

Studies on the Ginseng Plants(III)* Radioactive Sodium Acetate-U-C¹⁴ Feeding Experiments

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人蔘植物에 관한 연구(Ⅲ)*

同位元素化合物 Sodium Acetate-U-C¹⁴을 투여한 실험

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The radioactive compound sodium acetate-U-C¹⁴ (C¹⁴-acetate) was administered to two- and four-year-old July and September American ginseng (*Araliaceae*, *Panax quinquefolium* L.) plants and cuttings. The C¹⁴-acetate uptake was approximately 99 %. The autoradiochromatograms suggest that the saponins isolated by preparative thin-layer chromatography contained impurities, especially those isolated from the leaf and stem extracts. The root and fruit methanol extracts yielded relatively pure saponins. The large amounts of panaquilin B and its proximity to panaquilin C on preparative thin-layer plates resulted in some admixing. The average concentration (% plant dry weight) of semi-purified saponins were high in the leaves (13.8 %), as compared to fruits (9.8 %), stems (7.9 %) and roots (6.3 %). The average percentage of C¹⁴-acetate incorporation into panaquilins was 4.8 %. The average percentage of C¹⁴-acetate incorporation into panaquilins B and C was higher (1.40 % and 1.13 %, respectively) than that into panaquilins C, (d), G-1 and G-2 (0.75 %, 0.65 %, 0.13 % and 0.53 %, respectively). Panaquilin synthesis may be depending upon the part, collection period and age of the plant. The average percentage of C¹⁴-acetate incorporation into panaquilin B is high in roots (0.58 %) and stems (0.48 %); that into panaquilins C and (d) high in leaves (0.40 % and 0.45 %, respectively); and that into panaquilin E high in roots and leaves (0.55 % and 0.50 %, respectively). Panaquilin G-2 was synthesized in all parts of plants. The panaquilins appear to be biosynthesized more actively in July than September (exception-panaquilin G-1). Panaquilins B, C and G-1 may be biosynthesized more actively in four-year-old plants and panaquilins (d) and E more actively in two-year-old plants. The results from expectance with cuttings suggest that the panaquilins are synthesized *de novo* in the above-ground parts of ginseng plants, and that panaquilin G-1 may be synthesized *de novo* in the leaf. It is known from the tissue culture studies that panaquilins are produced by leaf, stem and root callus tissues and callus-root cultures of American and Korean ginseng plants. Panaquilins may actively be synthesized *de novo* in most any cell or organ of the ginseng plants. It was verified that C¹⁴-acetate was incorporated into the panaxadiol portions of the panaquilins of two-year-old plants (sp. act. 0.56 m μ Ci/mg) and four-year-old plants (sp. act. 0.54 m μ Ci/mg).

* This manuscript is abstracted in part from a dissertation submitted to the Graduate School of the University of Minnesota in partial fulfilment of the requirements for the degree of Doctor of Philosophy.

Introduction

The two previous reports considered the isolation, identification and distribution patterns of American ginseng saponins and sapogenins¹⁾, and H³-squalene feeding experiments²⁾. In this study, the purpose of the radioisotope experiments, therefore, was to confirm and to quantitate the panaquilin thin-layer chromatographic distribution patterns previously observed in plants not fed radioisotopes, and to obtain C¹⁴-labeled panaquilins for future studies.

Results and Discussion

Radioactive sodium acetate-U-C¹⁴ (C¹⁴-acetate) was fed by the wick method³⁾ to two- and four-year-old plants and cuttings at the beginning and the end of a growing season (July and September). After the feeding, the plants were allowed to grow in the field for 7 days, and the cuttings in a hydroponic solution for 5 days.

Aqueous C¹⁴-acetate (400 m μ Ci/mg) solutions were

fed to two-year-old plants (95 plants studied in July, 2.1 μ Ci/0.5 ml/plant; 100 plants studied in September, 2 m μ Ci/0.5 ml/plant), four-year-old plants (19 plants studied in July, 40 μ Ci/0.5 ml/plant; 20 plants studied in September, 40 μ Ci/0.5 ml/plant), two-year-old stem cuttings (40 cuttings studied in July, 1 μ Ci/0.5 ml/cutting; 41 cuttings studied in September, 1 μ Ci/0.5 ml/cutting) and four-year-old stem cuttings (20 cuttings studied in July, 10 μ Ci/0.5 ml/cutting; 20 cuttings studied in September, 10 μ Ci/0.5 ml/cutting). The approximate fresh weights for two-year-old plants and cuttings were 3 g and 1.5 g, respectively; and for four-year-old plants and cuttings were 80 g and 25 g, respectively. The uptake of C¹⁴-acetate solutions by two- and four-year-old plants was approximately 98.7 %, and that for two-year-old cuttings was approximately 98.7 %, and that for two- and four-year-old cuttings approximately 99.9 %.

The amounts of extracts obtained from the plants and cuttings (Tables I and II) are similar to those obtained from non-radioactive experiments¹⁾. The

Table I. Extracts from Two-year-old Ginseng Plants and Cuttings: Sodium Acetate-U-C¹⁴ Experiments*.

	Leaf		Stem		Root		Average	
	Jl	Sp	Jl	Sp	Jl	Sp	Jl	Sp
Plants								
Dry Wt. (g)	40.0	29.1	12.1	12.1	9.8	90.8	129.0	56.0
<i>Extract (%)**</i>								
Ether	13.0	5.5	1.1	4.6	1.4	1.3	5.2	3.8
Chloroform	1.8	2.7	0.2	0.7	0.4	1.7	0.8	1.7
Methanol-1	51.5	43.0	21.4	23.3	18.8	21.6	30.6	29.3
Residue	6.4	0.7	3.5	6.9	1.9	2.8	3.9	3.5
Methanol-2	40.9	42.3	17.8	16.4	16.9	18.8	25.2	35.8
Total	66.3	51.2	22.7	28.6	20.6	24.6	36.6	34.8
Cuttings								
Dry Wt. (g)	16.5	20.6	4.5	7.8	NA	NA	10.5	14.2
<i>Extract (%)**</i>								
Ether	1.8	3.7	0.9	4.4	NA	NA	1.4	4.1
Chloroform	2.6	2.0	0.4	1.4	NA	NA	1.5	1.7
Methanol-1	39.3	54.5	13.8	25.4	NA	NA	28.1	40.0
Residue	5.2	NA	1.1	8.5	NA	NA	3.2	4.3
Methanol-2	25.8	48.3	12.7	16.8	NA	NA	19.3	32.3
Total	43.7	60.2	15.1	31.2	NA	NA	31.0	45.8

* Abbreviations: Jl-July collection; Sp-September collection; NA-not available.

** Residue: Insoluble material of methanol-1 extracted with cold methanol (5° C); Methanol-2: Soluble extracts of methanol-1 extracted with cold methanol (5° C); Total=Ether (%) + Chloroform (%) + Methanol-1 (%).

Table II. Extracts from Four-year-old Ginseng Plants and Cuttings: Sodium Acetate-U-C¹⁴ Experiments*.

