# Secretions of the biliary mucosa in experimental clonorchiasis

Soon-Hyung Lee\*, Jong-Inn Lee<sup>1)</sup>, Sun Huh<sup>2)</sup>, Jae-Ran Yu<sup>3)</sup>, Sei Woo Chung<sup>4)</sup>, Jong-Yil Chai and Sung-Tae Hong

Department of Parasitology and Institute of Endemic Diseases Seoul National University College of Medicine, Seoul, 110-799, Korea

Abstract: The histological change of the biliary mucosa in clonorchiasis is characterized as adenomatous hyperplasia, and cross-sectioned mucosa looks like intestinal mucosa. In addition to the glandular hyperplasia, the metaplasia of mucin secreting cells is also known. The present study investigated the presence of intestinal secretion from the biliary mucosal cells of rabbits and rats with Clonorchis sinensis infection. The rabbit was infected with 300 and the rat was infected with 100 metacercariae of C. sinensis. A part of the animals were followed up after praziquantel treatment. The rabbit livers were prepared for histochemistry to observe any endocrine secretion and the bile duct mucosa of the mice was processed for the activity of brush border membrane (BBM)-bound enzymes of the small intestine. Immunohistochemistry with the polyclonal antibodies and biotinstreptavidin-peroxidase staining kit showed no positive cells for gastrin and secretin, but a few cells were positive for serotonin. The proliferated biliary mucosa of the mice revealed no activity of dissaccharidases and aminopeptidase. Only alkaline phosphatase activity was found both in the control and the infected. The hyperplastic biliary mucosal cells showed no gastrointestinal secretory functions. The serotonin secreting cells may be one of the inflammatory cells.

**Key words:** Clonorchis sinensis, biliary epithelial cells, gastrin, secretin, serotonin, brush border membrane enzymes, alkaline phosphatase

#### INTRODUCTION

Clonorchiasis is a liver fluke disease caused by Clonorchis sinensis which is prevalent in Chira, North Indochina, Japan and Korea, At present, it is the most commonly detected parasite of human by fecal examination (Chai, 1990), and the infected population is now estimated about one million in Korea. This is a remarkably decreased number when compared with previous data but still its prevalence is high along the rivers (Kim et al., 1990). Praziquantel is the drug of choice for fluke diseases, and also highly effective in clonorchiasis (Rim et al., 1981; Seo et al.,

<sup>·</sup> Received Nov. 2 1992, accepted Jan. 7 1993.

<sup>•</sup> This study was supported by the Research Grant from the Korea Science and Engineering Foundation (1992, number 921-1600-003-1).

<sup>\*</sup> Corresponding author

<sup>•</sup> Present address: 1) National Cancer Hospital. Dept. of General Surgery. Seoul 139-240; 2) Hallym University, Dept. of Parasitology. Chunchon 200-702; 3) Kon-Kuk University, Dept. of Parasitology. Chungju 380-701; 4) Dept. of Pediatrics, College of Medicine, Inha University, Inchon 402-751.

1983; Lee, 1984). The decrease of clonorchiasis owes mainly to the introduction of praziquantel in early 1980s. However, its prevalence is rather high in contrast to rapid and distinctive declining of the soil-transmitted helminthiases in Korea. The situation may be due to the obstinate habit of raw consumption of fresh water fish (MHSA & KAH, 1992).

The epithelium of the infected bile ducts showed marked glandular hyperplasia and goblet cell metaplasia (Lee et al., 1978a & 1978b; Song et al., 1989). The changes persisted more than 6 months after praziquantel treatment (Lee et al., 1987 & 1988). More residual changes were in the bile ducts with severe histopathological changes (Lee et al., 1987). Mucin secretion disappeared but the hyperplasia was still found in mild degree even 12 months after treatment (Hong et al., 1990).

Some sections of the heavily infected bile ducts simulate gross appearance of the cross sectioned small intestine because of the characteristic hyperplasia of the biliary mucosa. Furthermore, goblet cells appear at the base of the epithelium. Those findings suggest that *Clonorchis* worms make the epithelium of bile ducts proliferate in shape of the small intestine. Is the change confined only in the morphology?

