

SPECT Imaging of Dopamine Transporter with $[^{123}\text{I}]\beta\text{-CIT}$: A Potential Clinical Tool in Parkinson's Disease

Sang Eun Kim, M.D.*[†], Won Yong Lee, M.D.[†], Dae Yoon Chi, Ph.D.*
Yeorn Seong Choe, Ph.D.*[‡], Kyung Han Lee, M.D.*[‡], Yong Choi, Ph.D.*
Seung Jun Oh, M.S.[†] and Byung-Tae Kim, M.D.*

Department of Nuclear Medicine*, Department of Neurology[†], Samsung Medical Center;
and Samsung Biomedical Research Institute[‡], Seoul, Korea

=국문초록 =

파킨슨병에서 $[^{123}\text{I}]\beta\text{-CIT}$ SPECT를 이용한 도파민 운반체 영상

삼성의료원 핵의학과*, 신경과[†]; 삼성생명과학연구소 임상의학연구센터[‡]

김상은*[†] · 이원용[†] · 지대운* · 최연성*
이경한* · 최 용* · 오승준[†] · 김병태*

$[^{123}\text{I}]\beta\text{-CIT}$ [$2\beta\text{-carbomethoxy-3}\beta\text{-(4-iodophenyl)tropane}$]는 도파민 운반체(dopamine transporter)에 특이결합하며 $[^{123}\text{I}]\beta\text{-CIT}$ 의 도파민 운반체 결합정도는 파킨슨병에서 도파민 뉴우런의 변성정도를 반영하는 것으로 제안되어 왔다. 이 연구의 주요 목적은 파킨슨병 환자에서 $[^{123}\text{I}]\beta\text{-CIT}$ SPECT를 이용하여 측정된 $[^{123}\text{I}]\beta\text{-CIT}$ 의 선조체 결합지표들이 질병의 임상적 진행정도를 반영하는지를 검토하고, 간편화된 조직방사능비가 $[^{123}\text{I}]\beta\text{-CIT}$ 의 결합정도를 나타내는 정량적 지표로 이용될 수 있는지를 검증하는 것이었다. 파킨슨병 환자 30명(59±9세, 평균±표준편차; Hoehn-Yahr stage 1-3)과 정상인 6명(58±5세)을 대상으로 $[^{123}\text{I}]\beta\text{-CIT}$ SPECT 영상을 얻었다. $[^{123}\text{I}]\beta\text{-CIT}$ 선조체 결합의 정량적 지표로서(선조체 방사능-소뇌방사능)/소뇌방사능 비(specific binding ratio, SBR)와 추적자역학모델을 이용하여 측정된 선조체 결합능(binding potential)(k_3/k_4)을 구하였다. 파킨슨병 환자에서 $[^{123}\text{I}]\beta\text{-CIT}$ 의 선조체 결합역학은 정상인에 비하여 현저하게 느렸으며 그 결합지표들은 정상인에 비하여 뚜렷하게 낮았다. 한편, 편측파킨슨병 환자에서 $[^{123}\text{I}]\beta\text{-CIT}$ 결합은 증상 반대쪽 선조체 뿐만 아니라 같은 쪽 선조체에서도 정상인에 비해 유의하게 감소되어 있었다. 파킨슨병 환자에서 $[^{123}\text{I}]\beta\text{-CIT}$ 투여 후 24시간의 선조체 SBR 및 최대 SBR, 선조체 결합능은 모두, 유병기간, Hoehn-Yahr stage, UPDRS(Unified Parkinson's Disease Rating Scale) 총점, UPDRS 운동점수, UPDRS 일상활동점수와 유의한 상관관계를 나타내었다. 24시간 선조체 SBR과 최대 SBR은 선조체 결합능과 우수한 상관관계를 보였다. 이상의 결과로부터 $[^{123}\text{I}]\beta\text{-CIT}$ 의 선조체 결합은 파킨슨병의 진행정도를 나타내는 지표로 이용될 수 있다. 또 $[^{123}\text{I}]\beta\text{-CIT}$ 투여 후 24시간 영상으로부터 얻은 간편화된 조직방사능 비는 $[^{123}\text{I}]\beta\text{-CIT}$ 의 결합정도를 정량적으로 반영한다. $[^{123}\text{I}]\beta\text{-CIT}$ SPECT는 파킨슨병의 조기진단 및 진행 추적에 임상적으로 유용할 것으로 판단된다.

Key Words : $[^{123}\text{I}]\beta\text{-CIT}$; Dopamine transporter; Parkinson's disease; SPECT

INTRODUCTION

Parkinson's disease(PD) is a common move-

ment disorder associated with degeneration of dopaminergic neurons in the substantia nigra and a corresponding loss of dopamine(DA)-containing nerve terminals in the basal ganglia^{1-5). Dege-}

neration of the nigrostriatal pathway is accompanied by large decreases in a number of corresponding biochemical markers, including DA^{1-3, 6-10}), dopa decarboxylase^{2, 11, 12}), tyrosine hydroxylase^{2, 10, 13}), DA metabolites^{1, 2, 6, 8, 9}), and the DA transporter^{7, 14-17}).

Noninvasive imaging and quantitation of the loss of dopaminergic nerve terminals in PD have evolved over the last decade by the use of 6-L-[¹⁸F]fluoro-DOPA([¹⁸F]FDOPA) and positron emission tomography(PET)¹⁸⁻²⁸). This method has permitted several studies of the reduction in DA synthesis in PD, its relationship to neurological parameters of disease^{22, 23, 25}), and comparison of PD with other movement disorders^{19, 23, 24, 27, 29}).

The DA reuptake site that mediates reuptake of DA into presynaptic nerve terminals following its release is an active, sodium gradient-driven membrane transporter spanning the plasma membrane of dopaminergic terminals. It is the function of the transporter to rapidly deplete the intrasynaptic DA during a DA surge and to maintain normal DA concentrations in the intra- and extracellular spaces at other times. Cocaine blocks the DA transporter, thus increasing the levels of intrasynaptic dopamine, which may account for the central nervous system stimulant actions of the drug³⁰⁻³²). Recently, a series of cocaine analogues, including CFT [2 β -carbomethoxy-3 β -(4-fluorophenyl)tropane; also designated WIN 35,428], β -CIT [2 β -carbomethoxy-3 β -(4-iodophenyl)tropane; also designated RTI-55] and isopropyl- β -CIT(also designated RTI-121), have been developed with high affinity for the DA transporter³³⁻⁴³), and some have been labeled with positron emitting and single-photon emitting isotopes to permit imaging by PET and single-photon emission computed tomography(SPECT)⁴²⁻⁴⁶).

In vitro binding study showed that β -CIT, the iodo analog of CFT, has a high affinity for the

DA and serotonin(5-hydroxytryptamine: 5-HT) transporters from baboon brain, with an IC₅₀ of 1.6 nM against [³H]CFT and 3.8 nM against [³H]paroxetine³⁷). ¹²³I-Labeled β -CIT has been used in SPECT imaging for visualization of binding to DA and 5-HT transporters in the baboon brain in vivo^{36, 43, 47}). In vivo displacement studies in the monkey demonstrated that striatal uptake of [¹²³I] β -CIT was primarily due to DA transporters while uptake in hypothalamus-midbrain areas was mainly associated with 5-HT transporters^{36, 37}). Initial studies in human subjects confirmed the high and prolonged levels of activity in striatum and demonstrated significant reductions of tracer uptake in patients with PD⁴⁷⁻⁴⁹). These studies suggest that [¹²³I] β -CIT SPECT imaging is a promising technique for the diagnosis and evaluation of PD.

Attempts to estimate receptor binding characteristics in vivo using PET and SPECT have followed a variety of approaches. One method is to assume a particular model in which a measured plasma input function governs the uptake of the labeled ligand. Model parameters can then be determined by a nonlinear least-square fit to the experimental data⁵⁰). Graphical method of analysis applicable to ligands that are trapped in tissue for the duration of the experiment have been developed and applied by several investigators⁵¹⁻⁵⁶). These model-based methods usually require arterial sampling and repeated scans, procedures that are not easily implemented in the clinical setting. The simplest procedure is using the ratio among different regions(i.e., receptor-rich versus receptor-poor region) or the slope of the ratio over scanning time, which is a measure of the specific binding or the rate of binding of the ligand in limited conditions⁵⁷).