	Leaf		Stem		Fruit		Root	
	Jl	Sp	Jl	Sp	Jl	Sp	Jl	Sp
Plants								
Dry Wt. (g)	56.5	46.4	43.0	30.7	18.0	36.6	199.0	181.0
<i>Extract (%)**</i>								
Ether	8.9	2.9	2.9	0.8	5.2	15.8	1.5	0.5
Chloroform	2.0	1.7	0.3	0.8	0.7	9.5	0.9	0.4
Methanol-1	44.7	48.3	24.4	24.3	37.0	22.2	19.9	52.4
Residue	10.5	1.7	0.7	2.2	1.8	4.8	2.5	4.9
Methanol-2	27.4	28.3	23.7	22.1	35.3	17.3	17.3	47.5
Total	55.6	52.9	27.6	25.9	42.9	47.5	22.3	53.3
Cuttings								
Dry Wt. (g)	45.0	49.4	32.2	32.6	9.4	39.1	NA	NA
<i>Extract (%)**</i>								
Ether	3.3	5.0	1.2	1.3	3.7	17.2	NA	NA
Chloroform	3.9	1.7	1.0	1.0	1.3	2.0	NA	NA
Methanol-1	38.8	43.9	23.6	25.6	39.5	20.2	NA	NA
Residue	4.6	0.1	0.4	1.9	2.3	5.1	NA	NA
Methanol-2	30.4	38.8	23.2	23.7	37.2	15.1	NA	NA
Total	46.0	50.6	25.8	27.9	44.5	39.4	NA	NA

* Abbreviations: Jl-July collection; Sp-September collection, NA-not available.

** Residue: Insoluble material of methanol-1 extracted with cold methanol (5° C); Methanol-2: Soluble extracts of methanol-1 extracted with cold methanol (5° C); Total=Ether (%) + Chloroform (%) + Methanol-1 (%).

thin-layer procedures used for isolating the saponins from radioactive extracts were the same as those used for non-radioactive extracts¹⁾.

1. Extracts

The percentages of ether extracts were 1.5~8%; those of chloroform, 0.8~3.0%; and those of methanol-1, 20~40%. The total percentage of ether, chloroform and methanol-1 extracts were higher in the leaf than in the root (Tables I and II). This result could be anticipated because of the high amounts of pigments and lipids observed in the leaf extracts. The residue, data shown in Tables I and II, contains principally carbohydrates and is separated from methanol-1 by cold methanol (5° C).

The percentage of C¹⁴-acetate incorporation into ether extracts was generally higher than that of chloroform extracts, but was lower than that of methanol extract-1 (Table III). The percentage of C¹⁴-acetate incorporation into ether and chloroform extracts was consistently higher in plants studied in

July than those studied in September.

Methanol-2 saponin extracts were semi-purified in that considerable (2.4~4%) carbohydrate residue was removed (Tables I and II). The percentage of methanol-2 C¹⁴-acetate incorporation is less (average 26.1%) than that in methanol-1 (average 33.1%), and is probably due to its incorporation into carbohydrates (Table III and Fig. 1). The percentage of methanol-2 C¹⁴-acetate incorporation is high (15.4%) in two-year-old plants collected in July (Fig. 1). The average percentage of C¹⁴-acetate incorporation is low in fruits (1.5%) as compared to roots (3.1%), leaves (4.2%) and stems (6.2%). The average percentage of C¹⁴-acetate incorporation was higher in roots studied in September (3.8%) than those studied in July (2.4%). The leaves studied in July had higher average C¹⁴-acetate incorporation than the roots (6.0% and 2.3%, respectively), but the leaves studied in September were lower than the roots (2.4% and 3.8%, respectively).

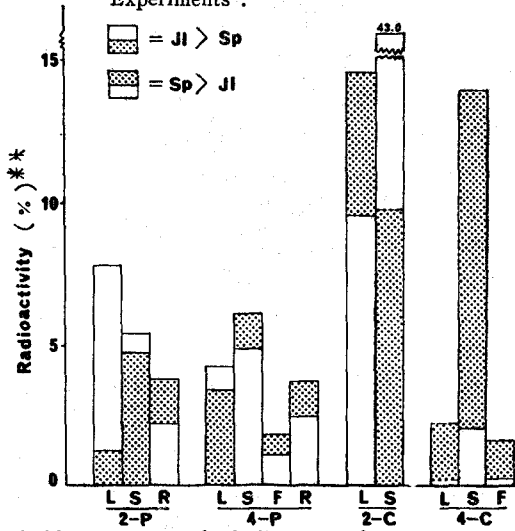
Table III. C¹⁴-Activity (%) of Ether, Chloroform and Methanol Extracts*.

Plant Material**	Two-year-old				Four-year-old			
	Ether	Chloroform	Methanol	Total	Ether	Chloroform	Methanol	Total
Plants								
JIL	0.1	0.3	15.4	15.8	0.02	0.3	9.9	10.2
JIS	2.8	0.4	8.2	11.4	0.9	0.3	5.2	6.4
JIF	NA	NA	NA	NA	0.3	0.1	2.0	2.4
JIR	0.6	0.2	3.6	4.4	0.4	0.1	3.0	3.5
Total	3.5	0.9	27.2	31.6	1.6	0.8	20.1	22.5
SpL	0.03	0.1	2.1	2.2	0.02	0.1	7.6	7.7
SpS	2.3	1.2	8.4	11.9	1.0	0.3	8.3	9.6
SpF	NA	NA	NA	NA	0.3	0.1	1.6	2.0
SpR	0.6	0.2	4.3	5.1	0.5	0.1	3.8	4.4
Total	2.9	1.5	14.8	19.2	1.8	0.6	21.3	23.4
Cuttings								
JIL	0.7	0.8	23.8	25.3	NA	0.01	0.4	0.4
JIS	13.6	6.0	61.5	81.1	1.3	0.5	2.4	4.2
JIF	NA	NA	NA	NA	0.1	0.02	0.3	0.4
Total	14.3	6.8	85.3	106.4	1.4	0.5	3.1	5.0
SpL	0.1	0.3	14.8	15.2	0.01	0.1	3.6	3.7
SpS	3.5	1.2	16.6	21.3	2.1	0.5	16.6	19.2
SpF	NA	NA	NA	NA	0.3	0.1	1.6	2.0
Total	3.6	1.5	31.4	36.5	2.4	0.7	21.8	23.9

* C¹⁴-Activity (%): (Total radioactivity in extract/Total radioactivity uptake)x100.

** Abbreviations: JI-July collection; Sp-September collection; L-leaf; S-stem; F-fruit; R-root; NA-not available.

Fig. 1. Radioactivity of Methanol-2 Extracts: C¹⁴-Experiments*.



*Abbreviations: L-leaf; S-stem; F-fruit; R-root; 2-P: Two-year-old plants; 4-P: Four-year-old plants; C: Cuttings; JI-July collection; Sp-September collection.

