

Fructan Synthesis in Transgenic Tomato Plants Expressing Jerusalem Artichoke(*Helianthus tuberosus* L.) 1-*sst* and 1-*fft*

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Objectives

To study the regulation of fructan synthesis in plants, we isolated two full size cDNA clones encoding the two enzymes responsible for fructan biosynthesis in Jerusalem Artichoke (*Helianthus tuberosus* L.) 1-sucrose:sucrose fructosyl transferase (1-*sst*) and 1-fructan:fructan fructosyl transferase (1-*fft*), transformed to tomato and investigated fructan synthesis in transgenic tomato.

Materials and Methods

1. Materials

Jerusalem Artichoke (*Helianthus tuberosus* L.) for gene cloning

Tomato (*Lycopersicon esculentum* M cv Chomyoung) for transformation

2. Methods

RT-PCR, Sequence analysis, Construction of 35S-*sst* and 35S-*fft* chimeric genes

Agrobacterium-mediated transformation, Southern blot and Northern blot.

Results and Discussion

cDNA clones (1-*sst*, 1-*fft*) encoding the two enzymes responsible for fructan biosynthesis were isolated from Jerusalem Artichoke (*Helianthus tuberosus* L.) tuber. The deduced amino acid sequence of both cDNAs perfectly matched the sequences of the corresponding purified proteins (Ingrid et al(1998) *The Plant Journal*, 15(4)489-500). Using the 1-*sst* and 1-*fft* cDNAs, chimeric genes were constructed driven by the CaMV 35S promoter. Analysis of transgenic tomato plants carrying these constructs showed that both cDNAs encoding functional fructosyltransferase enzymes. Plants transformed with the 35S-1-*sst* construct accumulated the oligofructans 1-kestose, 1,1-nystose and 1,1,1-fructosyl-nystose. However plants transformed with the 35S-1-*fft* construct did not accumulate fructans. TLC analysis shown that transgenic tomato containing 35S-1-*sst* accumulated with high molecular weight fructans in old, senescent leaves.