The aim of the present study was: 1) to characterize the pharmacokinetics and regional distribution of [¹²³I] β -CIT in healthy subjects and

PD patients, 2) to evaluate the correlation between SPECT measures of [¹²³I]β-CIT binding and motor symptoms in patients with PD, and 3) to validate the use of simplified ratio method for the assessment of [¹²³I]β-CIT binding by comparing with a more complete tracer kinetic approach.

METHODS

1. Synthesis of [¹²³I]β-CIT

[¹²³I]β-CIT was prepared from the corresponding tributylstannyl precursor (Research Biochemicals International, Natick, MA) and high radionuclidic purity [¹²³I]NaI (Korea Atomic Energy Research Institute, Seoul, Korea), using the method described by Zea-Ponce et al. (1995) with minor modification. [¹²³I]β-CIT was obtained in an average radiochemical yield of 64% ± 12% (n=21, mean ± s.d.) and a radiochemical purity of 96% ± 4%. Although specific activity of the radiotracer could not be measured because of limit of UV detection with our HPLC system, it might be higher than 67,000 Ci/mmol based on the literature⁵⁸⁾.

2. Subjects

Thirty patients [13 males and 17 females; age 59 ± 9 yr (mean ± s.d.)] with idiopathic PD (Hoehn-Yahr stages 1-3) and 6 age-matched healthy controls (4 males and 2 females; age 58 ± 5 yr) were enrolled in the study following the provision of informed consent. All patients had symptoms that were responsive to L-dopa and had at least three of the following symptoms: resting tremor, bradykinesia, rigidity, and postural instability. Fourteen of the patients were recent-onset patients and were not receiving any dopaminergic medication before the SPECT scan. The rest of the patients were at an advanced stage of PD and were on treatment with L-dopa, DA agonist, L-deprenyl, amantadine and anticholinergic drugs

in various combinations. Each patient was evaluated at drug-off state using the Hoehn-Yahr stage and the Unified Parkinson's Disease Rating Scale (UPDRS)⁵⁹⁾. The clinical characteristics of the patients are summarized in Table 1. The healthy controls were taking no medications and were free of serious medical illnesses by physical examination and laboratory testing.

3. Data Acquisition

SPECT studies were performed using a three-headed Triad XLT system (Trionix Research Laboratory, Twinsburg, OH) equipped with medium-energy collimators. Images were acquired with each head rotating 120° in 3° steps, creating 120 raw image sets. Antiparkinsonian medications were discontinued for 2 days prior to the scanning. In order to minimize radioiodine uptake in the thyroid gland, each patient was given oral Lugol's solution, 1 drop tid, for 1 day prior and for 3 days after intravenous administration of [¹²³I]β-CIT. Fiducial markers containing ~7 μCi of ¹²³I were attached to the skin along the canthomeatal line for realignment of all images from each subject in a plane parallel to the canthomeatal line. Each subject received an intravenous bolus injection of 185–370 MBq [¹²³I]β-CIT. In all of the healthy controls and 14 of the patients, a total of 15 SPECT scans was obtained for each subject over a 24 hr period following injection: ten sequential scans of 10 min starting immediately after injection, followed by scans of 20–30 min at 3 hr, 4 hr, 6 hr, 12 hr, and 24 hr postinjection. Based on the time-activity curve from serial scans, the rest of the patients were scanned at 12 hr and 24 hr postinjection. Images were acquired with a 10% symmetric window centered at 159 keV, reconstructed with a Butterworth filter (power=7; cutoff=0.4 cyc/cm) and displayed in 128 × 128 matrix (pixel size = 3.56 × 3.56 mm with a slice thickness of 3.56 mm).

Table 1. Clinical Characteristics of Patients

Patient	Sex	Age (yr)	Disease duration(mo)	UPDRS					
				H-Y	Total	Motor	ADL	Motor subscales*, Right	Motor subscales*, Left
1	F	80	96	3	61	39	18	13	12
2	F	59	51	1.5	52	34	14	0	21
3	F	60	48	1	20	14	4	12	0
4	F	60	51	2	30	24	6	2	14
5	M	40	5	2	20	14	4	3	10
6	F	61	27	1	17	8	9	0	7
7	F	57	36	2	27	23	4	10	11
8	M	51	8	2	37	28	9	17	7
9	M	60	2	2	31	23	7	3	14
10	F	73	12	1.5	30	21	5	0	15
11	F	63	17	3	59	43	15	11	16
12	F	55	55	1.5	23	19	4	0	15
13	F	59	48	2.5	42	32	9	14	9
14	F	55	12	2	15	11	4	3	6
15	M	55	30	2.5	41	34	5	6	12
16	F	77	65	2.5	51	40	10	11	18
17	M	50	29	2	27	22	4	1	14
18	M	58	28	NA	NA	NA	NA	14	8
19	M	54	26	NA	NA	NA	NA	0	3
20	M	69	16	NA	NA	NA	NA	9	3
21	F	39	18	NA	NA	NA	NA	7	0
22	M	53	58	2.5	55	43	10	19	12
23	M	45	NA	NA	NA	NA	NA	NA	NA
24	M	57	10	2	36	30	6	17	5
25	F	63	24	NA	NA	NA	NA	NA	NA
26	F	58	42	2.5	49	35	12	11	11
27	F	56	24	2	50	38	9	18	11
28	M	71	13	2.5	47	35	10	16	7
29	F	57	48	3	65	49	14	22	10
30	M	69	12	2	31	23	7	16	3
Mean±s.d.		59±9	31.4±21.7	2.1±0.6	38.2±14.8	28.4±11.0	8.3±4.0	9.1±6.9	9.8±5.3

* Sum of lateralizing motor UPDRS subscales(tremor, rigidity, bradykinesia)

UPDRS=Unified Parkinson's Disease Rating Scale; H-Y=Hoehn-Yahr stage; ADL=activities of daily living; NA=data not available

Attenuation correction was performed using Chang's method($\mu=0.11/\text{cm}$)⁶⁰, and SPECT activity (cpm) was converted to absolute units of radioactivity(μCi) based upon a calibration factor determined from a cylindrical phantom of 20 cm diameter filled with an ¹²³I solution.

4. Data Analysis

Three consecutive slices with highest striatal activities were summed to construct a 10.68 mm thick slice, and standardized size and shape region of interest(ROI) (10.68 mm×10.68 mm rectangle) was visually positioned on each striatum. The same procedures were performed for

the ROI placement on hypothalamus-midbrain regions. Three consecutive slices representing the cerebellum were also added and ROI(14.24 mm x 24.92 mm) was placed on each cerebellar hemisphere. Right and left cerebellar values were averaged for subsequent analysis.

$[^{123}\text{I}]\beta\text{-CIT}$ binding in the striatum was estimated using two quantitative indices: 1) the simplified ratio of specific to nonspecific binding, or the radioactivity ratio of striatum(an area rich in DA transporter) to cerebellum [an area containing few or no DA transporter⁶¹⁾] minus 1 (specific binding ratio: SBR) and 2) the binding potential, or the ratio of the rate constant of binding to the DA transporter(k_3) to that of dissociation from the DA transporter(k_4), as calculated based on a kinetic two-compartment analysis of radioligand binding.