**Radioactivity (%): (Total radioactivity in extract/Total radioactivity uptake) × 100.

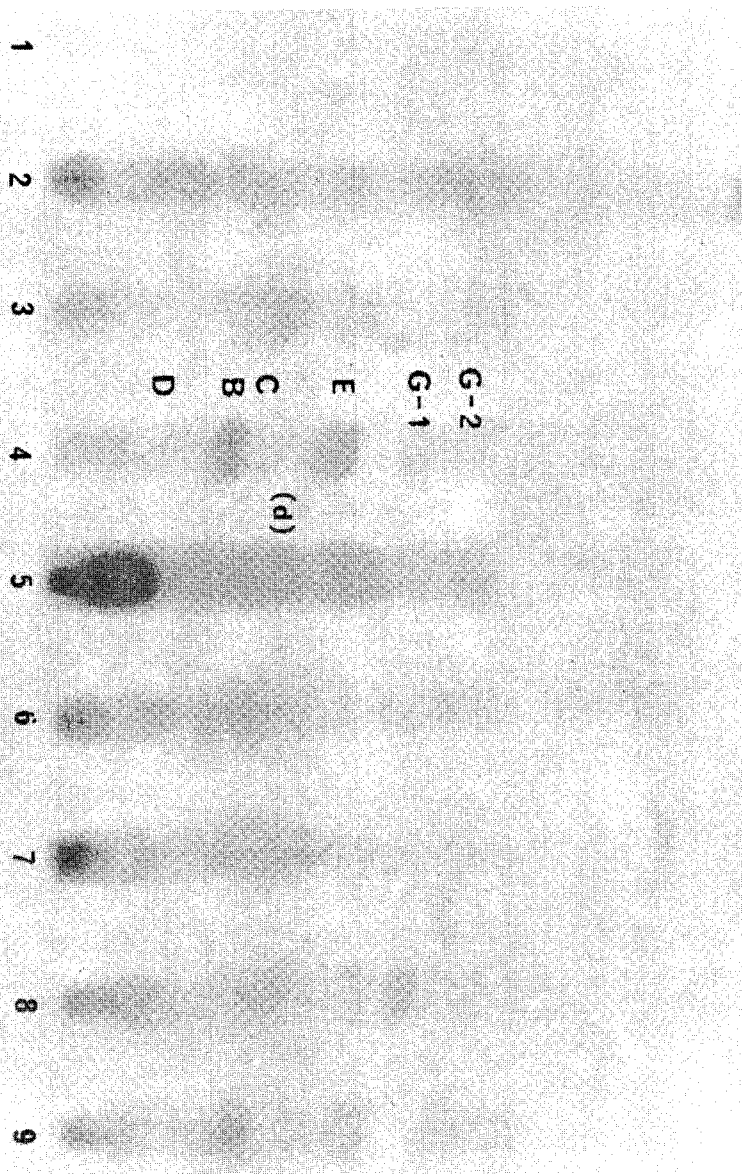
When extract activity was expressed as μCi per mg dry weight it was consistently higher in stems (7.4 $\mu\text{Ci}/\text{mg}$) and fruits (1.03 $\mu\text{Ci}/\text{mg}$) than in leaves (0.3 $\mu\text{Ci}/\text{mg}$) and roots (0.38 $\mu\text{Ci}/\text{mg}$). This is probably related to the fact that the stem wick method was used to administer the radioactive compound.

In cuttings, the percentage of C¹⁴-acetate incorporation into the methanol-2 leaf extracts of two-year-old cuttings was higher in September (14.6 %) than July (9.5 %). There were significant amounts of methanol-2 extract in two- and four-year-old July green immature fruits (average 37.8 %) as compared to the September mature fruits (average 16.2 %) (Table II).

2. Autoradiography of Methanol-2

The radioactive methanol-2 solutions (approximately 10 % in methanol, 20~60 lambda) were applied to silica gel plate (0.5 mm thick). The plates were exposed to X-ray films (Kodak Rp Royal X-Omat) for

Plate 1. One-dimensional Tlc Autoradiochromatograms of American Ginseng Methanol-2 Extracts*.

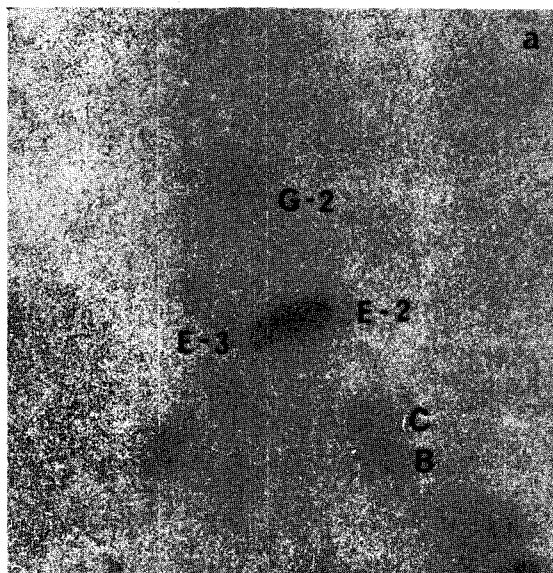


D-panaquillin D; B-panaquillin B; C-panaquillin C; (d)-panaquillin (d); E-panaquillin E; G-1-panaquillin G-1; G-2-panaquillin G-2. **Film:** Kodak RP Royal X-Omat (Eastman Kodak Co., Rochester, N. Y.); **Development:** Kodak X-ray developer; **Exposure time:** 42 days. **Sample:** 1. Four-year-old leaves collected in July (cutting, 20 lambda, 70 cpm); 2. Four-year-old stems collected in July (cutting, 20 lambda, 1,540 cpm); 3. Four-year-old fruits collected in July (cutting, 40 lambda, 520 cpm); 4. Two-year-old roots collected in July (30 lambda, 1,340 cpm); 5. Two-year-old leaves collected in July (cutting, 40 lambda, 2,980 cpm); 6. Four-year-old leaves collected in September (cutting, 20 lambda, 800 cpm); 7. Four-year-old fruits collected in September (cutting, 40 lambda, 1,340 cpm); 8. Two-year-old leaves collected in September (20 lambda, 760 cpm); 9. Two-year-old roots collected in September (40 lambda, 1,980 cpm).

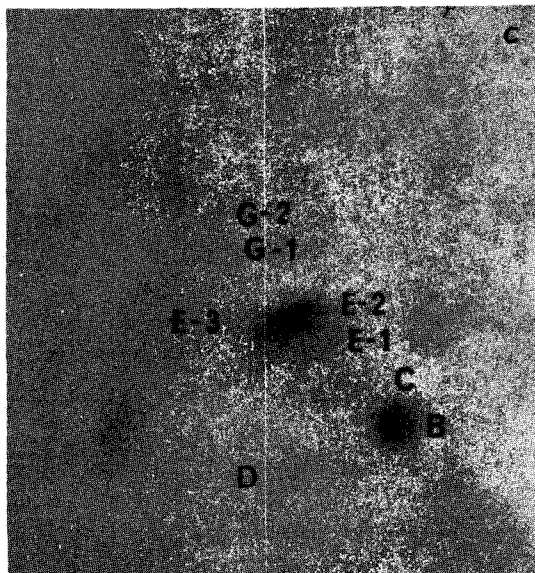
Silica gel plate: 20x20cm; **Thickness:** 0.5 mm; **Solvent system:** Chloroform: Methanol: Distilled water=65:35:10

* The background of original autoradiochromatograms negative was darker than that of this Plate, and panaquillins (d), G-1 and G-2 were visible in the original, but are not visible in the Plate.

Plate 2. Two-dimensional TLC Autoradiochromatograms of American Ginseng Methanol-2 Extracts from Four-year-old Plants Collected in July*.

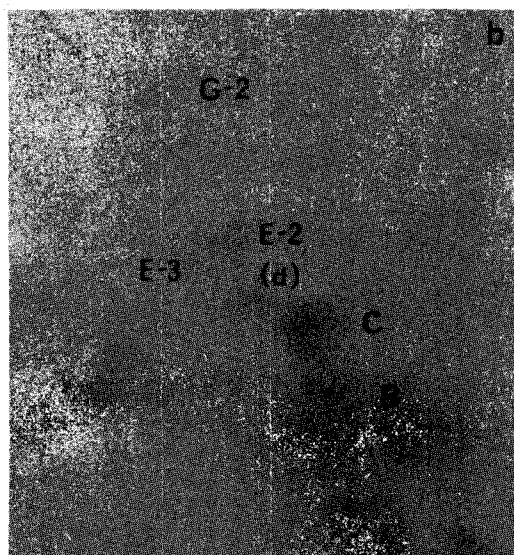


a. Leaf extracts; 20 lambda (6,120 cpm), 14 days exposure. Panaquilins B, C, E-2, E-3 and G-2.



c. Root extracts; 20 lambda (1,870 cpm), 28 days exposure. Panaquilins B, C, D, E-1, E-2, E-3, G-1 and G-2.

