NMR Assignments of Two Furofuran Lignans from Sesame Seeds

Sam Sik Kang, Ju Sun Kim, Jee Hyung Jung¹ and Young Hee Kim²

Natural Products Research Institute, Seoul National University, Seoul 110-460, ¹Korea Ocean Research and Development Institute, Ansan 425-600 and ²College of Natural Sciences, Sangji University, Wonju 220-702, Korea

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Two furofuran lignans, sesamolin and sesangolin were isolated from the seeds of *Sesamum indicum* and *S. angolense*, respectively. Detailed analysis of the ¹H- and ¹³C-NMR spectra of these lignans was carried out by the application of two-dimensional ¹H-¹H COSY and ¹H-¹³C multiple-bond, multiple-quantum spectroscopic correlation techniques.

Key words: Sesamum indicum, S. angolense, Pedaliaceae, furofuran lignans, NMR assignments, ¹H-¹H COSY, HMQC, HMBC

INTRODUCTION

The cultivated oilseed crop, Sesamum indicum L. (Pedaliaceae) is known to contain variable amounts of the furofuran lignans sesamin and sesamolin in its seed oil (Ryu et al., 1992; Kamal-Eldin and Appelqvist, 1994). Another related lignan sesangolin was isolated from the wild type sesame plant, S. an-

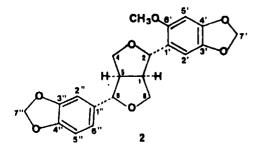


Fig. 1. Structures of sesamolin (1) and sesangolin (2).

Correspondence to: Sam Sik Kang, Natural Products Research Institute, Seoul National university, Seoul 110-460, Korea golense Welw. (Jones *et al.*, 1962; Ryu *et al.*, 1993). These sesamin type lignans are known for their strong antioxidative activities (Osawa, 1992). The structures of these lignans are well established (Haslam, 1970; Jones *et al.*, 1962) as shown in Fig. 1, but the ¹³C-NMR assignments for sesamolin (1) and sesangolin (2) are not fully characterized so far. In this paper, we report the NMR assignments of these compounds which were made with the aid of 2D correlation NMR techniques such as HMQC and HMBC.

MATERIALS AND METHODS

The general experimental methods are the same as those reported in Kang *et al.* (1994). Sesamolin (1) and sesangolin (2) were isolated from the seeds of *S. indicum* and *S. angolense*, respectively, as described in Ryu *et al.* (1992; 1993) and identified by direct comparison with authentic samples (mmp, UV, IR, NMR and MS).

RESULTS AND DISCUSSION

The 1 H-NMR chemical shifts of **1** was previously reported (Haslam, 1970) which were indistinguishable from those of our results as tabulated in Table I. The proton chemical shifts of **2** could be assigned in detail compared to data reported for similar and related compounds (Pelter *et al.*, 1976; Nagata *et al.*, 1987) as well as based on the 1 H- 1 H COSY spectra. It showed a multiple signal assigned to one methine proton at δ 2.94 (H-1), which was found to be coupled with a benzylic proton at δ 5.04 (d, H-2) and also with a pair of methylene protons at δ 4.30 and 4.01 (H-8) through 1 H- 1 H COSY-45 and HMQC experiments. In

Table I. Chemical shifts and correlations of sesamolin (1) in CDCl₃

Position	δ_{H}	δ _C	HMBC
C-1	3.30 (q, 8.5)	53.2	
C-2	5.49 (s)	106.8	C4, C5, C8
C-4	4.12 (dd, 8.5, 9.1)	69.7	C1, C2, C6
	3.96 (d, 9.1)		
C-5	2.94 (q, 8.5)	52. <i>7</i>	
C-6	4.39 (d, 8.5)	87.0	
C-8	4.43 (t, 9.0)	71.2	C2, C6
	3.63 (dd, 8.5, 9.0)		
C-1'		134.3	
C-21	6.87 (d, 1.2)	106.5	C4', C6'
C-3'		148.0	
C-4'		147.3	
C-5'	6.78 (d, 7.8)	108.1	C1', C3'
C-6'	6.82 (dd, 1.2, 7.8)	119.6	C21, C41
C-71	5.91	101.2	C3'
C-1"		151.8	
C-2"	6.62 (d, 2.5)	100.1	C4"
C-3"		148.1	
C-4"		142.6	
C-5"	6.70 (d, 8.5)	108.0	C1", C3"
C-6"	6.50 (dd, 2.5, 8.5)	108. 9	C2", C4"
C-7"	5.95 (s)	101.0	C3"

Coupling constants (J in Hz) in parentheses.