The small intestine has some secretory functions other than mucin production. Major secretions are gastrointestinal hormones and digestive enzymes. The APUD (amine precursor uptake and decarboxylation) cells secrete gastrin, secretin, serotonin, glucagon, and somatostatin to control the functions of the digestive tract (McGuigan, 1989). As for the final digestion in the intestine, the brush border membrane (BBM) retains several disaccharidases, aminopeptidases and phosphatases (Anderson et al., 1983).

The bile duct and the small intestine share their ancestral origin in the development. Therefore, a hypothesis can be proposed that some kind of stimulations from the fluke make the bile ducts change into the intestine. The present study was executed to observe any secretory functions of the small intestine from the infected biliary mucosa, and to reevaluate the pathogenesis of clonorchiasis.

#### MATERIALS AND METHODS

#### 1. Infection of animals with C. sinensis

Rabbits and rats were experimentally infected by introducing the metacercariae of C. sinensis into the stomach, 300 and 100 respectively. The metacercariae were collected from peptic digested material of Pseudorasbora parva which were captured at the endemic area, the Nakdong River, Kyongsangnam-do. The animals were reared in the laboratory by feeding commercial diet and tap water for 14 weeks. Total 8 rabbits were infected, and half of them were treated with praziquantel 100 mg/kg  $\times$  2 days. The treated rabbits were sacrificed 6 or 12 months after the treatment.

## 2. Immunohistochemical staining for the endocrine functions

Normal control, infected and treated rabbits were prepared for routine histopathological observation by sectioning their livers in paraffin blocks. The ABC staining kits with polyclonal antibodies and biotinylated anti-IgG antibody, peroxidase conjugated streptavidin (AtrAviGen, BioGenex Lab., U.S.A.) were used for detection of gastrin, secretin, and serotonin. Following the packaged procedure, the sections were deparaffinized and hydrated. Endogenous peroxidase was blocked by 10 minute incubation in PBS (pH 7.6) containing 3% H<sub>2</sub>O<sub>2</sub>. Polyclonal antibodies were reacted 30 minutes at room temperature with one negative serum control. After thorough rinsing with PBS, the link solution of biotinylated anti-IgG antibody was incubated for 30 minutes at room temperature. Washed several times, and peroxidase labeled streptavidin was to the sections for 20 minutes. The color reaction was developed by 3-amino-9-ethylcarbazole (AEC) in N,N-dimethylformamide, and controlled the intensity by the reaction time from 7 to 10 minutes. Counter stain used Mayer's hematoxylin.

#### 3. Activities of BBM enzymes

The common bile ducts of the infected rats and normal controls were isolated. Their mucosal layer was scraped on ice and stored at -70°C in 0.05 M mannitol 2 mM Tris HCl (pH 7.0). The tissue was homogenated with a teflon tipped homogenizer (Stir-R model S63C Tri-R Inst. Inc., U.S.A.) and then with an ultrasonic sonicator (Sonicator W-385, Heat Systems-Ultrasonics Inc., U.S.A.). Their protein contents were quantitated following modified Lowry method. The absorbances were checked through a UV/VIS spectrophotometer (Hewlett-Packard Co., U.S.A.). The mucosal homogenates were reacted with the substrates of disaccharides (sucrose, lactose, trehalose), pnitrophenyl phosphate (pH 10.0), and leucine naphtylamide HCl. The enzyme activities were estimated by the absorbances after color reaction. The activities of the infected rats were compared with those of the control.

#### RESULTS

#### 1. Endocrine secretions

ABC staining showed several gastrin positive cells in the stomach wall as a positive control, but no cells were stained in the bile ducts of the control, infected or treated rabbits (Figs. 1 & 2). Secretin staining was positive only in the small intestine as a positive control, and negative in the bile ducts of all group of rabbits (Figs. 3 & 4). The ABC staining for serotonin detected a few positively stained cells among the epithelial cells of infected rabbits. The cells were slender with a round nucleus, and included brown granules in their cytoplasm (Figs. 5-10). No cell was positive in the treated rabbits (Figs. 11-12). Also the edematous stroma layer of the hyperlastic tissue was stained for serotonin. These findings are summarized in Table 1.