* The original background of autoradiochromatograms negative was darker than that of this Plate, and panaquilins (d), G-1 and G-2 were visible, but are not visible in the Plate.



b. Fruit extracts; 20 lambda (5,300 cpm), 14 days exposure. Panaquilins B, C, (d), E-2, E-3 and G-2.

1~6 weeks depending on the activity (100~37,000 cpm) of the aliquot applied. Radioactive saponin patterns (Plates 1 and 2) were similar to non-radioactive saponin patterns verifying C^{14} -acetate incorporation into saponins.

3. Total Radioactivity of Semi-purified Saponins

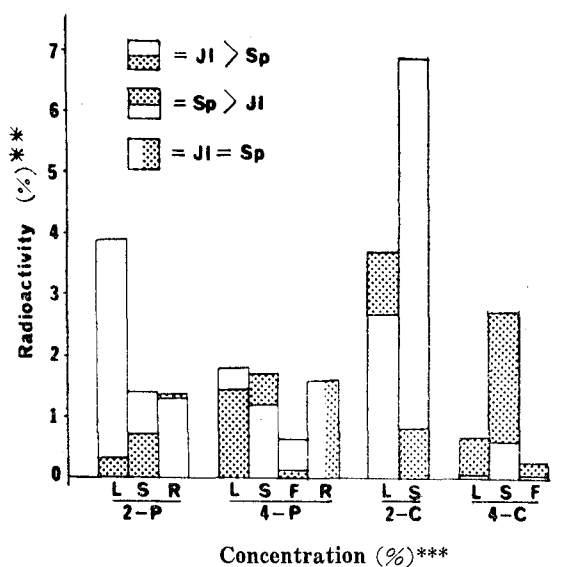
Semi-purified saponins were obtained from silica gel plates by eluting their zone with methanol. The semi-purified saponin concentration showed in Fig. 2 is based on the dry weight total of all the saponin band elutes. The semi-purified saponins did not contain visible silica gel residue. The five bands eluted and studied were panaquilin B (rf-value, 0.23), panaquilin C (Rf-value, 0.30), panaquilin E (Rf-value, 0.41), panaquilin G-1 (Rf-value, 0.55) and panaquilin G-2 (Rf-value, 0.63)¹⁾. The large amount of panaquilin B and its rf-

proximity to panaquilin C resulted in some admixing. The autoradiochromatography also suggests that saponin bands may have contained impurities, especially those isolated from the leaf and stem extracts (Plate 2). The root and fruit methanol extracts contained relatively pure saponins.

a. Plants

The total percentage of C¹⁴-acetate incorporation into the semi-purified saponins (Fig. 2) of two-year-old plants collected in July (6.6 %) was higher than that incorporated into September collected plants (2.4 %),

Fig. 2. Semi-purified Saponin C¹⁴-Radioactivity Distribution and Concentration (%) in American Ginseng Plants and Cuttings*.



	Concentration (%)***							
	2-P			4-P				
	L	S	R	L	S	F	R	
Jl	15.1	6.7	10.1	9.8	4.5	13.1	5.4	
Sp	16.0	5.2	4.2	14.3	15.2	6.5	5.5	
	2-C		4-C					
	L	S	L	S	F			
Jl	11.8	3.4	21.4	9.2	19.4			
Sp	15.7	4.8	16.8	5.8	6.4			

* Abbreviations: L-leaf; S-stem; F-fruit; R-root; Jl-July collection; Sp-September collection; 2-P: Two-year-old plants; 4-P: Four-year-old plants; C-cuttings.

** Radioactivity (%): (Radioactivity in semi-purified saponins (μCi)/Radioactivity uptake (μCi)) × 100.

*** Concentration (%): (Weight of total saponins(mg) / Weight of plants (mg dry weight)) × 100.

and the percentage incorporated in July (5.4 %) was slightly higher than that in September collected plants (4.8 %). The average percentage of C¹⁴-acetate incorporated into root saponins of July collected plants (1.45 %) was approximately the same as September collected roots (1.50 %). The radioactivity (%) data suggests that the physiological activity of two-year-old leaves with regard to C¹⁴-acetate incorporation into saponins is significantly higher in July than in September, and that the roots maintain a fairly constant physiological activity (Fig. 2). The percentage of C¹⁴-acetate incorporation in two-year-old plants collected in September (2.4 %) was lower than at any other time studied.

The concentration (%) of semi-purified saponins were similarly high (average 13.8 %) in both July and September leaves (Fig. 2). This observation is significant, as it is generally believed that the saponins existed principally or only in the root. There is but one related publication⁴⁰ which states the presence of ginseng sapogenins panaxadiol, panaxatriol and oleanolic acid in the above-ground parts of *Panax ginseng*. The saponin content of semi-purified saponins from the the matured red-ripened September fruits was approximately twice lower (6.5 %) than that from the immature green July fruits (13.1 %).

b. Cuttings

The radioactivity (%) of semi-purified saponins was high and localized in the stems. The unusually high localization of radioactivity in July collected may be related to their fragile and young structure and the fact that the stem wick method of radioisotope administration was employed. The average size of the two-year-old stems was 1~2 mm in diameter and 5~7 cm in stem height. Although the radioactivity (%) was generally lower than that in stem, it was present in both leaf and fruit crude saponin fractions.

4. Radioactivity of Semi-purified Saponins

a. Panaquilin B (Panaxadiol Genin)

(1) Plants

The percentage of C¹⁴-acetate incorporated into panaquilin B by the plant was higher in July than in September, with the exception of two-year-old roots

and four-year-old stems (Fig. 3). Four-year-old plants contained more panaquilin B activity (1.8 %) than two-year-old plants (1.1 %). Panaquilin B activity was low in four-year-old July and September fruits (0.05 %). The average panaquilin B activity was consistently higher in two- and four-year-old roots (0.6 %) than that in leaves (0.3 %).

The average concentration of panaquilin B in July collected roots was higher (2.6 %) than that in September collected roots (1.9 %). The above-ground parts (leaves and stems) contained significant amounts of panaquilin B in two-year-old (average 5.2 %) and four-year-old plants (average 7.7 %). Four-year-old stems collected in September contained the highest percentage of panaquilin B (6.2 %), and perhaps this observation

relates to its lower concentration in mature fruits (1.5 %) and immature fruits (4.2 %).

(2) Cuttings

The total percentage of C¹⁴-acetate incorporation was high in July in two-year-old cuttings (1.8 %), and high in September in four-year-old cuttings (1.2 %). The higher percentage of panaquilin B activity in four-year-old September stems (0.8 %) than that in four-year-old July stem (0.1 %) may be related to fruit maturation.

b. Panaquilin C (Panaxadiol Genin)

(1) Plants

The percentage of C¹⁴-acetate incorporation into panaquilin C was with one exception (four-year-old stems) higher in July than in September (Fig. 4). The average panaquilin C activity in the roots (0.15 %) was generally lower than that of leaves (0.4 %) or stems (0.18 %). Four-year-old roots contained more panaquilin C activity (0.2 %) than two-year-old roots (0.1 %). With the exception of two-year-old September leaves (0.02 %), the leaves contained the highest percentage of panaquilin C activity (average 0.5 %). As with panaquilin B, higher panaquilin C activity in September stems may be related to fruit maturation.