5. Kinetic Analysis of $[^{123}\text{I}]\beta\text{-CIT}$ Binding

By using a two-compartment kinetic model, the rate constants k_3 and k_4 were determined in the striata of healthy subjects and PD patients. Since

the number of DA transporter is negligible in the cerebellum, the assumption was made that the concentration and kinetics of $[^{123}\text{I}]\beta\text{-CIT}$ in the cerebellum is the same as in the free plus nonspecifically bound space in the striatum^{62, 63)}. Therefore, the time-activity curve of the cerebellum was used as an input function for the two-compartment model. The concentration of specifically bound radioligand in the striatum was determined by subtracting the radioactivity concentration in the cerebellum from that in the striatum. The optimum values of k_3 and k_4 were obtained using a nonlinear least square fitting procedure. The binding of $[^{123}\text{I}]\beta\text{-CIT}$ to the DA transporter in the striatum was evaluated using the binding potential, defined as k_3/k_4 ratio⁶⁴⁾.

6. Statistical Analysis

Results are expressed as the mean \pm s.d. Comparisons of the SPECT measures of $[^{123}\text{I}]\beta\text{-CIT}$ binding between patients and controls were made with the Mann-Whitney U-test; comparisons between contralateral and ipsilateral striatum were performed using the Wilcoxon signed rank test.

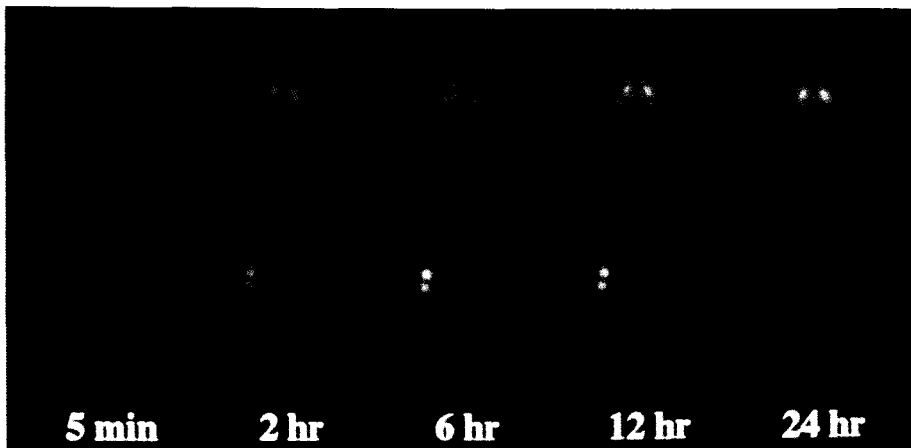


Fig. 1. Brain images obtained with $[^{123}\text{I}]\beta\text{-CIT}$ in a healthy subject. The images illustrate a plane of scanning through the basal ganglia and cerebellum. With increasing time after injection $[^{123}\text{I}]\beta\text{-CIT}$ concentrates highly in the striatum, an area rich in dopamine transporter; low activity is observed in the cerebral cortex and cerebellar regions, which contain few or no dopamine transporters.

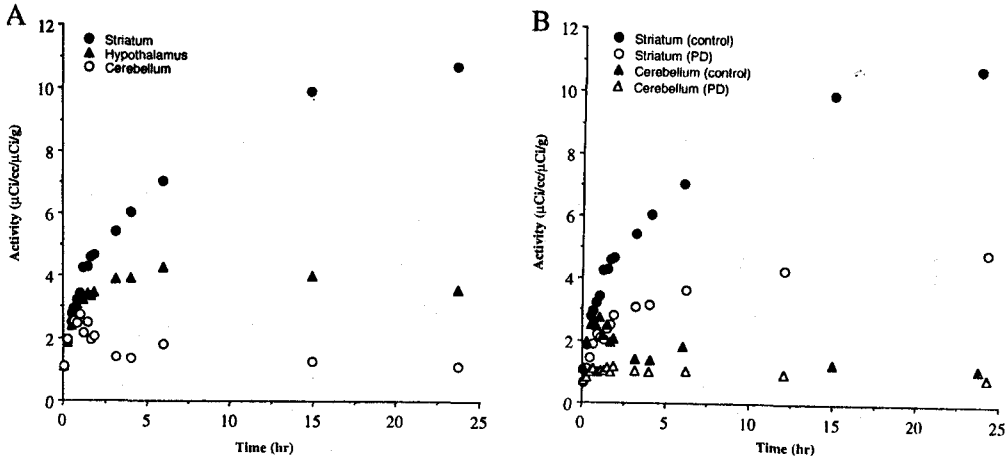


Fig. 2. Change in the level of activity in different regions of the brain with time in a healthy control (A) and in a patient with Parkinson's disease (PD) (B). Activity is expressed as $\mu\text{Ci/cc}/\mu\text{Ci}$ injected/g body weight.

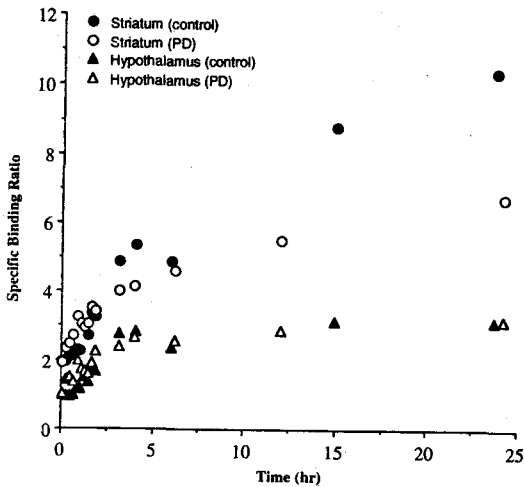


Fig. 3. Change of the specific binding ratio in the striatum and hypothalamus with time in a healthy control and in a patient with Parkinson's disease (PD).

Linear regression analysis by the least squares method was used to assess the relationship between the SPECT measures of $[^{123}\text{I}]\beta\text{-CIT}$ binding and between the SPECT measures and motor ratings (Hoehn-Yahr stage and UPDRS scores). Probability values of less than 0.05 were considered significant.

RESULTS

1. Pharmacokinetics of $[^{123}\text{I}]\beta\text{-CIT}$

With increasing time after injection $[^{123}\text{I}]\beta\text{-CIT}$ concentrated highly in the striatum, an area rich in DA transporter, followed by hypothalamus and midbrain, regions rich in 5-HT transporters (Fig. 1-3). Low activity was observed in the cerebral cortex and cerebellar regions, which contain few or no DA transporters. Fig. 2 and 3 show representative regional time-activity curves for the binding of $[^{123}\text{I}]\beta\text{-CIT}$. The striatal activity in all healthy subjects increased over time during the 24 hr scanning period. In the patients with PD, $[^{123}\text{I}]\beta\text{-CIT}$ accumulated more slowly and the peak striatal activity was clearly lower than in the healthy subjects. In the healthy subjects, the cerebellar activity reached a peak by 1 hr postinjection with a rapid washout thereafter; a similar uptake and washout was shown in the patients with PD. As a consequence, the striatal SBR in the healthy subjects increased steadily over time, while it peaked earlier at 12-24 hr postinjection and attained lower peak levels in the

Parkinsonian patients.

2. Striatal Binding of [¹²³I]β-CIT in PD

In the patients with PD, the binding of [¹²³I]β-CIT in the striatum was markedly reduced; a greater reduction occurred in the putamen than in

the caudate (Fig. 4). The results of SPECT measurement of [¹²³I]β-CIT binding in the Parkinsonian patients and healthy controls are shown in Table 2 and Fig. 5. The mean of right and left SBR at 24 hr postinjection and the mean peak SBR in the striatum were reduced to 48% and

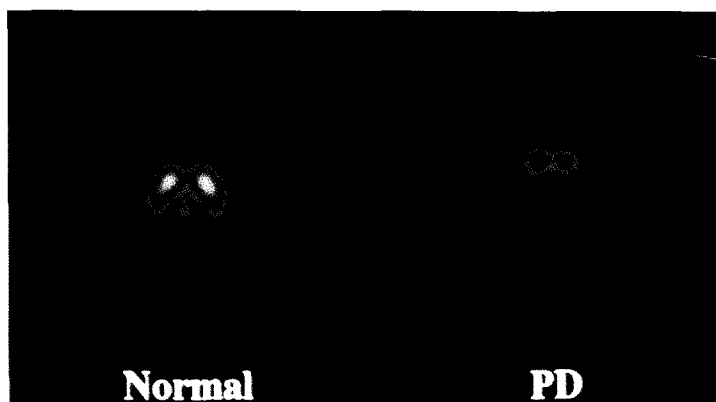


Fig. 4. [¹²³I]β-CIT SPECT images of a healthy subject and of a Parkinson's disease (PD) patient. In the patient with PD, [¹²³I]β-CIT binding in the striatum is markedly reduced; a greater reduction occurred in the putamen than in the caudate.

