

**F314 Characterization of Poliovirus isolated from patients with aseptic meningitis.**

Ki Soon Kim, Yoon Seok Chung<sup>†</sup>, Yoon Sung Lee<sup>†</sup>, Young Sun Kim, Hong Rae Lee, Moon Bo Kim<sup>†</sup>, Sang Hyuk Ma<sup>\*</sup>, Kwang Ho Lee<sup>†</sup>, Chul Yong Song<sup>†</sup> and Jae Deuk Yoon

Laboratory of Enteroviruses, Department of Virology, NIH

<sup>†</sup> Department of Biology, Faculty of Natural Science, Chung-Ang University

<sup>\*</sup> Department of Pediatrics, Fatima Hospital, Masan.

Poliovirus, a member of picornaviridae, major causative agent of paralytic poliomyelitis was isolated from feces of patients with aseptic meningitis in 1997. The virus was identified by cell culture methods and typed with WHO/RIVM enterovirus typing serum pool. RNA was prepared from characterized virus and reverse transcription was done using a commercially available random hexamer. After cDNA synthesis, a part of VP1 regions were amplified by PCR with UG1, UC1 primer set. Amplified viral cDNA was then confirmed by 0.8% AGE and specific fragment of 481 bp was obtained. PCR products were examined for restriction fragment length polymorphism(RFLP) with Hae III and confirmed as Sabin type 3. To make sure that this virus was originated from vaccine strain, RCT(reproductive capacity at different temperature) test was performed. The virus didn't show any plaque in case of elevated temperature. Sequence analysis of VP1 region and 5' noncoding regions of the virus has shown that there was over 99% homogeneity between reference virus and isolate. Our findings suggest that there may be a circulating polio viruses which were originated from vaccine and polio virus can induce aseptic meningitis, even in case of vaccine derived polio viruses.

**F315 Counterbalances on CuZnSOD and Cat G conjugated Gene expression in Escherichia coli Double Mutants to Oxidative Stress**

Y. G. KIM,<sup>1</sup> H. Y. PARK<sup>1</sup> and H. M. HASSAN<sup>2</sup>

*Department of Biological Science, College of Natural Sciences, Chosun University, Kwangju 501-759, Korea<sup>1</sup>, and Department of Microbiology, North Carolina State University, Raleigh, NC 27695-7622, USA<sup>2</sup>*

In *Escherichia coli*, the counterbalances between superoxide dismutase (SOD) and catalase are more important for overall sensitivity to oxidative stress such as paraquat(PQ, 0.1mM), H<sub>2</sub>O<sub>2</sub> (1mM), CuSO<sub>4</sub> (0.1mM) and heat shock (37°C→42°C) than the level of either SOD or HPI alone. To evaluate relative coordinate defense, *E. coli* SOD and/or hydroperoxidase double mutants were conducted by the transformation of plasmid oriented in wild type, SOD<sup>+</sup>HPI<sup>-</sup>, SOD<sup>-</sup>HPI<sup>+</sup> and SOD<sup>-</sup>HPI<sup>-</sup> mutant cells, and their physiological response to oxidative stress was measured. The toxicity observed increasing expression of SOD than wild type by CuSO<sub>4</sub> or heat shock treatment results in more sensitivity to oxidative stress. This might be a result of an increase in hydrogen peroxide. The same pattern was found in the H<sub>2</sub>O<sub>2</sub> treatment. However, SOD was found to be more important than hydroperoxide in preventing oxygen-mediated stress except in the case of H<sub>2</sub>O<sub>2</sub> treatment in the presence of SOD. Nevertheless, both enzymes were necessary for an effective defense against oxygen mediated radicals. Moreover, although HPI could reduce the H<sub>2</sub>O<sub>2</sub> or hydroxyl radicals, it also reduced the SOD function against oxygen radicals. This system offers much greater analysis on triple mutant including glutathione peroxidase.