Panaquilin C concentration (%) was highest in two-year-old September leaves (5.6 %), and four-year-old July fruits (5.3 %). The high concentration of panaquilin C present in immature fruits (5.2 %) may be related to its low concentration in the stems (0.9 %).

(2) Cuttings

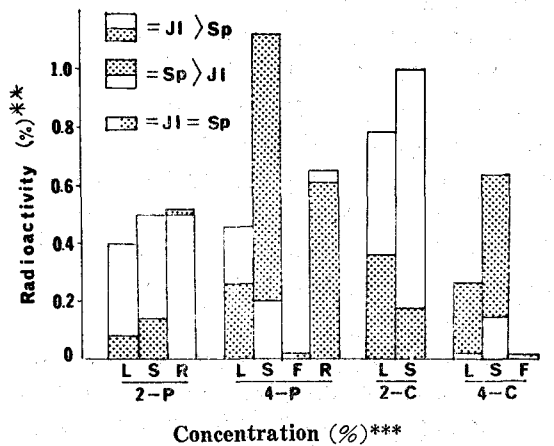
The percentage of C¹⁴-acetate incorporation into panaquilin C in two-year-old cuttings collected in July was higher (2.4 %) than in September (0.5 %). Four-year-old cuttings collected in September contained more panaquilin C activity (1.0 %) than those collected in July (0.1 %). The unusually high four-year-old stem panaquilin C activity may be related to fruit maturation.

c. Panaquilin (d) (Unknown Genin)

(1) Plants

Panaquilin (d) was not present in American ginseng root. This confirms the two-dimensional thin-layer

Fig. 3. Panaquilin B C¹⁴-Radioactivity Distribution and Concentration (%) in American Ginseng Plants and Cuttings*.

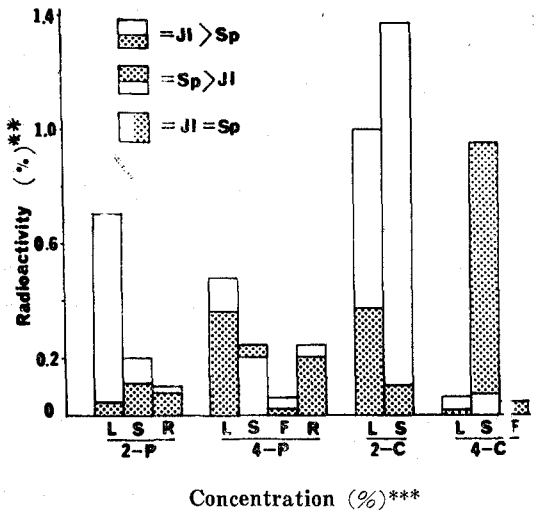


* Abbreviations: L-leaf; S-stem; F-fruit; R-root; Jl-July collection; Sp-September collection; 2-P: Two-year-old plants; 4-P: Four-year-old plants; C-cuttings.

** Radioactivity (%): (Radioactivity in panaquilin B (μCi)/Radioactivity uptake (μCi)) × 100.

*** Concentration (%): (Weight of panaquilin B (mg)/Weight of plants (mg dry weight)) × 100.

Fig. 4. Panaquilin C ¹⁴-Radioactivity Distribution and Concentration (%) in American Ginseng Plants and Cuttings*.



		Concentration (%)***							
		2-P			4-P				
		L	S	R	L	S	F	R	
Jl		4.0	1.3	0.8	3.0	0.9	5.2	0.6	
Sp		5.6	1.3	0.4	3.1	3.8	2.5	1.0	
		2-C			4-C				
		L	S		L	S	F		
Jl		2.9	0.5		6.2	1.9	7.7		
Sp		3.7	1.3		4.1	1.4	2.4		

* Abbreviations: L-leaf; S-stem; F-fruit; R-root; JI-July collection; Sp-September collection; 2-P: two-year-old plants; 4-P: four-year-old plants; C-cuttings.

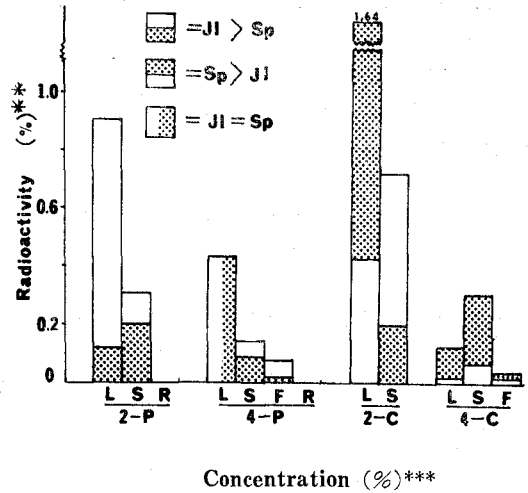
** Radioactivity (%): (Radioactivity in panaquilin C (μ Ci)/Radioactivity uptake (μ Ci)) \times 100.

*** Concentration (%): (Weight of panaquilin C (mg)/Weight of plants (mg dry weight)) \times 100.

chromatography observation made with non-radioactive materials. Also, panaquilin (d) was absent from autoradiochromatograms (Plates 1 and 2).

The average percentage of ¹⁴C-acetate incorporation into panaquilin (d) was highest in two-year-old leaves collected in July (0.9%), and was in general higher in the leaves (0.5%) than the stems (0.2%) and fruits (0.05%) (Fig. 5). The average panaquilin (d) activity was also higher in July collected two- and four-year-old plants (0.6% and 0.2%, respectively) than in September collected plants (0.15% and 0.17%, respectively).

Fig. 5. Panaquilin (d) ¹⁴C-Radioactivity Distribution and Concentration (%) in American Ginseng Plants and Cuttings*.



		Concentration (%)***							
		2-P			4-P				
		L	S	R	L	S	F	R	
Jl		2.9	0.5	—	1.8	0.3	1.2	—	
Sp		2.3	0.4	—	2.4	1.8	1.2	—	
		2-C			4-C				
		L	S		L	S	F		
Jl		2.7	0.3		4.3	1.2	1.9		
Sp		1.5	0.7		1.4	0.8	1.2		

* Abbreviations: L-leaf; S-stem; F-fruit; R-root; 2-P: two-year-old plants; 4-P: four-year-old plants; C-cuttings; JI-July collection; Sp-September collection.

** Radioactivity (%): (Radioactivity in panaquilin (d) (μ Ci)/Radioactivity uptake (μ Ci)) \times 100.

*** Concentration (%): (Weight of panaquilin (d) (mg)/Weight of plants (mg dry weight)) \times 100.

Panaquilin (d) concentration (%) was higher in the leaves of the plants (2.4%) as compared to its stems (0.8%) or fruits (2.2%). The concentration of panaquilin (d) in July immature fruits was the same (1.2%) as that in September mature fruits.

(2) Cuttings

The average percentage of ¹⁴C-acetate incorporation into panaquilin (d) was higher in September (0.44%) than in July (0.23%), and in two-year-old cuttings (0.73%) than in four-year-old cuttings (0.09%). The average concentration of panaquilin (d) was consistently higher in the leaves (2.5%) than in the stems

(0.75 %) or fruits (1.6 %). The average concentration of panaquilin (d) in immature fruits (1.9 %) is slightly higher than that in September fruits (1.2 %).

d. Panaquilin E (Panaxadiol and Panaxatriol Genins)

(1) Plants

The average percentage of C¹⁴-acetate incorporation into panaquilin E in plants collected in July was higher (1.9 %) than those collected in September (0.8 %) (Fig. 6). With the exception of two-year-old July collected leaves (1.5 %), the panaquilin E activity

in the above-ground parts was consistently lower than that in the roots (0.55 %). The panaquilin E activity in July collected roots was slightly lower (0.5 %) than that in September collected roots (0.6 %).

