

## Studies on the Korean Indigenous Plants\* Isolation of 1-eicosanoyl caffeate from *Echinosophora koreensis*

Sam Sik Kang and Chang Min Kim<sup>†</sup>

Natural Products Research Institute, Seoul National University,

Seoul 110 and <sup>†</sup>Kangwon National University

Chunchun 200, Korea

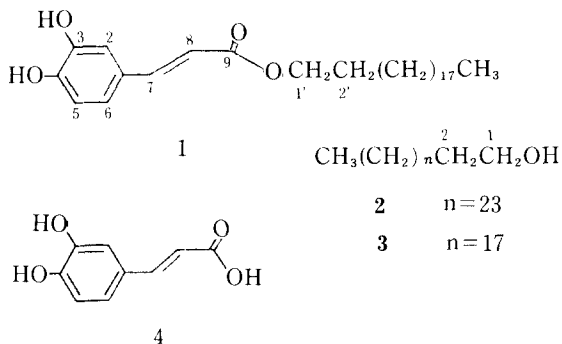
(Received December 29, 1986)

**Abstract** □ 1-Eicosanoyl caffeate, mp 109-110°, was isolated from the underground parts of *Echinosophora koreensis* together with hexacosanol, mp 75-6°. 1-Eicosanoyl caffeate was isolated for the first time from plant source.

**Keywords** □ *Echinosophora koreensis*, Leguminosae, 1-Eicosanoyl caffeate, Hexacosanol.

*Echinosophora koreensis* Nakai(Leguminosae) is indigenous to Korea and the only species of this genus of the family Leguminosae. Earlier investigations on this plant showed the presence of sterols, coumarin and triterpenoids<sup>1)</sup> and flavonoids<sup>2)</sup>. Further studies on the underground parts of this plants has resulted in the isolation of a new compound, 1-eicosanoyl caffeate together with hexacosanol.

Repeated chromatographic separations of ether soluble portion of the MeOH extract over silica gel and LiChroprep<sup>®</sup> RP-8 yielded compound **1** as needles, mp 109-110°. The IR spectrum of **1** showed the presence of hydroxyl at 3490 and 3320 cm<sup>-1</sup>, α, β-unsaturated ester carbonyl at 1688 cm<sup>-1</sup>, aromatic at 1609 and 1536 cm<sup>-1</sup> and polymethylene groups at 715 cm<sup>-1</sup> which suggested to be that aromatic ester of higher alcohol. This was further corroborated by its UV spectrum which was virtually identical to that of ethyl caffeate<sup>3)</sup>. The NMR spectrum of **1** showed signals for three aromatic protons at δ 6.94(1H, d, J=7.2 Hz, H-5), 7.03(1H, brd, J=7.2 Hz, H-6) and 7.08(1H, brs, H-2) and shows a pair of one proton doublets appearing at δ 6.25 and 7.57(1H each, J=15.9 Hz, H-8 and 7) typical of a trans olefin. In the region of upfield, the n-alkyl ester functional group was observed which appeared methyl signal at δ 0.87(3H, brt, J=6 Hz), (CH<sub>2</sub>)<sub>n</sub> at 1.26(34 H, s), -COOCH<sub>2</sub>CH<sub>2</sub> at 1.67(2H, m), and -COOCH<sub>2</sub> at 4.18(2H, t, J=6 Hz)<sup>4,5)</sup>. These data indicated that **1** was a caffeic acid ester of higher alcohol.



The higher alcohol was deduced as eicosanol (**3**) from the mass spectrum of **1**. The ester **1** on alkaline hydrolysis yielded eicosanol (**3**) and caffeic acid (**4**) which were identified by direct comparison with authentic samples. Therefore the compound **1** was identified as eicosanoyl caffeate that hitherto had not been found in plants.

The second compound (**2**) was crystallized from petroleum ether to give colorless flakes, mp 75-6°. The IR spectrum of this compound indicated the presence of hydroxyl at 3350 cm<sup>-1</sup> and polymethylene at 726 and 716 cm<sup>-1</sup> typical of higher alcohols. The NMR spectrum of **2** was similar to compound **1**: methyl signal at δ 0.88 (3H, brt, J=6 Hz), long chain methylene protons at δ 1.25(46 H, s), methylene protons adjacent to primary hydroxyl group at δ 1.52(2H, m) and methylene protons adjacent to an oxygen at δ 3.63(2H, m). This data suggested that **2** was hexacosanol. This assumption was supported by mass spectrum of **2**. It showed an ion peak at m/z 364

\*Part 2 in the series "Korean indigenous plants". For part 1 see ref (1).

in the high mass region which corresponded to the loss of one molecule of H<sub>2</sub>O from the molecular ion<sup>6</sup>). Other fragment peaks for [C<sub>n</sub>H<sub>2n+1</sub> O<sup>+</sup> - H<sub>2</sub>O] and [M<sup>+</sup> - (C<sub>n</sub>H<sub>2n</sub> + H<sub>2</sub>O)] ions<sup>6</sup>) strongly supported that **2** was hexacosanol. Direct comparison with an authentic sample established its identity.

## EXPERIMENTAL METHODS

Melting points were determined on a Mitamura-Riken apparatus and are uncorrected. IR spectra were recorded on a Perkin-Elmer 283B spectrophotometer. NMR spectra were recorded on a Varian FT-80A spectrometer and are given in ppm( $\delta$ ) downfield from an internal TMS standard. Mass spectra were determined on a Hewlett-Packard 5985B GC/MS system at 70 eV using direct inlet system. UV spectra were runned with Gilford System 2600 spectrophotometer.

### Plant collection and extraction

This was carried out as described previously<sup>1)</sup>.