Table 2. [¹²³I]β-CIT SPECT Measures in Parkinson's Disease Patients and Healthy Controls

	Controls			PD patients			Symptomatic striatum*		
	24hr SBR [†]	Peak SBR [†]	BP [†]	24hr SBR [†]	Peak SBR [†]	BP [†]	24hr SBR [†]	Peak SBR [†]	BP [†]
Mean ± s.d.	8.0 ± 0.7	8.0 ± 0.7	7.9 ± 0.6	3.9 ± 1.3	4.0 ± 1.1	3.7 ± 1.1	3.7 ± 1.2	3.9 ± 1.1	3.6 ± 1.2
p	-	-	-	0.0001	0.0001	0.0005	<0.0001	<0.0001	0.0002
% of control mean	-	-	-	48%	50%	47%	46%	48%	46%

	Patients with hemiparkinsonism					
	Contralateral striatum			Ipsilateral striatum		
	24hr SBR	Peak SBR	BP	24hr SBR	Peak SBR	BP
Mean ± s.d.	3.4 ± 1.2	3.8 ± 1.0	3.7 ± 1.1	4.0 ± 1.2	4.4 ± 1.0	4.5 ± 0.8
p	0.0027	0.0039	0.0105	0.0027	0.0039	0.0105
% of control mean	42%	48%	47%	50%	56%	57%

* Striatum contralateral to symptoms

[†] Mean of right and left striatal values

SBR=specific binding ratio calculated as the specific(striatal minus cerebellar activity) to cerebellar activity ratio; BP=binding potential(k_3/k_4) measured using the two-compartment model; p=probability, Parkinson's disease patients vs controls, according to the Mann-Whitney U-test

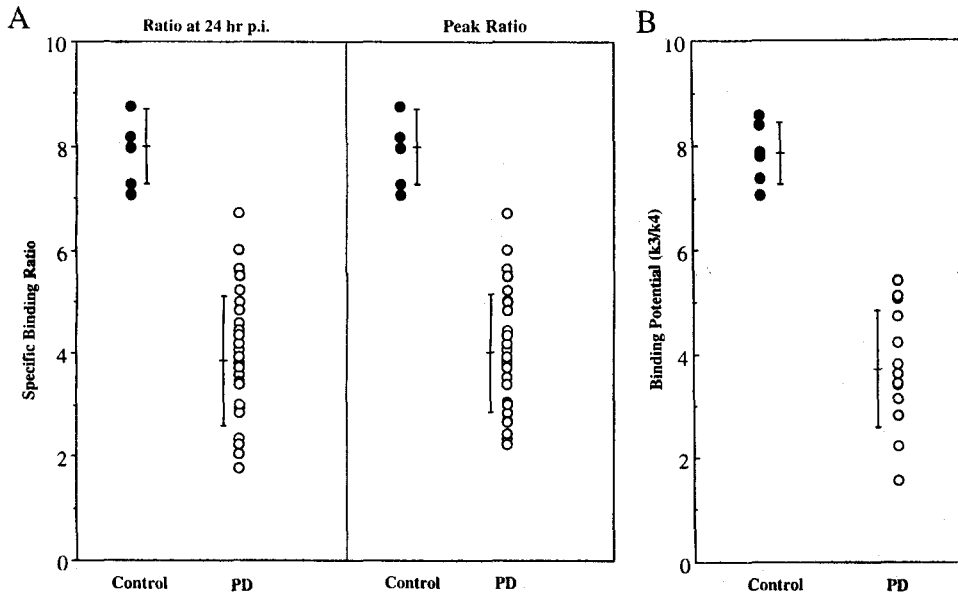


Fig. 5. Specific binding ratio(A) and binding potential(B) in healthy controls and Parkinson's disease (PD) patients.

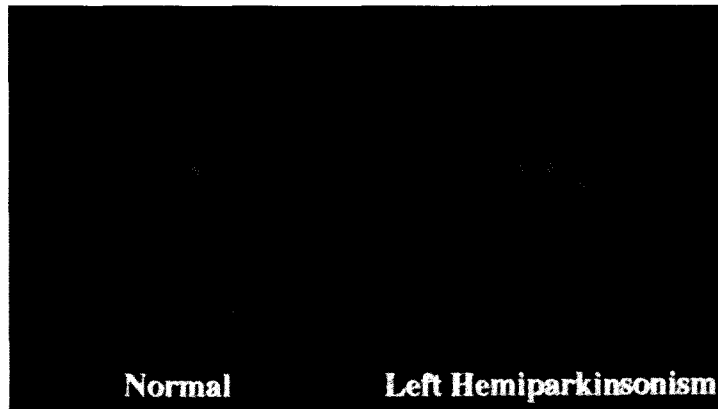


Fig. 6. [^{123}I] β -CIT SPECT images of a healthy subject and of a patient with hemiparkinsonism. In the patient with hemiparkinsonism, [^{123}I] β -CIT binding is reduced not only in the striatum contralateral to the clinical symptoms but also in the ipsilateral striatum. Note that left is on the right in the figure.

50%, respectively, of the control mean (3.9 ± 1.3 versus 8.0 ± 0.7 , $p=0.0001$; 4.0 ± 1.1 versus 8.0 ± 0.7 , $p=0.0001$, respectively). The mean binding potential in the striatum was also reduced to 47% of the control mean (3.7 ± 1.1 versus 7.9 ± 0.6 , $p=0.0005$). The SBR at 24 hr postinjection, the peak SBR

and the binding potential in the striatum corresponding to the clinical symptoms were reduced to 46%, 48%, and 46%, respectively, of the control mean (3.7 ± 1.2 versus 8.0 ± 0.7 , $p<0.0001$; 3.9 ± 1.1 versus 8.0 ± 0.7 , $p<0.0001$; 3.6 ± 1.2 versus 7.9 ± 0.6 , $p=0.0002$). In the patients with hemiparkinsonism,

Table 3. Correlation Coefficients for SPECT Measures and Motor Ratings in Parkinson's Disease Patients

	SBR at 24hr p.i.	Peak SBR	Binding potential
Disease duration(mo)	-0.372	-0.379	-0.572
p	0.0510	0.0466	0.0325
Hoehn-Yahr stage	-0.508	-0.564	-0.453
p	0.0133	0.0050	0.1042
Total UPDRS	-0.557	-0.591	-0.697
p	0.0058	0.0030	0.0056
Motor UPDRS	-0.542	-0.568	-0.681
p	0.0075	0.0047	0.0073
ADL score of UPDRS	-0.487	-0.543	-0.599
p	0.0183	0.0075	0.0235

Note: Correlations are for means of right and left striatal values.

p = probability according to linear regression analysis; SBR = specific binding ratio calculated as the specific(striatal minus cerebellar activity) to cerebellar activity ratio; UPDRS = Unified Parkinson's Disease Rating Scale; ADL = activities of daily living.

the 24 hr and peak SBR and the binding potential were reduced not only in the striatum contralateral to the clinical symptoms [42%(3.4±1.2 versus 8.0±0.7, p=0.0027), 48%(3.8±1.0 versus 8.0±0.7, p=0.0039), and 47%(3.7±1.1 versus 7.9±0.6, p=0.0105), respectively, of the control mean] but also in the ipsilateral striatum [50%(4.0±1.2 versus 8.0±0.7, p=0.0027), 56%(4.4±1.0 versus 8.0±0.7, p=0.0039), and 57%(4.5±0.8 versus 7.9±0.6, p=0.0105), respectively, of the control mean](also see Fig. 6). In the patients with hemiparkinsonism or predominantly unilateral symptoms, the reduction was greater on the side opposite to the predominant symptoms than on the ipsilateral side(24 hr SBR, 3.6±1.0 versus 4.3±1.1, p=0.0046; peak SBR, 3.9±0.8 versus 4.5±0.9, p=0.0047; binding potential, 3.5±0.8 versus 4.4±0.6, p=0.0180).