The average concentration of panaquilin E in the roots collected in July was higher (1.9 %) than that in September (1.7 %), and four-year-old roots (2.0 %). The leaf (average 3.2 %) consistently contained more panaquilin E than the root (average 1.8 %). In four-year-old plants, July collected stems (0.6 %) and September mature fruits (0.8 %) contained less panaquilin E than the green immature fruits (1.6 %).

(2) Cuttings

The average percentage of C¹⁴-acetate incorporation into panaquilin E in two-year-old cuttings collected in July was higher (1.0 %) than that in September cuttings (0.4 %). The average concentration (%) of panaquilin E was high in four-year-old cuttings (2.4 %) as compared to that in two-year-old cuttings (1.2 %). The concentration of panaquilin E in July-collected green immature fruits was higher (2.2 %) than that in September collected mature fruits (0.8 %).

e. Panaquilin G-1 (Panaxatriol Genin)

(1) Plants

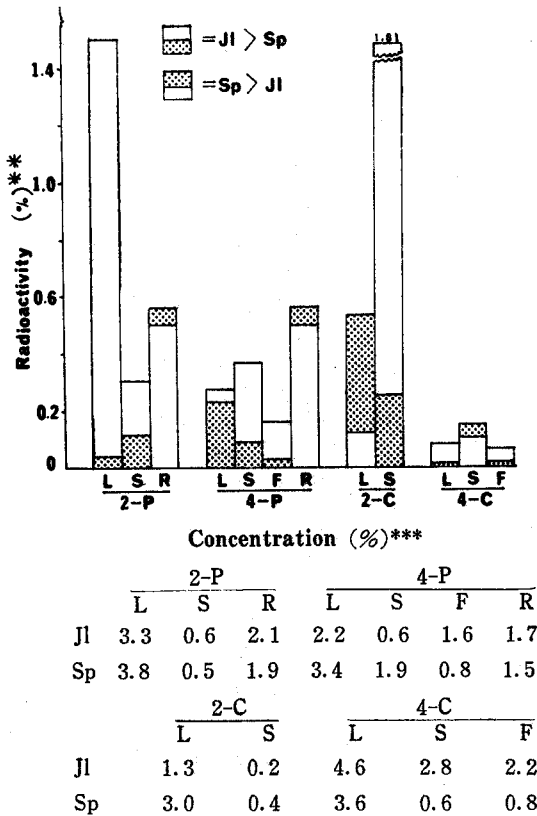
The average percentage of C¹⁴-acetate incorporation into panaquilin G-1 was similar (0.9 %) in both two- and four-year-old roots (Fig. 7). Panaquilin G-1, in both the labeled and non-labeled form, was absent from the stems and leaves (exception-leaves from four-year-old plants). The panaquilin G-1 concentration was approximately 8 times higher (2.4 %) in two-year-old roots collected in July than September (0.3 %).

The data suggest that panaquilin G-1 may be synthesized *de novo* in either the leaf and root, or may be present and translocated in some other chemical forms through the stems.

(2) Cuttings

The two-year-old leaves collected from July and September cuttings contained radioactive panaquilin G-1. In four-year-old cuttings, the July collected leaves contained only trace amounts of panaquilin G-1 (0.002 %). Panaquilin G-1 was not present in the stems of the cuttings.

Fig. 6. Panaquilin E C¹⁴-Radioactivity Distribution and Concentration (%) in American Ginseng Plants and Cuttings*.

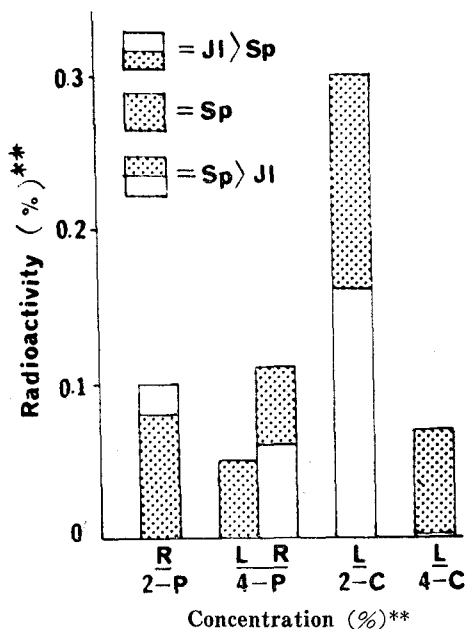


* Abbreviations: L-leaf; S-stem; F-fruit; R-root; 2-P: two-year-old plants; 4-P: four-year-old plants; C-cuttings; Jl-July collection; Sp-September collection.

** Radioactivity (%): (Radioactivity in panaquilin E (μCi) / Radioactivity uptake (μCi)) × 100.

*** Concentration (%): (Weight of panaquilin E (mg) / Weight of plants (mg dry weight)) × 100.

Fig. 7. Panaquinin G-1 C¹⁴-Radioactivity Distribution and Concentration (%) in American Ginseng Plants and Cuttings*.



	2-P		4-P		2-C	4-C
	R	L	R	L	L	L
Jl	2.4	—	0.3	0.4	0.2	0.2
Sp	0.3	0.1	0.1	1.1	0.8	0.8

* Abbreviations: L-leaf; S-stem; F-fruit; R-root; JI-July collection; Sp-September collection; 2-P: two-year-old plants; 4-P: four-year-old plants; C-cuttings.

** Radioactivity (%): (Radioactivity in panaquinin G-1 (μCi)/Radioactivity uptake (μCi))×100.

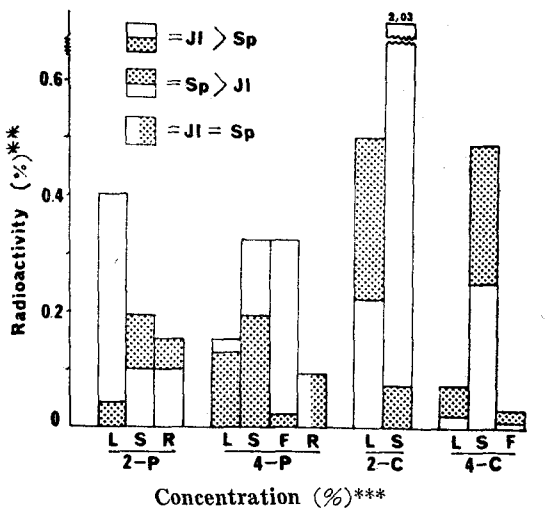
*** Concentration (%): (Weight of panaquinin G-1 (mg)/ Weight of plants (mg dry weight))×100.

f. Panaquinin G-2 (Unknown Genin)

(1) Plants

The average percentage of C¹⁴-acetate incorporation into panaquinin G-2 was high (0.3 %) in two- and four-year-old leaves collected in July as compared to two- and four-year-old leaves collected in September (0.07 %) (Fig. 8). Panaquinin G-2 activity was particularly high in two-year-old leaves collected in July (0.4 %) and in four-year-old July stems (0.3 %) and fruits (0.3 %). The average panaquinin G-2 activity in two-year-old roots (0.15 %) was higher than that in four-year-old roots (0.10 %).