### Fractionation and isolation

The MeOH extract was partitioned between ether and water. The ether fraction was chromatographed over silica gel and eluted with CHCl<sub>3</sub> and then MeOH-CHCl<sub>3</sub> (1 : 49) to give subfractions 1 to 3 and 4 and 5, respectively. Subfraction 5 was concentrated and allowed to stand at room temperature. The precipitate was filtered and the filtrate was subjected to column chromatography on a prepacked LiChroprep<sup>R</sup> RP-8 reversed phase column with MeOH-H<sub>2</sub>O (8 : 2) to afford compound **1** and **2**. Compound **1** was crystallized from MeOH as needles. Compound **2** was crystallized from petroleum ether as colorless flakes.

### Compound 1

mp 109-110°. IR  $\nu_{max}^{KBr}$  cm<sup>-1</sup> 3490, 3320(OH), 2922, 2850, 1475(CH), 1688, 1285, 1170 ( $\alpha$ ,  $\beta$ -unsaturated ester carbonyl), 1609, 1536(aromatic C=C), 975(trans double bond), 715[(CH<sub>2</sub>)<sub>n</sub>]. UV  $\lambda_{max}^{MeOH}$  nm(log  $\epsilon$ ) 221(3.84), 236(sh. 3.69), 246(3.71), 303(3.85), 330(3.96). NMR(CDCl<sub>3</sub>, TMS)  $\delta$  0, 87(3H, brt, J=6Hz, CH<sub>3</sub>), 1, 26[34 H, s, (CH<sub>2</sub>)<sub>n</sub>], 1, 67(2H, m, H-2'), 4, 18(2H, t, J=6 Hz, H-1'), 6, 25(1H, d, J=15.9 Hz, H-8), 6, 94(1H, d, J=7, 2 Hz, H-5), 7, 03(1H, brd, J=7.2 Hz, H-6), 7, 08(1H, brs, H-2), 7, 57(1H, d, J=15, 9 Hz, H-7). MS,  $m/z$ (rel. int.) 460(M<sup>+</sup>, 1.0), 404(M<sup>+</sup>-C<sub>4</sub>H<sub>8</sub>, 22.0), 260(1.1), 245(0.8), 236(2.6), 219(3.0), 182(29.7), 181(17.7), 180(caffeic acid, 100), 163(56.0), 145(10.7), 136(180-CO<sub>2</sub>, 19.0), 135(163-CO, 12.3), 134(12.

2), 123(11.3), 97(5.9), 83(8.4), 69(13.4), 55(15.9).

### Hydrolysis of 1

**1** (30 mg) was hydrolyzed with 5% KOH in EtOH for 3 hr at 60-70°. The reaction mixture was partitioned with ether and water. The ether layer was concentrated and the residue was crystallized from petroleum ether to afford colorless flakes. mp 63-5°, which was identified as eicosanol (**3**) by direct comparison with an authentic sample. The aqueous layer was acidified with d-HCl and extracted with ethylacetate. After evaporation the residue was crystallized from MeOH-H<sub>2</sub>O to yield caffeic acid (**4**) as pale yellow powder, mp 220-4°, and direct comparison with an authentic sample established its identity.

### Compound 2

mp 75-76° [Lit.<sup>7)</sup> mp 78°]. IR  $\nu_{max}^{KBr}$  cm<sup>-1</sup> 3350, 1060(OH), 2922, 2855, 1475, 1465(CH), 726, 716[(CH<sub>2</sub>)<sub>n</sub>]. NMR(CDCl<sub>3</sub>, TMS)  $\delta$  0, 88(3H, brt, J=6Hz, CH<sub>3</sub>), 1, 25[46 H, s, (CH<sub>2</sub>)<sub>n</sub>], 1, 52(3H, m, H-2 and OH), 3, 63(2H, m, H-1). MS,  $m/z$  (rel. int.) 364(M<sup>+</sup>-H<sub>2</sub>O, 1.5), 336(3.8), 308(0.9), 250(0.7), 237(0.9), 223(1.7), 209(1.9), 195(2.2), 181(2.9), 167(4.0), 153(5.9), 139(10.3), 125(24.5), 111(48.2), 97(95.0), 83(100), 69(78.4), 57(65.0), 55(58.3), 43(33.7), 41(20.1).

## LITERATURE CITED

1. Kim, C.M. and Kang, S.S.: Studies on the Constituents of the Stems of *Echinosophora kor-eensis*. *Yakhak Hoeji* **30**, 139(1986).
2. Kim, C.M., Sankawa, U. and Ebizuka, Y.: Abstracts of Papers, the 17th Annual Meeting of the Korean Society of Pharmacognosy, Seoul, Korea, pp 22-23(1986).
3. Daniels, D.G.H., King, H.G.C. and Martin, H. F.: Antioxidants in Oats: Esters of Phenolic Acids. *J. Sci. Food Agric.* **14**, 385(1963).
4. Isobe, T., Noda, Y. and Kubota, T.: The Chemical Constituents from the Roots of *Rabdosia japonica*. *Nippon Kagaku Kaishi* 799(1985).
5. Chatterjee, A., Dhara, K.P., Rej, R.N. and Ghosh, P.C.: Hexacosylferulate, A Phenolic Constituents of *Pinus roxburghii*. *Phytochem.* **16**, 397(1977).
6. McLafferty, F.W.: *Interpretation of Mass Spectra*, 2nd ed., The Benjamin/Cummings Publishing Company, Inc., London. pp 113-116(1973).
7. Banerji, R., Misra, G. and Nigam, S.K.: Butyric Acid, A New Sapogenin from *Madhuca butyracea*. *Planta Med.* 280(1985).