Characterization of *Bacillus* species occurring anaerobic denitrification in night soil treatment

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Abstract

Bacillus species predominantly outgrown in a night soil treatment system were isolated and characterized. Cell interactions took place among them and cell population changed under various culture conditions. Maximum removal of NH₄⁺-N and cell production occurred under the conditions of 30% DO and C/N ratio of 8. Additions of 0.8% peptone and 0.3% yeast extract to a basal medium influenced the growth of isolates and the removal of NH₄⁺-N in flask culture, and metal ions such as Ca, Fe and Mg also did. During the flask experiment of nitrogen removal under an aerobic condition, active nitrification by the isolates occurred largely in 1 h with the decrease of COD and alkalinity destructed was only 74.6% of theoretical value. From the nitrogen balance, the percentage of nitrogen lost in the flask culture was estimated to be 29.0%. This conversion of ammonia to N₂ under an aerobic condition was confirmed by GC analysis. The B3 process using the Bacillus species seemed to have some economic advantage.

Introduction

Nitrogen removal is an important aspect of present day wastewater treatment processes, and biological nitrification-denitrification is one of the most economical processes for nitrogen removal from municipal wastewaters. It has commonly been accepted that denitrification requires anoxic conditions, but there have been periodic reports of aerobic denitrification¹⁾. The nitrification- denitrification process has been also challenged by a one-step process in which ammonium is oxidized directly to $N_2^{(2)}$. Recently, researches focus on nitrite nitrification, which might be a short cut process for savings in oxygen for nitrification and carbon requirements for denitrification³⁾.

In Korea, B3 (Bio Best Bacillus) process (Korean Patent No. 151928) has been known as an advanced wastewater treatment system in which *Bacillus* species predominantly

outgrown. Bacillus species has known to occur heterotrophic nitrification, but other characteristics of the Bacillus species have not studied in detail so far. Understanding characteristics of microorganisms used in the biological nitrification- denitrification process is very important in order to remain high efficiency of treatment at all times, and the possible microbiological nitrogen conversions should be provided. The purpose of this study was to understand Bacillus species predominantly outgrown in a night soil treatment system.

Materials and Methods

Isolation of Bacillus species

Bacillus sludge were obtained from night soil treatment systems. The samples were first agitated to obtain homogeneous suspensions in sterile 0.2% NaCl. 1 ml of the suspended liquid was pipetted into a 10 ml tube that contained a 0.8% nutrient broth medium. After 2 days of incubation at 30°C and 150 rpm, colonies were formed separately on 1.5% nutrient agar plates.

Characterization of isolates

To observe the change of cell population among *Bacillus* species under various culture conditions, a 5 l- bioreactor was used. The composition of a medium used in this experiment was (g/l): glucose, 1.05; NH₄Cl, 0.382; KH₂PO₄, 0.131; peptone, 0.05; yeast extract, 0.05; and 1 ml- mineral solution. In flask experiments to study effects of growth factors on growth of isolates, a basal synthetic medium was used, which contained (g/l): KNO₃, 1; K_2HPO_4 , 1; EDTA, 3.5×10^{-3} ; $ZnSO_4\cdot7H_2O$, 2×10^{-3} ; $FeSO_4\cdot7H_2O$, 10×10^{-3} ; $MnSO_4\cdot7H_2O$, 2×10^{-3} ; $CuSO_4\cdot5H_2O$, 1×10^{-3} ; $Co(NO_3)_2\cdot6H_2O$, 0.2×10^{-3} ; and H_3BO_3 , 1×10^{-3} .

Capacity of isolates for aerobic denitrification was tested in a tightly sealed 11- branched flask. Pure oxygen was pressurized into the flask, and a septum was equipped on the mouth of the flask for gas analysis. The composition of the substrate was (mg/l): COD, 250; NH₄⁺-N, 35; NO₂⁻-N, 0; NO₃⁻-N, 0; TN, 50; and TP, 7. 10% of active isolated *Bacillus* species were inoculated into the flask. The flask was incubated at 150 rpm and 30℃.

Analyses

Growth of isolates was determined spectrophotometrically at 510 nm. The concentrations of NH₄⁺-N, NO₂⁻-N, NO₃⁻-N COD, TN, TP, alkalinity of medium were determined by procedures given in the Standard Methods. O₂ and N₂ gases were determined by GC/TCD analysis.

Results and discussion

Twenty-four species could be isolated. PK15 grew very well under aerobic conditions (more than 15% DO) and PK5 and PK16 did under the condition of 5% DO. Maximum removal of NH₄⁺-N and cell production occurred under the conditions of 30% DO and C/N ratio of 8 (Fig. 1). When DO concentration maintained in a range of 15-30%, DO did not affect the removal efficiency of NH₄⁺-N significantly, but the efficiency almost halved under the condition of 5% DO. The removal efficiency of NH₄⁺-N by the isolates was higher at a C/N ratio of 8 than that at C/N ratio of 4 (Fig. 2). At a C/N ratio of 8, there was not any effect of the substrate composition on the removal efficiency of NH₄⁺-N, but the effect of substrate composition was significant at the C/N ratio of 4.

The concentration of NH₄⁺-N decreased significantly in 1 h, and that of TN also did (Fig. 3). The same trend could be seen in both the removal of COD and the destruction of alkalinity. After then, the concentrations of NH₄⁺-N, TN, COD and alkalinity did not decrease significantly. The removal efficiencies of NH₄⁺-N, TN, COD and TP for 4 h were found to be 72.0, 53.8, 89.5 and 21.4%, respectively. When the concentration of organic nitrogen decreased, cell production almost maintained and the substantial removals of COD and NH₄⁺-N did not occur further.

From the nitrogen balance (Table 1), 14.5 mg/l (29% of initial TN) of nitrogen in the medium disappeared after 4h. From the above results, the mixed *Bacillus* cells were found to be able to occur aerobic denitrification indeed. This fact has not been reported yet.

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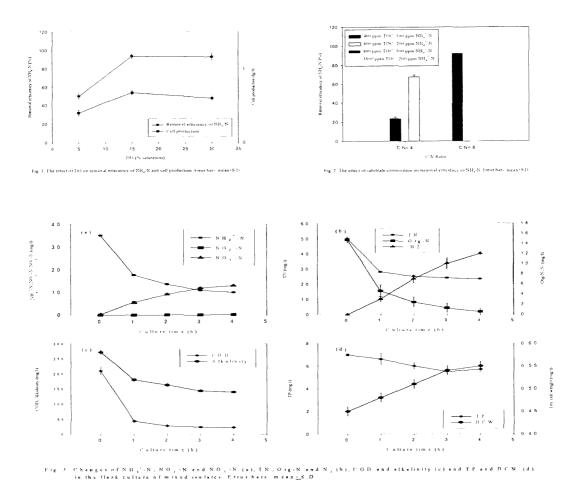


Table 1. Nitrogen balance for the flask culture (units: mg/l)

Initial TN	Final	Final	Final	Final	N in	% N lost ³
	NH₄⁺-N	NO ₂ -N	NO ₃ -N	Org-N ¹	Biomass ²	
50.0	9.8	0	12.8	0.5	12.4	29.0

Calculated value. ²Biomass composition was assumed to be C₅H₇O₂N.

 3 % N lost = $100 \times \{(initial\ TN)-(final\ NH_{4}^{+}-N)-(final\ NO_{2}^{-}-N)-(final\ NO_{3}^{-}-N)-(final\ Org-N) -(N in biomass)\}/$ (initial TN)