3. Correlation of SPECT Measures with Motor Symptoms

Table 3 shows the correlations of SPECT measures with motor ratings in PD patients. The mean SBR at 24 hr postinjection, the mean peak SBR and the mean binding potential in the striatum were significantly correlated with disease

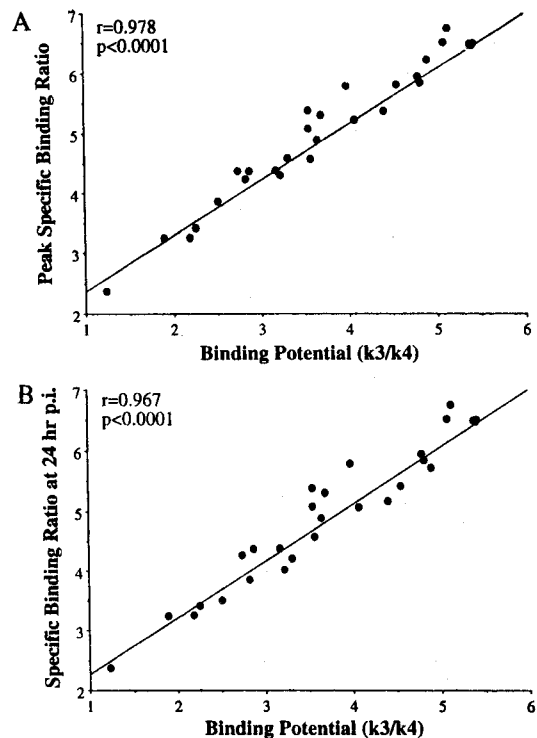


Fig. 7. Correlation of the peak specific binding ratio (A) and the specific binding ratio at 24 hr postinjection (B) with the binding potential.

duration, Hoehn-Yahr stage, total score of

UPDRS, motor score of UPDRS, and activities of daily living score of UPDRS.

4. Correlation between Simplified Ratio Index and Kinetic Parameters

There was an excellent correlation between the peak striatal SBR and the binding potential ($r=0.978$, $p<0.0001$)(Fig. 7). Correlation between the striatal SBR at 24 hr postinjection and the binding potential was also shown to be excellent ($r=0.967$, $p<0.0001$).

DISCUSSION

The present study demonstrates that SPECT measurement of [^{123}I] β -CIT binding clearly distinguishes patients with PD from healthy subjects. Between symptomatic patients and healthy subjects, the striatal SBR and binding potential values were not only significantly different but also showed a significant interval. The wide gap in the striatal [^{123}I] β -CIT binding between symptomatic patients and healthy subjects suggests that in vivo imaging may be able to identify patients before the development of definite clinical symptoms. Indeed, we found that in patients with hemiparkinsonism, the [^{123}I] β -CIT binding reduced not only in the striatum contralateral to the clinical symptoms but also in the ipsilateral striatum. The ability to identify biochemically patients with PD who have early symptoms or presymptomatic individuals at risk for PD might be helpful in light of recent studies suggesting that early treatment with L-deprenyl, a monoamine-oxidase B inhibitor, slows the progression of disability in PD⁶⁵⁻⁶⁷. Additionally, the measurement of [^{123}I] β -CIT binding in the striatum corresponding to the symptomatic or asymptomatic side may provide information regarding the threshold for dopaminergic terminal loss at which symptoms become clinically apparent.

The decrease of [^{123}I] β -CIT binding in the striatum contralateral to the clinical symptoms (approximately 50%) was not as great as the loss of endogenous DA and DA transporter reported in postmortem human tissue samples(>80%)^{3, 68}. This may be due to difference in the patient population: the patients in the present study (Hoehn-Yahr stages 1-3) appear much less severely affected than those from postmortem examinations. Although it has been proposed that Parkinsonian symptoms develop only after 85-90% depletion of endogenous DA levels, this imaging study suggests that symptoms may begin with only a 50% decrease in striatal DA terminal innervation.

We found a clear negative correlation between [^{123}I] β -CIT binding in the striatum and the disability of the patients, assessed by the Hoehn-Yahr stage and the UPDRS scores. This finding from [^{123}I] β -CIT and SPECT is in accordance with other SPECT or PET studies using the same ligand^{69, 70} or [^{18}F]FDOPA⁷¹. Also in vitro studies have indicated a correlation of the degree of hypokinesia and rigidity of PD patients with striatal DA deficiency¹ and nigral neuronal loss^{1, 4, 72, 73}.

The loss of midbrain DA in PD is accompanied by a rise in the DA D₁ and D₂ receptor densities^{17, 74}. This is found in the putamen and caudate tissues from unmedicated patients, and may account for the clinical supersensitivity to DA agonists in PD patients^{75, 76}. However, little is known about the DA transporter regulation in residual neurons following DA neuronal loss. The density of DA transporter on the plasma membrane of dopaminergic terminals is commonly believed to be so constant that the number of terminals can be inferred from the DA transporter density^{14, 77, 78}, but there is very little direct evidence to support this notion. Therefore, it is not entirely certain whether alterations of the

number of dopaminergic terminals can be revealed by measurements of the density of DA transporters in such diseases as Parkinson's. Although we found a linear relationship between [¹²³I]β-CIT binding and the motor symptoms of the patients, the patients in the present study were in relatively early stages of the disease. Studies of a large series of patients with a wide range of symptom severity may clarify this issue.

While the agonist-induced down-regulation of postsynaptic DA receptors is established^{17, 75}, the effect of long-term treatment with L-dopa on the DA transporter density has still to be determined. In particular, the duration of the L-dopa effect in Parkinson's patients taking high doses of levodopa on a daily basis and undergoing SPECT imaging with [¹²³I]β-CIT, should be investigated further. Following chronic treatment with L-deprenyl, Wiener et al.⁷⁹ found an up-regulation of the DA transporter *in vitro* in the mouse brain using [³H]mazindol, whereas Ursula et al. failed to find a significant change in *in vivo* [³H]CFT accumulation in the mouse striatum (unpublished observation). It remains possible that at least some of the present findings can be related to the long-term dopaminergic treatment of the patients.

In the present study, antiparkinsonian medications were discontinued for 2 days prior to the scanning. Competition by endogenous DA has previously been reported for the binding of the radioligand [¹¹C]raclopride to postsynaptic DA D₂ receptors, and the consequences of these findings on the interpretation of PET studies have been discussed⁸⁰⁻⁸². Since the affinity of β-CIT for DA transporter is approximately 3 orders of magnitude greater than that of DA, it appears unlikely that the binding of this ligand is influenced by modest fluctuations in intrasynaptic DA levels. With large doses of L-dopa(50-200 mg/kg) *in vivo* microdialysis studies in normal rat striata

showed DA concentrations to be rising either not at all⁸³ or only 2 to 4 times above baseline levels⁸⁴⁻⁸⁷. Infusion of L-dopa(50 mg/kg) failed to displace striatal [¹²³I]β-CIT binding in nonhuman primates⁸⁸, suggesting that the binding would not be affected by L-dopa administration in Parkinsonian patients. However, it has been shown that in 6-hydroxydopamine lesioned rats, L-dopa infusion(100 mg/kg) increases striatal extracellular DA 30-fold, compared with less than 2-fold in normal striatum⁸⁹. This difference has been attributed to reduced buffering capacity in denervated striatum as a result of loss of DA terminals. Supporting this notion, Antonini et al.⁹⁰ found that several hours of continuous L-dopa infusion(60-80 mg/hr) reduced [¹¹C]raclopride binding in the putamen by 20%-27% but not in the caudate which is less severely affected than the putamen in PD. In Parkinson's patients, therefore, L-dopa therapy may have to be temporarily interrupted to avoid its potential interference with the binding of [¹²³I]β-CIT to DA transporter. In addition, L-deprenyl and its major metabolite, L-methamphetamine⁹¹, enhance intrasynaptic DA levels⁹²⁻⁹⁴. However, it is presently unknown whether acute administration of L-deprenyl affects [¹²³I]β-CIT binding.