#### 2. BBM enzyme activities

The activities are expressed in units as one unit corresponds to hydrolysis of 1 M substrate per minute at 37°C. Sucrase activities were from 2.6 to 12.1 mu/mg protein while 4.4 mu/mg protein in the control. Lactase activities ranged from 0 to 34.4 mu/mg protein and that of control was 0. Trehalase activities were in the range of 0 to 32.9 mu/mg protein (control 6.9 mu/mg protein). Alkaline phosphatase showed higher activities than other enzymes, control 63.8

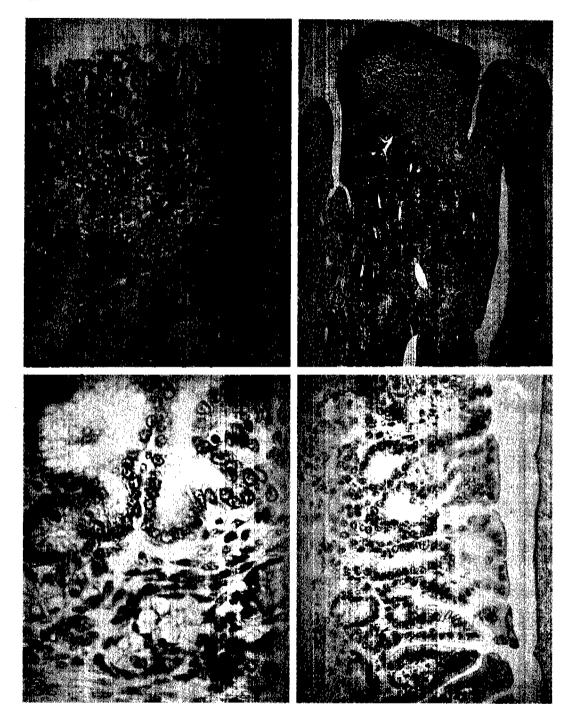
mu/mg protein and the infected rats 30.6-156. 1 mu/mg protein. Leucine aminopeptidase activities were 11.6 mu/mg protein in the control, and 6.4 to 18.1 mu/mg protein in the infected rats. The activities are presented in Table 2.

#### DISCUSSION

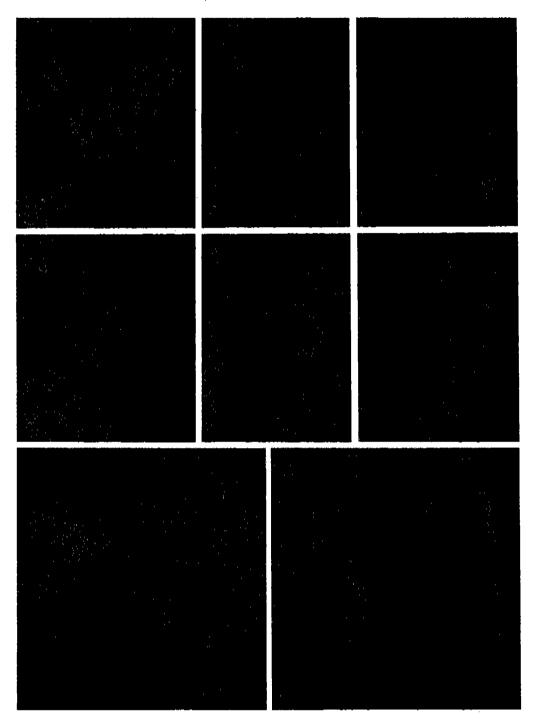
As observed in the bile ducts of rabbits infected with C. sinensis, the hyperplastic biliary mucosa is known to secrete neutral and acid mucopolysaccharide (Chou and Gibson, 1970; Hong et al., 1990). The mucopolysaccharide producing cells are normally absent in the biliary mucosa. The mucopolysaccharide in the intestinal tract is known to play a role in protection of the host by two ways. The one is covering the brush border membrane, and the other is expediting expulsion of foreign materials especially wellknown in nematode infections (Roitt, 1988). The goblet cells increase by parasitic infections in the intestine, and the function is associated with cellular or humoral immunity (Tse and Chadee, 1991).