Fig. 8. Panaquinin G-2 C¹⁴-Radioactivity Distribution and Concentration (%) in American Ginseng Plants and Cuttings*.



	2-P			4-P			
	L	S	R	L	S	F	R
Jl	2.9	1.7	2.2	0.2	0.5	0.9	0.2
Sp	0.8	0.8	0.4	0.9	1.5	0.5	0.4

	2-C		4-C		
	L	S	L	S	F
Jl	0.4	0.6	1.8	1.1	1.5
Sp	1.1	0.7	1.2	0.9	0.5

* Abbreviations: L-leaf; S-stem; F-fruit; R-root; JI-July collection; Sp-September collection; 2-P: two-year-old plants; 4-P: four-year-old plants; C-cuttings.

** Radioactivity (%): (Radioactivity in panaquinin G-2 (μCi)/Radioactivity uptake (μCi))×100.

*** Concentration (%): (Weight of panaquinin G-2 (mg)/ Weight of plants (mg dry weight))×100.

The concentration of panaquinin G-2 was highest in July collected plants (2.3 %). The immature (0.9 %) and mature (0.5 %) fruits also contained panaquinin G-2. The leaves contained more panaquinin G-2 than the roots (average 1.2 % and 0.8 %, respectively).

(2) Cuttings

The average percentage of C¹⁴-acetate in panaquinin G-2 in two-year-old cuttings (1.4 %) was higher than that of four-year-old cuttings (0.5 %). The stems collected in July contained a very high percentage of panaquinin G-2 activity (2.0 %). Except for these stems, the percentage into panaquinin G-2 was higher

in September cuttings (0.6 %) than July cuttings (0.3 %).

The concentration (%) of panaquilin G-2 was low in two-year-old cuttings collected in July (1.0 %) as compared to two-year-old cuttings collected in September (1.8 %) or four-year-old cuttings (3.5 %). A relatively high concentration of panaquilin G-2 was present in the immature fruits (1.5 %).

g. Summary

The results of C¹⁴-acetate incorporation into each panaquilin are summarized in Table IV.

Table IV. Panaquilin Radioactivity Distribution in American Ginseng Plants*.

Plant Part	Panaquilin						
	B	C	(d)	E	G-1	G-2	Total
Collection							
July	1.45	1.00	0.90	1.85	0.10	0.70	6.00
September	1.35	0.50	0.40	0.80	0.15	0.35	3.60
Age							
Two-year-old	1.05	0.60	0.75	1.50	0.10	0.50	4.50
Four-year-old	1.75	0.90	0.55	1.15	0.15	0.55	5.10

* Radioactivity (%): (Radioactivity in panaquilins (μCi)/Radioactivity C¹⁴-acetate uptake by plant (μCi)) \times 100. Average of four studies (July and September collections from two- and four-year-old plants).

5. Hydrolysis of Radioactive Methanol-2

a. Radioactivity of Hydrolysates

Aliquots of radioactive methanol-2 extracts (0.96~1.16 g, activity 0.37~2.52 $\mu\text{Ci}/\text{mg}$) (Table V) were hydrolyzed with a mixture of methanol and 30 % hydrochloric acid (4:1) by refluxing on a steam bath for 5 hrs. The hydrolyzed solution was evaporated and the residue was extracted with ether. The ether extracts weighed 0.09~0.30 g, were radioactive 0.76~6.13 $\mu\text{Ci}/\text{mg}$, and contained ginseng genins and

Table V. Hydrolysates of Methanol-2: C¹⁴-Activity.

Plant Material*	Methanol-2**		Hydrolysates***		Ratio
	Aliquot (g)	Activity ($\mu\text{Ci}/\text{mg}$)	Amount (g)	Activity ($\mu\text{Ci}/\text{mg}$)	
Two-year-old					
JIA	1.16	1.58	0.21	4.53	2.9
JIR	1.03	0.33	0.20	1.14	3.5
Average	1.10	0.96	0.21	2.84	3.2
SpA	1.10	0.69	0.33	1.01	1.5
SpR	0.96	0.41	0.23	0.76	1.9
Average	1.03	0.55	0.28	0.89	1.7
SpA (C)	1.06	1.04	0.30	1.54	1.5
Four-year-old					
JIA	1.11	2.52	0.22	6.04	2.4
JIR	1.14	0.57	0.27	1.72	3.0
Average	1.13	1.55	0.25	3.88	2.7
SpA	1.09	2.95	0.23	6.13	2.1
SpR	1.14	0.37	0.09	2.10	5.7
Average	1.12	1.66	0.16	4.12	3.9
SpA (C)	1.11	1.21	0.26	1.70	1.4

* Abbreviations: JI-July collection; Sp-September collection; A-above-ground parts; R-root; C-cutting.

** Methanol-2: Soluble extracts of methanol-1 with cold methanol (5° C).

*** Hydrolysates: Ether extracts of hydrolysates of methanol-2 with 30 % hydrochloric acid and methanol (1:4).

**** Ratio: Hydrolysate activity/Methanol-2 activity.

probably impurities such as fatty acids and flavonoids.

The average activity ($\mu\text{Ci}/\text{mg}$) of hydrolysates in the above-ground parts of plants (4.43 $\mu\text{Ci}/\text{mg}$) was higher than that in the roots (1.43 $\mu\text{Ci}/\text{mg}$). In cuttings, the hydrolysates from the above-ground parts contained approximately 1.60 $\mu\text{Ci}/\text{mg}$. The average activity of hydrolysates was lower in two-year-old plants (1.87 $\mu\text{Ci}/\text{mg}$) than that in four-year-old plants (4.00 $\mu\text{Ci}/\text{mg}$). The average activity of hydrolysates in four-year-old plants (4.00 $\mu\text{Ci}/\text{mg}$) was higher than either two-year-old plants collected in July (2.84 $\mu\text{Ci}/\text{mg}$) or September (0.89 $\mu\text{Ci}/\text{mg}$).

The ratio of hydrolysate activity to methanol-2 activity was 1.4~5.7 (Table V). The ratio was lower in the above-ground parts (average 2.2) than in the root (average 3.5). This may indicate that the root contained more ether-soluble hydrolysates or less impurities, such as pigments and carbohydrates than

the above-ground parts. The ratio (average) is higher in two-year-old plants collected in July (3.2) than in September (1.7), whereas the ratio is lower in four-year-old plants collected in July (2.7) than in September (3.6).

6. Radioactivity of Panaxadiol

Radioactive panaxadiol (0.42~2.96 m μ Ci/mg, 9~67 mg) was isolated from the ether-soluble extracts of hydrolysates by methanol elution of its silica gel tlc band (Table VI). This radioactivity was lower than that of the original ether extracts of the hydrolysate (0.67~6.13 m μ Ci/mg). The radioactive panaxadiol obtained from tlc could not be perfectly recrystallized, probably due to the presence of impurities. Non-radioactive panaxadiol (10~17 mg) was added to the panaxadiol (9~67 mg), and crystallized (Purified (1 \times): 6~24 mg, Table VI).

The average radioactivity of the combined, purified (1 \times) panaxadiol was 0.51 m μ Ci/mg in two-year-old plants and 0.69 m μ Ci/mg in four-year-old plants (Table VI). After three additional recrystallizations, the specific activity of the panaxadiol from two-year-old plants was 0.91 m μ Ci/mg (Table VI). Dilution and Recrystallization of C¹⁴-Panaxadiol.