For [¹²³I]β-CIT SPECT to be easily applicable in the clinical setting, relatively simple methods of quantification will be required. In the present study, we found an excellent correlation between the simplified tissue ratio obtained either at peak striatal binding or 24 hr postinjection and the binding potential from kinetic analysis. This finding indicates that the simplified ratio index obtained at 24 hr postinjection may be feasible for the assessment of [¹²³I]β-CIT binding. The combination of SPECT camera availability with a simple accurate method that does not require repeated scanning or complicated modeling procedures would certainly increase the clinical use

of [123 I] β -CIT SPECT in the diagnosis and treatment of PD.

The diagnosis of PD remains a clinical judgment based primarily upon motor examination and the patient's response to L-dopa. A prior study correlating clinical impression with subsequent pathology showed only 75-80% agreement between clinical and pathological diagnoses⁹⁵⁾. Moreover, evaluation of disease progression based on clinical examination is complicated by the drug therapy of motor symptoms. Hence, an objective marker of DA neuronal loss is essential for the diagnosis and serial monitoring of the disease and for improved understanding of the pathophysiology of disease onset and progression. The results of the present study demonstrate marked differences in [123 I] β -CIT SPECT measures between healthy subjects and PD patients. The significant correlation of SPECT measures with motor severity suggests that [123 I] β -CIT may be a useful marker of disease severity in PD. Additionally, the simplified tissue ratio obtained at 24 hr postinjection may be feasible for the assessment of [123 I] β -CIT binding, avoiding repeated scanning and complicated modeling procedures. [123 I] β -CIT SPECT may be clinically useful for the early diagnosis and serial monitoring of PD.

ACKNOWLEDGMENT

The authors thank Korea Atomic Energy Research Institute, Seoul, Korea, for supplying the high radionuclidic purity [123 I]NaI solutions, Jae Dong Seok and Kyu Bok Lee for their technical assistance, and Yeon Sook Yoon for her assistance in preparing the manuscript. This work was supported in part by Samsung Biomedical Research Institute grant C-95-025.

REFERENCES

- 1) Bernheimer H, Birkmayer W, Hornykiewicz O, et al.: *Brain dopamine and the syndromes of Parkinson and Huntington. Clinical, morphological and neurochemical correlations. J Neurol Sci* 1973;20:415-455
- 2) Hornykiewicz O, Kish SJ: *Biochemical pathophysiology of Parkinson's disease. Adv Neurol* 1986; 45:19-34
- 3) Kish SJ, Shannar K, Hornykiewicz I: *Uneven pattern of dopamine loss in the striatum of patients with idiopathic Parkinson's disease. Pathophysiologic and clinical implications. N Engl J Med* 1988;318:876-880
- 4) Rinne JO, Rummukainen J, Paljarvi L, Rinne UK: *Dementia in Parkinson's disease is related to neuronal loss in the medial substantia nigra. Ann Neurol* 1989;26:47-50
- 5) German DC, Manaye K, Smith WK, et al.: *Midbrain dopaminergic cell loss in Parkinson's disease: Computer visualization. Ann Neurol* 1989;26:507-514
- 6) Scatton B, Rouquier L, Javoy-Agid F, Agid Y: *Dopamine deficiency in the cerebral cortex in Parkinson disease. Neurology* 1982;32:1039-1040
- 7) Pimoule C, Schoemaker H, Jovoy-Agid F, et al.: *Decrease in [3 H]cocaine binding to the dopamine transporter in Parkinson's disease. Eur J Pharmacol* 1983;95:145-146
- 8) Bokobza B, Ruberg M, Scatton B, et al.: [3 H] spiperone binding, dopamine and HVA concentration in Parkinson's disease and supranuclear palsy. *Eur J Pharmacol* 1984;99:167-175
- 9) Perry TL, Wright JM, Berry K, et al.: *Dominantly inherited apathy, central hypoventilation, and Parkinson's syndrome: clinical, biochemical, and neuropathologic studies of 2 new cases. Neurology* 1990;40:1882-1887
- 10) Dubois B, Pillon B: *Biochemical correlates of cognitive changes and dementia in Parkinson's Disease. In: Huber J, Cummings JL, eds. Parkinson's disease: Neurobehavioral aspects New York: Oxford University Press. 1992:178-198*
- 11) Lloyd KG, Hornykiewicz O: *Parkinson's disease activity of L-dopa decarboxylase in discrete brain regions. Science* 1970;170:1212-1213
- 12) Nagatsu T, Kato T, Nagatsu I, et al.: *Cacholamine-related enzymes in the brain of patients*

- with Parkinsonism and Wilson's disease. *Adv Neurol* 1979;24:283-292
- 13) Goto S, Hirano A, Matsumoto S: Subdivision involvement of nigrostriatal loop in idiopathic Parkinson's disease and striatonigral degeneration. *Ann Neurol* 1989;26:766-770
 - 14) Janowsky A, Vocci F, Berger P, et al.(1987): [³H]GBR-12935 binding to the dopamine transporter is decreased in the caudate nucleus in Parkinson's disease. *J Neurochem* 49:617-621
 - 15) Maloteaux J-M, Vanisberg M-A, Laterre C, et al.: [³H]GBR 12935 binding to dopamine uptake sites: Subcellular localization and reduction in Parkinson's disease and progressive supranuclear palsy. *Eur J Pharmacol* 1988;156:331-340
 - 16) Hirai M, Kitamura N, Hashimoto T, et al.: [³H]GBR-12935 binding sites in human striatal membranes: Binding characteristics and changes in Parkinsonians and schizophrenics. *Jpn J Pharmacol* 1988;47:237-243
 - 17) Pearce RKB, Seeman P, Jellinger K, Tourtellotte WW: Dopamine uptake sites and dopamine receptors in Parkinson's disease and schizophrenia. *Eur Neurol* 1990;30(Suppl 1):9-14
 - 18) Garnett ES, Nahmias C, Firnau G: Central dopaminergic pathways in hemiparkinsonism examined by positron emission tomography. *Can J Neurol Sci* 1984;1:174-179
 - 19) Leenders KL, Frackowiak RSJ, Quinn N, et al.: Ipsilateral blepharospasm and contralateral hemidystonia and parkinsonism in a patient with a unilateral rostral brainstem-thalamic lesion: Structural and functional abnormalities studied with CT, MRI and PET scanning. *Movement Dis* 1986;1:51-58
 - 20) Leenders KL, Salmon EP, Tyrrell P, et al.: Brain L-(¹⁸F)-6-fluorodopa and (¹¹C)-nomifensine uptake in patients with Parkinson's disease and healthy volunteers. *J Cereb Blood Flow Metab* 1989;9(Suppl 1):S723
 - 21) Martin WRW, Palmer MR, Patlak CS, Calne DB: Nigrostriatal function in humans studied with positron emission tomography. *Ann Neurol* 1989; 26:535-542
 - 22) Lang AE, Garnett ES: Dopa-responsive parkinsonism with normal 6[¹⁸F]-fluorodopa positron emission tomography scans. *Ann Neurol* 1990; 28:592-593
 - 23) Brooks DJ, Ibanez V, Sawle GV, et al.: Differing patterns of striatal ¹⁸F-dopa uptake in Parkinson's disease, multiple system atrophy, and progressive supranuclear palsy. *Ann Neurol* 1990;28:547-555
 - 24) Brooks DJ, Salmon EP, Mathias CJ, et al.: The relationship between locomotor disability, autonomic dysfunction, and the integrity of the striatal dopaminergic system in patients with multiple system atrophy, pure autonomic failure, and Parkinson's disease, studied with PET. *Brain* 1990;113:1539-1552
 - 25) Leenders KL, Salmon EP, Tyrrell P, et al.: The nigrostriatal dopaminergic system assessed in vivo by positron emission tomography in healthy volunteer subjects and patients with Parkinson's disease. *Arch Neurol* 1990;47:1290-1298
 - 26) Sawle GV, Colebarch JG, Shah A, et al.: Striatal function in normal aging: Implications for Parkinson's disease. *Ann Neurol* 1990;28:799-804
 - 27) Sawle GV, Leenders KL, Brooks DJ, et al.: Dopa-responsive dystonia: [¹⁸F]dopa positron emission tomography. *Ann Neurol* 1991;30:24-30
 - 28) Sawle GV, Bloomfield PM, Bjorklund A, et al.: Transplantation of fetal dopamine neurons in Parkinson's disease: PET [¹⁸F]6-L-fluorodopa studies in two patients with putaminal implants. *Ann Neurol* 1992;31:166-173
 - 29) Brooks DJ, Playford ED, Ibanez V, et al.: Isolated tremor and disruption of the nigrostriatal dopaminergic system: an ¹⁸F-dopa PET study. *Neurology* 1992;42:1554-1560
 - 30) Church WH, Justice JB, Byrd LD: Extracellular dopamine in rat striatum following uptake inhibition by cocaine, nomifensine and bztropine. *Eur J Pharmacol* 1987;139:343-348
 - 31) Hurd YL, Ungerstedt U Cocaine: An in vivo microdialysis evaluation of its acute action on dopamine transmission in rat striatum. *Synapse* 1989;3:48-54
 - 32) Kuhar MJ, Ritz MC, Boja JW: The dopamine hypothesis of the reinforcing properties of cocaine. *TINS* 1991;14:299-302
 - 33) Boja JW, Carroll FI, Rahman MA, et al.: New, potent cocaine analogs: Ligand binding and transport studies in rat striatum. *Eur J Pharmacol* 1990;184:329-332
 - 34) Boja JW, Patel A, Carroll FI, et al.: [¹²⁵I]RTI-55: A potent ligand for dopamine transporters. *Eur J Pharmacol* 1991;194:133-134
 - 35) Carroll FI, Rahman MA, Abraham P, Parham K, Lewin AH, Dannals RF, Shaya E, Scheffel U, Wong DF, Boja JW, Kuhar MJ: [¹²³I]3β-carboxylic acids methyl ester (RTI-55), a unique