Little has been revealed for the cell reaction at the biliary mucosa in clonorchiasis. Hong et al. (1990) suspected any role of hypersensitive reaction in the pathogenesis of adenomatous hyperplasia and goblet cell metaplasia in clonorchiasis. In fascioliasis, proline secreted from the fluke was known to induce proliferation of the biliary mucosa (Isseroff et al., 1977).

Other than mucopolysaccharides, the glandular structured mucosa contains a few serotonin secreting cells and the activity of alkaline phosphatase. Serotonin (5-hydroxytryptamine) is one of major neurotransmitters in the central nervous system and the vasoconstricters. The majority of its content is stored in the enterochromaffin cells of the small intestine. The mast cell of rodents harbours it, but not the cell of humans (Douglas, 1987). The serotonin containing cells were one or two on a section of the biliary tree, and not found after treatment though mucin secretion and the hyperplasia lasted more than 6 months after treatment (Hong et al., 1990). The serotonin positive cells seem to be mast



**Fig. 1.** Mucosa of the stomach as a positive staining for gastrin,  $\times$  200. **Fig. 2.** The proximal intrahepatic bile duct of an infected rabbit with negative gastrin staining,  $\times$  100. **Fig. 3.** Mucosa of the jejunum as a positive staining for secretin,  $\times$  400. **Fig. 4.** Negative secretin staining in the biliary mucosa of an infected rabbit,  $\times$  200.



**Fig. 5.** Serotonin positive cells in the jejunum as a positive reference staining,  $\times$  400. **Figs. 6-10.** Serotonin positive cells (arrows) in the biliary mucosa of the infected rabbits,  $\times$  400. **Fig. 11.** Negative serotonin staining in the biliary mucosa of the infected rabbits 6 months after treatment,  $\times$  200. **Fig. 12.** Negative serotonin staining in the biliary mucosa of the infected rabbits 12 months after treatment.  $\times$  100.

**Table 1.** Immunohistochemical findings of the biliary mucosa in rabbit clonorchiais

Group	Location	ABC staining for		
		gastrin	secretin	serotonin
Uninfected	Intrahepatic duct	<del>_</del>	_	_
control	Extrahepatic duct	+*	_	+
	Stomach	+++	nd	nd
v	Jejunum	_	++	++
Infected	Intrahepatic duct	_	_	+
6M treatment	Intrahepatic duct	_	_	<u>.</u>
12M treatment	Intrahepatic duct	_	_	_

<sup>\*+:</sup> minimal changes found in less than 1/4 field of a section duct

nd: not done

**Table 2.** Activities of the brush border membrane bound enzymes in the biliary mucosa of rats infected with *C. sinensis* 

unit:	mu	/mg	protein
-------	----	-----	---------

Enzymes	Uninfected control	Infected
Sucrase	4.4	4.9± 3.1
Lactase	0.0	$4.3 \pm 12.7$
Trehalase	6.9	$2.9 \pm 19.6$
Alkaline phosphatase	63.8	$67.3 \!\pm\! 46.6$
Leucine aminopeptidas	se 11.6	$10.1 \pm 4.8$

cells or other inflammatory cells rather than metaplastic enterochromaffin cells.

Still any information on mast cell reaction in clonorchiasis is not available, however, the cells are known to increase locally in the intestine by parasite infections (Woodbury et al., 1984; Guy-Grand et al., 1984; Novak and Nombrando, 1988; Kho et al., 1990). Mast cells can be infiltrated into the mucosa with other inflammatory cells by the irritation of the liver fluke. This reaction should be evaluated further.

The activity of alkaline phosphatase was found in the proliferated epithelium, but not increased compared to that of the control. The enzyme is universal in the epithelium-lined tubular organs of mammals, such as the stomach, intestine, bile duct, kidney tubule, etc. (Miura et al., 1982). Since the enzyme activity was not changed by the infection

though the epithelial cells proliferated severely, the infection and associated hyperplasia might not influence the cell maturity. Other BBM enzymes than alkaline phosphatase were of negligible activity in the biliary mucosa.

