3', 4', 7-Trihydroxyflavone in Albizzia julibrissin

Prapaipit Chamsuksai*, Jae Sue Choi and Won Sick Woo

Natural Products Research Institute, Seoul National University, Seoul 110, Korea (Received 10 October 1981)

Abstract From the stembark of *Albizzia julibrissin*, 3', 4', 7-trihydroxyflavone and α -spinasteryl-D-glucoside were isolated.

Keywords Albizzia julibrissin; Leguminosae 3', 4', 7-trihydroxyflavone; α -spinasteryl-D-glucoside.

Previous workers have demonstrated that *Albizzia julibrissin* contains quercetin 3–O-galactoside and quercetin 3–O-rhamnoside¹⁾. We now report the isolation and identification of a rare 5-deoxyflavone, 3',4',7-trihydroxyflavone, from the stembark of the plant.

Column chromatography of the ethylacetate soluble fraction of the stem bark methanol extract on silica gel using a solvent (CHCl₃–MeOH-7%HAc=25:8:5) to give the flavonoid together with α -spinasteryl-D-glucoside.

3',4',7-*Trihydroxyflavone*, mp 314–316°, yellow needles crystallized from MeOH, showed M⁺ at m/e 270 (100%) and other peaks at m/e 242 (M–CO, 29), 137 (RDA fragment with A ring +H, 73) and 134 (RDA fragment with B ring, 31). The UV (in MeOH) showed absorption peaks characteristic of a flavone at 239 and 345 nm,

which were shifted by addition of NaOMe (259, 408 nm), NaOAc (255, 375 nm), NaOAc $+H_3BO_3(259, 372 nm)$ and AlCl₃ (239, 377 nm). Addition of HCl resulted in the decomposition of AlCl₃ complex, returning to original spectrum.

The ¹H NMR (60 MHz, DMSO-d₆, TMS) exhibited signals at δ 7.90 (1H, d, J=9Hz, H-5), 7.43(1H, d, J=2Hz, H-2'), 7.40(1H, dd, J=2 and 6.5 Hz, H-6') 6.96(1H, d, J=2Hz, H-8), 6.93(1H, dd, J=2 and 9 Hz, H-6), 6.89(1H, d, J=6.5Hz, H-5') and 6.62 (1H, s, H-3). These spectral data were in agreement with those for the structure of 3',4',7-trihydroxyflavone.

It was confirmed by the following ¹³C NMR data (22.50 MHz, DMSO-d₆, TMS) δ 176.6 (C-4) 163.0 (C-2) 162.8 (C-7) 157.6 (C-9) 149.1 (C-4') 145.8 (C-3') 126.2 (C-5), 122.2 (C-1') 118.2 (C-6') 116.2 (C-5') 116.0 (C-10) 114.6 (C-6) 113.2 (C-2') 104.8 (C-3) 102.5 (C-8).

The presence of this compound in the plants has previously been reported in *Trifolium* spp.^{2,3)} *Medicago sativa*⁴⁾ *Baptisia* spp^{5,6)}., *Juncus trifidus*, *Luzula purpurea*⁷⁾, *Salvertia convallariodora* and *Vochysia* spp⁸⁾.

This is the first report of its occurrence

^{*}Present address: Department of Chemistry, Faculty of Science, Chulalongkorn University, Bangkok, Thailand

in Albizzia genus.

α-Spinasteryl-D-glucoside, mp 278-281°, white needles crystallized from MeOH, is a well-known natural product; the aglycone, mp 167-169°, obtained by acid hydrolysis was identified by direct comparison with authentic sample (TLC, GLC and MS⁹⁾);the sugar was shown to be D-glucose (TLC and GLC as TMSi ether).

EXPERIMENTAL

Isolation of α -Spinasterol-D-glucoside.

The commercially available stem bark of *Albizzia julibrissin* reduced to a coarse powder was extracted with methanol to exhaustion and the extract was concentrated *in vacuo* to a drak brown viscous residue which was partitioned between equal volumes of n-hexane and water. The aqueous layer was partitioned with an equal volume of chloroform and subsequently partitioned with ethylacetate to effect an ethylacetate soluble fraction.

Every partition was complete when the organic solvent layer from the last partition was nearly colorless.