- cocaine receptor ligand for imaging the dopamine and serotonin transporters in vivo. *Med Chem Res* 1991;1:289-294
- 36) Innis R, Baldwin R, Sybirska E, Zea Y, Laruelle M, Al-Tikriti M, Charney D, Zoghbi S, Wisniewski G, Hoffer P, Wang S, Millius R, Neumeyer J: Single photon emission computer tomography imaging of monoamine reuptake sites in primate brain with [¹²³I] CIT. *Eur J Pharm* 1991;200:369-370
- 37) Neumeyer JL, Wang SY, Milius RA, et al.: [¹²³I]-2β-carbomethoxy-3β-(4-iodophenyl)tropane: High affinity SPECT radiotracer of monoamine reuptake sites in brain. *J Med Chem* 1991;34:3144-3146
- 38) Boja JW, Cline EJ, Carroll FI, et al.: High potency cocaine analogs: Neurochemical, imaging, and behavioral studies. *Ann NY Acad Sci* 1992;654:282-291
- 39) Boja JW, Mitchell WM, Patel A, et al.: High affinity binding of [¹²⁵I]RTI-55 to dopamine and serotonin transporters in rat brain. *Synapse* 1992;12:27-36
- 40) Carroll FI, Abraham P, Lewin AH, Parham KA, Boja JW, Kuhar MJ: Isopropyl and phenyl esters of 3ν(4-substituted phenyl) tropan-2νcarboxylic acids. Potent and selective compounds for the dopamine transporter. *J Med Chem* 1992;35:2497-2500
- 41) Cline EJ, Scheffel U, Boja JW, et al.: In vivo binding of [¹²⁵I]RTI-55 to dopamine transporters: Pharmacology and regional distribution with autoradiography. *Synapse*. 1992;12:37-46
- 42) Scheffel C, Dannals RF, Wong DF, et al.: Dopamine transporter imaging with novel, selective cocaine analogs. *Neurol Rep* 1992;3:969-972
- 43) Shaya EK, Scheffel U, Dannals RF, Ricaurte GA, Carroll FI, Wagner HN Jr, Kuhar MJ, Wong DF: In vivo imaging of dopamine reuptake sites in the primate brain using single photon emission computed tomography (SPECT) and iodine-123 labeled RTI-55. *Synapse* 1992;10:169-172
- 44) Dannals RF, Neumeyer JL, Milius RA, et al.: Synthesis of a radiotracer for studying dopamine uptake sites in vivo using PET. 2β-carbomethoxy-3β-(4-fluorophenyl)-[N-¹¹C-methyl]-tropane [¹¹C]CFT or [¹¹C]WIN-35,428). *J Labelled Compd Radiopharm* 1993;33:147-152
- 45) Wong DF, Yung B, Dannals RF, Shaya EK, Ravert HT, Chen CA, Chan B, Folio T, Scheffel U, Kuhar MJ: In vivo imaging of baboon and human dopamine transporters by positron emission tomography using [¹¹C]WIN 35,428. *Synapse* 1993;15:130-142
- 46) Fowler JS, Volkow ND, Wolf AP, et al.: Mapping cocaine binding sites in human and baboon brain in vivo. *Synapse* 1993;4:371-377
- 47) Brücke T, Kornhuber J, Angelberger P, Asenbaum S, Frassine H, Podreka I: SPECT imaging of dopamine and serotonin transporters with [¹²³I]β-CIT. Binding kinetics in the human brain. *J Neural Transm* 1993;94:137-146
- 48) Innis RB, Seibyl JP, Scanley BE, Laruelle M, Abi-Dargham A, Wallace E, Zea-Ponce Y, Zoghbi S, Wang S, Gao Y, Neumayer JL, Charney DS, Hoffer PB, Marek KL: SPECT imaging demonstrates loss of striatal transporters in Parkinson's disease. *Proc Natl Acad Sci USA* 1993;90:11,965-11, 969
- 49) Kuikka JT, Bergström KA, Vannien E, Laulumaa V, Hartikainen P, Länsimies E: Initial experience with single-photon emission tomography using iodine-123-labelled 2β-carbomethoxy-3β-(4-iodophenyl)tropane in human brain. *Eur J Nucl Med* 1993;20:783-786
- 50) Carson RE: Parameter estimation in positron emission tomography. In: Phelps M, Mazziotta J, Schelbert H, eds. *Positron emission tomography and autoradiography: Principles and applications for the brain and heart*. New York: Raven Press 1986:347-390
- 51) Gjedde A: High- and low-affinity transport of D-glucose from blood to brain. *J Neurochem* 1981;36:1463-1471
- 52) Gjedde A, Wong DF, Wagner HN Jr: Transient analysis of irreversible and reversible tracer binding in human brain in vivo. In: Battistin L, ed. *PET and NMR: New perspectives in neuroimaging and in clinical neurochemistry*. New York: Alan R Liss; 1986:223-235
- 53) Patlak CS, Blasberg RG, Fenstermacher JD: Graphical evaluation of blood-to-brain transfer constants from multiple-time uptake data. *J Cereb Blood Flow Metab* 1983;3:1-7
- 54) Patlak CS, Blasberg RG: Graphical evaluation of blood-to-brain transfer constants from multiple-time uptake data. Generalizations. *J Cereb Blood Flow Metab* 1985;5:584-590
- 55) Wong DF, Gjedde A, Wagner HN Jr: Quantification of neuroreceptors in the living human brain. I. Irreversible binding of ligands. *J Cereb Blood Flow Metab* 1986;6:137-146