The present findings confirm that the mucosal change of the biliary mucosa in clonorchiasis confines to the morphological intestinalization. Though cross sections of the bile ducts give the appearance of the sectioned intestine, the ducts showed no secretory functions of the gastrointestinal tract except production of mucopolysaccharide. The epithelial cells lining the biliary mucosa seem to differentiate enough not to convert to the intestinal epithelial cells. The change of biliary mucosa in clonorchiasis is not the metamorphosis to the small intestine.

#### ACKNOWLEDGEMENT

The present study was supported by the Research Grant from the Korea Science and Engineering Foundation, No. 921-1600-003-1.

#### REFERENCES

Andersen KJ, Schionsby H, Skagen DW (1983) Jejunal mucosal enzymes in untreated and treated coeliac disease. *Scand J Gastroenterol* **18**: 251-256.

Chai JY (1990) Positive rates of intestinal parasites by stool examination in the Seoul

<sup>++:</sup> moderate changes found in 1/2 field of a sectioned duct

<sup>+++:</sup> heavy changes found in more than 3/4 field of a sectioned duct

- National University Hospital (unpublished data).
- Chou ST, Gibson JB (1970) The histochemistry of biliary mucins and the changes caused by infestation with *Clonorchis sinensis*. *J Pathol* **101**: 185.
- Douglas WW (1987) Histamine and 5-hydroxytryptamine (serotonin) and their antagonists. The pharmacological basis of therapeutics, 7th ed.: 628-633, edited by Gilman AG, Goodman LS, Rall TW, Murad F, Macmillan Publishing Co., N.Y.
- Guy-Grand D, Michel DY, Luffau G, Vassalli P (1984) Gut mucosal mast cells. Origin, traffic, and differentiation. J Exp Med 160: 12-28.
- Hong ST, Huh S, Kho WG, et al (1990) Changes of histopathological and serological findings of the liver after treatment in rabbit clonorchiasis. Seoul J Med **31**(2): 117-127.
- Isseroff H, Sawma J.T, Reino D (1970) Fascioliasis. Role of proline in bile duct hyperplasia. Science 198: 1157-1159.
- Kho WG, Chai JY, Chun CH, Lee SH (1990) Mucosal mast cell responses to experimental Fibricola seoulensis infection in rats. Seoul J Med 31(3): 191-199.
- Kim SS, Han MH, Park SG, Lim HS, Hong ST (1990) A survey on the epidemiological factors of clonorchiasis in the Pohang industrial belt along the Hyungsan River, Kyongsangbuk-do. Korean J Parasit 28(4): 213-219.
- Lee SH (1984) Large scale treatment of Clonorchis sinensis infection with praziquantel under the field conditions. Arzneim-Forsch/Drug Res 34: 1227-1230.
- Lee SH, Chai JY, Yang EC, Yun CK, Hong ST, Lee JB (1988) Observation of liver pathology after praziquantel treatment in experimental Clonorchis sinensis infection in guinea pigs. Seoul J Med 29(3): 253-262.
- Lee SH, Hong ST, Kim CS, Sohn WM, Chai JY, Lee YS (1987) Histopathological changes of the liver after praziquantel treatment in Clonorchis sinensis infected rabbits. Korean J Parasit 25(2): 110-122.
- Lee SH, Shim TS, Lee SM, Chi JG (1978a) Studies on pathological changes of the liver in albino rats infected with *Clonorchis sinensis*. Korean