a similar manner, another multiplet proton at δ 2.93 (H-5) was coupled with another pair of methylene protons at δ 4.20 and 3.91 (H-4), and another benzylic proton at δ 4.66 (H-6). These signals supported the presence of unsymatrically substituted 2,6-diaryl cis-3,7-dioxabicyclo[3.3.0]octane in 2 (Pelter et al., 1976). The proton signals attributable to the 3,4methylenedioxy-6-methoxyphenyl and piperonyl (3,4methylenedioxyphenyl) moieties in 2 were assigned as follows. The two singlet signals at δ 6.90 and 6.52 ppm were shown to be correlated with methylenedioxy doublet protons (J=1.5 Hz) at δ 5.90 and 5. 91 ppm, which permitted that these protons were discernible from those for 3,4-methylenedioxyphenyl residue in 2. The presence of long-range couplings between H-2 and one of singlet signals at δ 6.90, and between a methoxy signal at δ 3.77 and a singlet signal at δ 6.52 ppm allowed the assignments of the signals at δ 6.90 and 6.52 ppm ascribable to H-2' and H-5', respectively in 3,4-methylenedioxy-6-methoxyphenyl moiety in 2. The assignments of protons in piperonyl moiety were easily made with the aid of the coupling constants. Two doublets for one of each oand m-coupled proton at δ 6.77 (J=7.6 Hz) for H-5" and δ 6.86 (J=1.5 Hz) for H-2", respectively, and double doublets at δ 6.81 (J=1.5, 7.6 Hz) for H-6" were observed. The H-2" and H-6" protons were longrange coupled with H-6 proton at δ 4.66 ppm. On the basis of the above results, the proton chemical shifts for 2 were assigned as shown in Table II.

¹³C-NMR spectroscopy has proved to be a valuable

Table II. Chemical shifts and correlations of sesangolin (2) in CDCl₃

III CDCI3			
Position	δ_{H}	$\delta_{\scriptscriptstyle C}$	НМВС
C-1	2.94 (m)	54.6	
C-2	5.04 (d, 4.2)	82.0	C4, C8, C1', C2'
C-4	4.20 (dd, 6.1, 8.8)	71.4	C1, C2, C6
	3.91 (dd, 3.4, 8.8)		
C-5	2.93 (m)	54.3	
C-6	4.66 (d, 5.2)	85.3	C4, C5", C6"
C-8	4.30 (dd, 7.1, 9.0)	73.1	C2, C5, C6
	4.01 (dd, 3.8, 9.0)		
C-1'		123.1	
C-21	6.90 (s)	105.8	C4', C6'
C-31		141.1	
C-4'		147.1	
C-5'	6.52 (s)	94.5	C1', C3', C4'
C-6 ¹		151.4	
C-7'	5.91 (d, 1.5)	101.0*	C3', C4'
	5.90 (d, 1.5)		
OCH_3	3.77 (s)	56.2	C6'
C-1"		135.3	
C-2"	6.86 (d, 1.5)	106.5	C4", C6"
C-3"		147.9	
C-4"		147.1	
C-5"	6.77 (d, 7.6)	108.1	C1", C3"
C-6"	6.81 (dd, 1.5, 7.6)	119.4	C2", C4"
C-7"	5.94 (s)	101.1*	C3"

^{*}Assignment may be reversed. Coupling constants (J in Hz) in parentheses.

technique for solving the problems of structure and stereochemistry in the furofuran lignans. Thus, the aromatic resonances C-1' and C-1" are dependant on the configuration, as well as on the nature of the substituents on the aryl ring, but carbons 1, 2, 8, 4, 5 and 6 are affected by the stereochemistry only (Pelter et al., 1976; Agrawal and Thakur, 1985). The 13C chemical shifts of furofuran ring in 2 except for C-2 and 8 agreed well with previously reported data (Agrawal and Thakur, 1985; Ina et al., 1987; Mivazawa et al., 1994), but C-2 and 8 chemical shifts were resonated at δ 82.0 and 73.1 ppm, respectively, as can be seen in sesaminol glucosides (Katsuzaki et al., 1994). However, the ¹³C chemical shifts of furofuran ring in 1 were somewhat different from those of 2,6-diaryl furofuran lignans due to aryloxy substituent at C-2 position, but assignment of each carbon could be determined on the basis of HMQC and HMBC experiments. The carbon chemical shifts for piperonyl moieties in 1 and 2 could be straightforwardly assigned with the aid of DEPT and HMQC experiments as well as comparing the literature data for sesamin analogues. The resonances for C-1' in 1 and C-1" in 2 at δ 134.3 and 135.3, respectively follow the literature data and are associated with equatorial piperonyl group (Pelter et al., 1977). The chemical shifts corresponding to 3,4-methylenedioxyphenoxy residue in 1 and 3,4-methylenedioxy-6-methoxyphenyl moiety

in 2 could be assigned in a similar manner. The above assignments were further confirmed by HMBC experiments as indicated in Tables I and II.

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