Glycoengineering of Yeast Cells for Humanized N-glycans

강현아

한국생명공학연구원 대사공학실

TEL: +82-42-869-4378, FAX: +82-42-860-5495, E-mail: hyunkang@kribb.re.kr

Due to its capacity to glycosylate proteins, yeasts have been considered for the production of glycoproteins derived from higher eukaryotes. However, yeast N-glycosylation pathway is only partially homologous to the pathway in human cells. Here we report the remodeling of glycosylation pathway for biosynthesis of humanized N-glycans in the methylotrophic yeast Hansenula polymorpham, which is rapidly gaining favor as a promising host for the production of recombinant proteins (1). Especially, H. polymorpha shows some advantages over the traditional yeast Saccharomyces cerevisiae in the production of recombinant glycoproteins for human therapeutic use. Most N-linked oligosaccharide species attached to H. polymorpha-derived glycoproteins have core-sized structures (Man₈₋₁₂GlcNAc₂) without highly immunogenic terminal a-1,3-linked mannose residues. Moreover, the outer chains of H. polymorpha N-linked glycans were shown to be extended mostly in 1,2-mannose linkages and to have very short 1,6 extensions, mainly composed of single a-1,6-linked mannose (2). As a first step toward humanizing H. polymorpha N-glycosylation pathway, the H. polymorpha och2 mutant strain having a defect in the outer chain initiation on the core oligosaccharide Man₈GlcNAc₂ was developed and further engineered with the targeted expression of Aspergillus saitoi a -1,2-mannosidase in the ER. The engineered H. polymorpha och2 strain was shown to produce the human high mannose-type Man₅GlcNAc₂ oligosaccharide as a major N-glycan (3). As an alternative approach, the remodeling of core oligosaccharide assembly pathway was carried out with additional deletion of theH. polymorpha ALG3 gene (HpALG3), encoding Dol-P-Man:-Man₅GlcNAc₂-PP-Dol mannosyltransferase. The engineered double deletion (alg3och2) mutant strain expressing A. saitoi a-1,2-mannosidase generated mainly the trimannosyl-core form glycan (Man3GlcNAc2), which is an intermediate for further maturation to human-like complex *N*-glycans. These results demonstrate the potential of *H. poly-morpha* to be developed as a host for the production of therapeutic glycoproteins with homogeneous complex *N*-glycan structures.

References

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