- J Parasit 16: 148-155 (in Korean).
- Lee SK (1983) Histopathological study on the liver with infection of *Clonorchis sinensis*. *J Pusan Med College* **23**(2): 41-48 (in Korean).
- Lee SY, Lee SH, Chi JG (1978b) Ultrastructural changes of the hepatocytes and biliary epithelia due to Clonorchis sinensis in guinea pigs. Korean J Parasit 16(2): 88-102 (in Korean).
- McGuigan JE (1989) Hormones of the gastrointestinal tract. In DeGroot L.J. Endocrinology, 2nd ed: 2764-2768. Saunders Co, Philadelphia.
- Ministry of Health and Social Affairs and Korea Association of Health (1992) Evaluation of the mass-treatment project of clonorchiasis in Korea.
- Miura S, Asakura H, Morishita T, et al (1992) Changes in intestinal alkaline phosphatase activity in cholera toxin-treated rats. Gut 23: 507-512.
- Novak M, Nombrando S (1988) Mast cell responses to *Hymenolepis microstoma* infection in mice. *J Parasit* **74**: 81-88,
- Rim HJ, Lyu KS, Lee JS, Joo KH (1981) Clinical evaluation of the therapeutic efficacy of praziquantel (Embay 8440) against Clonorchis sinensis infection in man. Ann Trop Med Parasit 75: 27-33.
- Roitt I (1988) Essential immunology. 6th ed Blackwell Sci Publ, Oxford.
- Seo BS, Lee SH, Chai JY, Hong ST (1983) Praziquantel (Distocide) in treatment of Clonorchis sinensis infection. Korean J Parasit **21**(2): 241-245.
- Song GA, Kim JD, Lee DW, et al (1989) Histopathological and histochemical studies on the intrahepatic duct in rabbits experimentally infected with Clonorchis sinensis. Korean J Int Med 37(3): 344-355 (in Korean).
- Tse SK, Chadee K (1991) The interaction between intestinal mucus glycoproteins and enteric infections. *Parasit Today* **7**(7): 163-172.
- Woodbury RG, Miller HRP, Huntrey JF, Newlands GFJ. Palliser AC, Wakelin D (1984) Mucosal mast cells are functionally active during spontaneous expulsion of intestinal nematode infections in rats. *Nature* **321**: 450-452.

=국문초록=

### 간횹층에 감염된 토끼 및 흰쥐 담관상피의 분비기능

서울대학교 의과대학 기생충학교실 및 풍토병연구소", 원자력병원 일반외과<sup>2</sup>, 한림대학교 의과대학 기생충학교실<sup>3</sup>, 건국대학교 외과대학 기생충학교실<sup>4</sup> 인하대학교 의과대학 소아과학교실<sup>9</sup>

이순형" · 이종인" · 허선" · 유재란 · 정세우 · 채종일" · 홍성태"

간흡충에 감염되었을 때 나타나는 조직병리학적 소견의 변화로는 담관 직경이 확장되고 담관벽이 비후되며, 담관 상피층에 선종성 중식과 배세포 확생이 관찰되었다. 치료 후에는 이러한 변화 중에서 화생이 소실되나 선종성 증식은 상당히 완화되어 잔존하고 담관벽의 비후는 지속된다. 이러한 형태학적인 변화는 선종성인 특징이 있어 어떠한 분비기능이 예상되었다. 이 연구에서는 특히 소장의 분비물이 담관 점막층에서 분비되는가를 확인하고자 하였다. 간흡충 피낭유충을 300개씩 8마리의 토끼에 감염시킨 후, 14주에 프라지콴텔로 치료하여 구층 후 6개월 및 1년에 간을 적출하고, 파라핀 포매하여 절편을 만들어 분비물에 대하여 면역조직화학법을 시행하였다. Gastrin과 secretin은 정상토끼의 간내 담관 및 감염된 담관에서 나타나지 않았다. Serotonin은 비감염 간 내당관에서는 관찰되지 않았으나, 감염 대조군의 담관상피충에서 몇 개의 양성 세포가 관찰되었다. 그러나 치료 후 6개월에는 관찰되지 않았다. 또한 흰쥐에 100개씩 간흡충 피낭유충을 감염시키고 중식된 상피층에서 소장 미소용모막 효소의 활성을 관찰한 바, alkaline phosphatase의 활성이 감염군에서 대조군과 같은 정도로 관찰되었으나 sucrase, trehalase, lactase, leucine aminopeptidase의 활성은 나타나지 않았다. 이 결과를 통하여 간흡충 감염시에 중식된 담관상피에 소수의 serotonin 분비세포가 있음을 관찰하였고 다른 소장의 분비기능은 생기지 않음을 확인하였다. 이 serotonin을 포합한 세포는 비만세포로 추정된다.

(기생충학잡지 31(1): 13~20, 1993년 3월)