Plant Material*	Panaxadiol** Amount (mg)	Panaxadiol** Activity (m μ Ci/mg)	Panaxadiol Added*** Amount (mg)	Purified (1 \times)**** Amount (mg)	Purified (1 \times)**** Activity (m μ Ci/mg)
Two-year-old					
J1A	23	2.77	15	18	0.91
J1R	11	1.29	10	15	0.54
SpA	67	0.42	10	6	0.10
SpR	48	0.67	11	24	0.49
Four-year-old					
J4A	14	2.96	12	13	0.91
J4R	9	2.23	17	19	0.65
SpA	42	1.11	15	17	0.23
SpR	15	1.93	11	15	0.98

* Abbreviations: J1-July collection; Sp-September collection; A-above-ground; R-root.

** Panaxadiol: Radioactive panaxadiol fractions obtained from preparative tlc bands.

*** Panaxadiol Added: Non-radioactive panaxadiol addition to radioactive panaxadiol fraction (mg).

**** Purified(1 \times): First recrystallized panaxadiol.

plants was 0.56 m μ Ci/mg and 0.54 m μ Ci/mg from four-year-old plants.

Experimental

The analytical procedures for the radioactive panaxadiol and panaxadiol were essentially identical to that has been described^{1,2)} except for the following:

1. Isotope Materials

Sodium acetate-U-C¹⁴ (1 mCi/1 ml, 102 mCi/mM, Lot No. C 32711) was purchased from Dhom Products, Ltd., North Hollywood, California. Toluene-C¹⁴ (5.67 \times 10⁵ \pm 3% dpm/g) for isotope standard was purchased from the Packard Instrument Co., Inc., Downers Grove, Illinois.

2. Feeding of Sodium Acetate-U-C¹⁴ to Intact Plants

An aqueous labeled acetate stock solution (10 ml) was prepared (100 μ Ci/mM) by adding non-radioactive sodium acetate (200 mg) to sodium acetate-U-C¹⁴ (1.0 ml, 1 mCi). Aliquots of the stock solution were then used to prepare feeding solutions (1 μ Ci/0.5 ml; 10 μ Ci/0.5 ml; 40 μ Ci/0.5 ml).

Tracer solutions (10 μ Ci/10~20 g fresh plant weight) were then fed to 100 two- (1 μ Ci/0.5 ml) and 20 four-year-old (40 μ Ci/0.5 ml) plants by the wick method³⁾. The plants took up the radioactive material in approximately 2 hrs. The tracer solution vials were washed with distilled water (0.2 ml) and the resulting solution was also taken up by the plant. The washing procedure was again repeated. The following day the thread and vials were removed from the plants and bamboo supports provided for plant. After one week, the plants were collected. Approximately 0.1~0.15 % of the radioactivity remained in the thread and vials.

3. Feeding of Sodium Acetate-U-C¹⁴ to Stem Cuttings

Labeled acetate solutions were fed by the wick method to 40 two-year-old stem cuttings (1 μ Ci/0.5 ml) and to 20 four-year-old stem cuttings (10 μ Ci/0.5 ml). The stem cuttings were maintained in a Hyponex solution (7-6-19, Hydroponic Chemical Co., Inc.,

Copley, Ohio). Approximately 0.004~0.06 % of the radioactivity remained in the thread and vials. The plants were collected for analysis after 5 days.

4. Autoradiochromatography⁵⁾

Kodak RP Royal X-Omat and liquid X-ray developer were used (Eastman Kodak Co., Rochester, N.Y.). The radioactivity of samples and standards was determined prior to their application to tlc plates to more properly estimate the X-ray film exposure time. The thin-layer chromatogram was placed directly on an unexposed X-ray film and placed in a cardboard X-ray cassette. The cassettes were covered with a heavy steel plate (approximately 1.5 kg) to maintain uniform contact between the film and plate, and the film exposed for the required time. Developed X-ray films were examined against a white background. The thin-layer chromatograms used to prepare the X-ray films were then sprayed with a ceric sulfate solution, and then compared.

요 약

방사성 동위원소화합물 소듐 아세테이트-U-C¹⁴(C¹⁴-아세테이트)를 2년생과 4년생 7월, 9월 미국인삼(오갈피 나무과, *Panax quinquefolium* L.) 식물과 컷팅(cutting)에 자기 심지법에 의하여 투여하였다. C¹⁴-아세테이트 섭취율은 약 99%였다. 오토라디오크로마토그램은 제조적 박층 크로마토그래피로 분리한 사포닌들이 불순물을 함유하고 있음을 제시하였으며 특히 잎이나 줄기 엑기스에서 분리한 사포닌들에 불순물이 많이 섞여 있음을 알았다. 뿌리 및 과실 메타놀 엑기스는 상대적으로 순수한 사포닌들을 얻을 수 있다. 페나퀼린 B의 과량으로 인한 페나퀼린 C에 대한 페나퀼린 B의 꼬리는 제조적 박층 크로마토그래피에서 서로 혼교되는 결과를 얻었다. 일차 정제된 사포닌들의 평균 함량(건조 식물에 대한 %)은 과실(9.8%), 줄기(7.9%), 뿌리(6.3%)에 비하여 잎(13.8%)에 높았다. C¹⁴-아세테이트가 페나퀼린으로 인코포레이트되는 평균 %는 48%였다. 페나퀼린 B와 C로 C¹⁴-아세테이트가 인코포레이트되는 평균 %는 페나퀼린 C (0.75%), (d) (0.65%), G-1 (0.13%) G-2 (0.53%)보다 높았다(페나퀼린 B 1.40%, C 1.13%). 페나퀼린 합성은 식물의 채취 부위, 채취

시기 및 연령에 따라 틀린다고 사료된다. 페나퀼린 B에 C¹⁴-아세테이트가 인코포레이트되는 평균 함량은 뿌리(0.58%)와 줄기(0.48%)에서 높았고, 페나퀼린 C (0.40%)와 (d) (0.45%)는 잎에서 높았고, 페나퀼린 E는 뿌리(0.55%)와 잎(0.50%)에서 자기 높았다. 페나퀼린 G-2는 식물의 모든 부위에서 생합성되어 졌다. 페나퀼린은 9월에 채집한 식물에서 보다 7월에 채집한 식물에서 보다 활발하게 생합성되는 것처럼 보였다(예의 페나퀼린 G-1). 페나퀼린 B, C, G-1은 4년생 식물에서 활발하게 생합성되고 페나퀼린 (d)와 E는 2년생 식물에서 활발하게 생합성된다고 사료된다. 컷팅에서 기대된 결과들은 페나퀼린들이 인삼 식물의 지상부위에서 새로히 합성되고 페나퀼린 G-1은 잎에서 새로히 합성된다고 하는 것들이다. 인삼 조직 배양 연구에서 알려진 바와 같이 페나퀼린들은 미국인삼과 한국인삼의 잎, 줄기, 뿌리 캘러스 조직에 의하여서도 합성될 뿐만 아니라 또한 그들의 캘러스 뿌리에서도 페나퀼린들이 합성된다. 따라서 인삼 사포닌인 페나퀼린은 인삼 식물의 세포나 기관등 모든 곳에서 새로히 합성된다고 사료할 수 있다. C¹⁴-아세테이트가 페나퀼린의 비당체 부분에도 인코포레이트되는 것을 입증하였는데, 2년생 식물에서 페낙사다이올은 0.56 μ Ci/mg, 4년생 식물에서 페낙사다이올은 0.54 μ Ci/mg 스페시픽 액티비티를 갖었다.

〈Received 6 January 1974〉

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