The ethylacetate phases were combined and concentrated *in vacuo* to dryness and dissolved in small amount of methanol and then stood at room temperature to yield colorless needles, mp 278–281°, IR $_{max}^{K3r}$ cm⁻¹: 3330 (OH) 1100–1000 (glycoside) 965 (*trans* disubstituted double bond) 840–790(trisubstituted double bond) MS m/e: 574 (M⁺), 395 (M–C₆H₁₁O₆). Acetate obtained by treatment with acetic anhydride in pyridine, mp 174–175°, NMR

(CDCl₃, TMS) δ : 0.55, 0.79 (3H, each, all s), 2.00, 2.02, 2.04 and 2.09 (3H each, all s), 4.60 (1H, d, J=7Hz).

Hydrolysis of α -Spinasteryl-D-glucoside.

The sample was dissolved in 5%HCl-MeOH and refluxed for 2 hr.

After cooling the reaction mixture was poured in water to give solids which were collected by filtration, when crystallized from MeOH, separated in colorless needles melting at mp 167–169°, identified by the direct comparison with an authentic sample of α -spinasterol.⁹⁾

The co-existance of trace amounts of Δ^7 -stigmastenol in the sample was shown by MS data (a small peak at m/e 414 besides the parent peak at m/e 412) and GC data (column, OVI, 60-80 mesh, $1.5m\times4mm$; column temp., 170° ; inj. temp. 190° ; detector temp. 200° ; N₂ 45ml/min; tr for α -spinasterol 5,8, for Δ^7 -stigmastenol, 6.5).

The filterate was concentrated *in vacuo* and refluxed with acid to hydrolyze methyl glucoside.

After neutralizing with Ag_2CO_3 , the aqueous solution was concentrated, in which only D-glucose was detected by TLC (MeOH-CHCl₃-Acetone-NH₄OH=5:2:2:3, Rf=0.23; CHCl₃-MeOH-H₂O=52:25:8, Rf=0.15) and GC of the TMS derivative (conditions, same as above; tr, 6.5, 7.2).

Isolation of 3',4',7-trihydroxyflavone.

The methanol solution, from which α -spinasteryl-D-glucoside had been separated by filtration, was evaporated to dryness, the residue subjected to column chromatography over silica gel using CHCl₃-MeOH-

7% HOAc = 25:8:5 to yield the flavonoid, crystallized from MeOH as yellow needles, mp 314–316°, UV $_{max}^{MOH}$ nm (log₈): 239 (4.32), 312 (4.21), 345 (4.36); with NaOMe, 259 (4.46), 318 (3.58), 340 (3.90), 408 (4.47); with NaOAc, 255 (4.13), 375 (4.25); with NaOAc+ H_3BO_3 , 259 (4.16), 308 (4.03), 372 (4.33); with AlCl₃, 239 (4.32), 305 (4.01), 377 (4.19); with AlCl₃+HCl 239 (4.25). 256 (4.11), 309(4.03), 347 (4.16).

ACKNOWLEDGEMENT

The authors are grateful to KOSEF for financial support and to UNESCO for a research fellowship to P.C.

LITERATURE CITED

- Kaneta, M., Hikichi, H., Endo, S. and Sugiyama, N., Identification of flavones in sixteen Leguminosae species. *Agric. Biol. Chem.*, 44, 1407 (1980).
- 2) Livingston, A. L. and Bickoff, E. M., Identification

- of 3', 4', 7-trihydroxyflavone in ladino clover. J. Pharm. Sci., 53, 1557 (1964).
- 3) Wong, E. and Francis, C. M., Flavonoids in genotypes of *Trifolium subterraneum*. *Phytochemistry*, 7, 2123, 2131, 2139 (1968).
- 4) Bickoff, E. M., Witt, S. C. and Livingston, A. L., 3', 4', 7-Trihydroxyflavone in alfalfa. *J. Pharm. Sci.*, 54, 1555 (1965).
- Markham, K. R. and Mabry, T. J., The identification of twenty-three 5-deoxy and ten 5-hydroxyflavonoids from *Baptisia lecontei* (Leguminosae). *Phytochemistry*, 7, 791 (1968).
- Markham, K. R., Mabry, T. J. and Swift, W. T., Jr., Distribution of flavonoids in the genus Baptisia (Leguminosae). Phytochemistry, 9, 2359 (1970).
- Williams, C. A. and Harborne, J. B.. Leteolin and daphnetin derivatives in the Juncaceae and their systematic significance. *Biochem. Syst. Ecol.*, 3, 181 (1975).
- Lopes, J. L. S., Lopes, J. N. C. and Lueitao Filho, H. F., 5-Deoxyflavones from the Vochysiaceae. *Phytochemistry*, 18, 362 (1979).
- Woo, W. S., Steroids and pentacyclic triterpenoids from *Phytolacca americana*. *Phytochemistry*, 13, 2887 (1974).