean soy paste). J. Food Sci. Nutr., 1, 151 (1996)
- 31. Park, K. Y.: The safety and antitumor activity of doenjang. *Rural Life Science*, 17, 36 (1996)
- Park, K. Y., Lim, S. Y. and Rhee, S. H.: Antimutagenic and anticarcinogenic effects of doenjang. J. Korean Assoc. Cancer Prevention, 1, 99 (1997)
- Park, K. Y.: Antimutagenic and anticancer functions of Korean traditional fermented foods (doenjang, kimchi). Food Sci. Indust., 30, 89 (1997)
- 34. Park, K. Y.: Destruction of aflatoxins during the manufacture of doenjang by traditional method and cancer preventive effects of doenjang. J. Korean Assoc. Cancer Prevention, 2, 27 (1997)
- Jung, K. S., Yoon, K. D., Hong, S. S. and Kwon, D. J.: Antimutagenic and anticancer effect of Korean fermented soybean products. J. Food Sci. & Technol., 1, 75 (1996)

- 36. Takahashi, M. and Hasegawa, R.: In "Nutrition and cancer" Hayashi, Y., Nagao, M., Sugimura, T., Takayama, S., Tomatis, L., Wattenberg, L. W. and Wogan, G. N. (eds.), Japan Sci. Soc. Press, Tokyo: VNU Sci. Press, UTrecht, p. 169 (1986)
- Park, K. Y., Kim, S. H. and Suh, M. J.: Comutagenicity of high salted kimchi in the Samonella assay system. J. Colle. Home Eco., Pusan Nat'l Univ., 16, 45 (1990)

(Received January 25, 1999)