- 56) Wong DF, Gjedde A, Wagner HN Jr, et al.: Quantification of neuroreceptors in the living human brain. II. Inhibition studies of receptor density and affinity. *J Cereb Blood Flow Metab* 1986;6:147-153
- 57) Wong DF, Wagner HN Jr, Dannals RF, et al.: Effects of age on dopamine and serotonin receptors measured by positron tomography in the living human brain. *Science* 1984;226:1393-1396
- 58) Zea-Ponce Y, Baldwin RM, Laruelle M, Wang S, Neumeyer JL, Innis RB: Simplified multidose preparation of iodine-123-β-CIT: a marker for dopamine transporters. *J Nucl Med* 1995;36:525-529
- 59) Fahn S, Elton R: Members of the UPDRS Development Committee. Unified Parkinson's disease rating scale. In: Fahn S, Marsden CD, Calne DB, Goldstein M, eds. *Recent developments in Parkinson's disease*. Florham Park, NJ: Macmillan Healthcare Information 1987:153-164
- 60) Chang L: A method for attenuation correction in computed tomography. *IEEE Trans Nucl Sci* 1987;NS-25:638-643
- 61) De Keyser J, De Baecker JP, Ebinger G, Vauquelin G: [³H]GBR 12935 binding to dopamine uptake sites in the human brain. *J Neurochem* 1989;53:1400-1404
- 62) Farde L, Hall H, Ehrin E, Sedvall G: Quantitative analysis of D2 dopamine receptor binding in the living human brain by PET. *Science* 1986;231:258-261
- 63) Eckernäs SÅ, Aquilonius SM, Hartvig P, Häggglund J, Lunqvist H, Nägren K, Långström B: Positron emission tomography (PET) in the study of dopamine receptors in the primate brain: Evaluation of a kinetic model using ¹¹C-N-Methyl-spiperone. *Acta Neurol Scand* 1987;75:168-178
- 64) Mintun MA, Raichle ME, Kilbourn MR, Wooten GF, Welch MJ: A quantitative model for the in vivo assessment of drug binding sites with positron emission tomography. *Ann Neurol* 1984;15:217-227
- 65) Tetrad JW, Langston JW: The effect of deprenyl (selegiline) on the natural history of Parkinson's disease. *Science* 1989;245:519-522
- 66) The Parkinson Study Group: Effect of deprenyl on the progression of disability in early Parkinson's disease. *N Engl J Med* 1989;321:1364-1371
- 67) The Parkinson Study Group: Effects of toco-pherol and deprenyl on the progression in early Parkinson's disease. *N Engl J Med* 1993;328:176-183
- 68) Kaufman MJ, Madras BK: Severe depletion of cocaine recognition sites associated with the dopamine transporter in Parkinson's disease striatum. *Synapse* 1991;49:43-49
- 69) Brücke T, Asenbaum S, Pozzera A: Hornykiewicz S, Haraskovan der Meer, Wenger S, Koch G, Pirker W, Wöber Cj, Müller Ch, Kornhuber J, Angelberger P, Podreka I. Dopaminergic nerve cell loss in Parkinson's disease quantified with [¹²³I]-2β-CIT and SPECT correlates with clinical findings. *Mov Disord* 1994;9(Suppl 1):120
- 70) Seibyl JP, Marek KL, Quinlan D, et al.: Decreased single-photon emission computed tomographic [¹²³I]β-CIT striatal uptake correlates with symptom severity in Parkinson's disease. *Ann Neurol* 1995;38:589-598
- 71) Brooks DJ, Salmon EP, Mathias CJ, Quinn N, Leenders KL, Bannister R, Marsden CD, Frackowiak RSJ: The relationship between locomotor disability, autonomic dysfunction and the integrity of the striatal dopaminergic system in patients with multiple system atrophy, pure autonomic failure and Parkinson's disease, studied with PET. *Brain* 1990;113:1539-1552.
- 72) Fearnley J, Lees AJ: Aging and Parkinson's disease: Substantia nigra regional selectivity. *Brain* 1991;114:2283-2301
- 73) Paulus W, Jellinger K: The neuropathologic basis of different clinical subgroups of Parkinson's disease. *J Neuropathol Exp Neurol* 1991;50:743-755
- 74) Seeman P, Niznik HB: Dopamine receptors and transporters in Parkinson's disease and schizophrenia. *FASEB J* 1990;4:2737-2744
- 75) Birkmayer W, Danielczyk W, Neumayer E, Riederer P: Dopaminergic supersensitivity in Parkinsonism. *Adv Neurol* 1975;9:121-129
- 76) Lee T, Seeman P, Rajput A, Farley IJ, Hornykiewicz O: Receptor basis for dopaminergic supersensitivity in Parkinson's disease. *Nature* 1978;273:59-61
- 77) Javitch JA, Blaustein RO, Snyder SH: [³H]mazindol binding associated with neuronal dopamine and norepinephrine uptake sites. *Mol Pharmacol* 1984;26:35-44
- 78) Battaglia G, Sharkey J, Kuhar, MJ, De Souza, EB: Neuroanatomic specificity and time course

- of alterations in rat brain serotonergic pathways induced by MDMA (3,4-methylenedioxyamphetamine): Assessment using quantitative autoradiography. *Synapse* 1991;8:249-260
- 79) Wiener HL, Hashin A, Lathja A, Sershen H: Chronic L-deprenyl-induced up-regulation of the dopamine uptake carrier. *Eur J Pharmacol* 1989; 163:191-194
- 80) DeJesus OT, Van Moffaert GJC, Dinerstein RJ; Friedman AM. Exogenous l-DOPA alters spiperidol binding, in vivo, in the mouse striatum. *Life Sci* 1986;39:341-349
- 81) Seeman P, Guan HC, Niznik HB: Endogenous dopamine lowers the dopamine D2 receptor density as measured by [³H]raclopride: Implications for positron emission tomography of the human brain. *Synapse* 1989;3:96-97
- 82) Young LT, Wong DF, Goldman S, et al.: Effects of endogenous dopamine on kinetics of [³H]N-methylspiperone and [³H]raclopride binding in the rat brain. *Synapse* 1991;9:188-194
- 83) Wachtel SR, Abercrombie ED: L-3,4-Dihydroxyphenylalanine-induced dopamine release in the striatum of intact and 6-hydroxy-dopamine-treated rats: Differential effects of monoamine oxidase A and B inhibitors. *J Neurochem* 1994; 63:108-117
- 84) Kaakkola S, Wurtman RJ: Effects of catechol-O-methyltransferase inhibitors and L-3,4-dihydroxyphenylalanine with or without carbidopa on extracellular dopamine in rat striatum. *J Neurochem* 1993;60:37-44
- 85) Buu NT: Vesicular accumulation of dopamine following L-DOPA administration. *Biochem Pharmacol* 1989;38:1787-1792
- 86) Brannan T, Knott P, Kaufman H, Leung L, Yahr M: Intracerebral dialysis monitoring of striatal dopamine release and metabolism in response to L-DOPA. *J Neural Transmission* 1989;75:149-157
- 87) Koshimura K, Ohue T, Akiyama Y, Iton A, Miwa S: L-DOPA administration enhances exocytotic dopamine release in vivo in the rat striatum. *Life Sci* 1992;51:747-755
- 88) Laruelle M, Baldwin R, Malison R, Zea-Ponce Y, Zoghbi S, Al-Tikriti M, Sybirska E, Zimmermann R, Wisniewski G, Newmeyer J, Milius R, Wang S, Smith E, Roth R, Charney D, Hoffer P, Innis R: SPECT imaging of dopamine and serotonin transporters with [¹²³I]β-CIT: Pharmacological characterization of brain uptake in nonhuman primates. *Synapse* 1993;13:295-309
- 89) Abercrombie ED, Jacobs BL: Dopaminergic modulation of sensory responses of striatal neurons: Single unit studies. *Brain Res* 1985;358:27-33
- 90) Antonini A, Schwarz J, Oertel WH, Beer HF, Madeja UD, Leenders KL: [¹¹C]raclopride and positron emission tomography in previously untreated patients with Parkinson's disease: Influence of L-dopa and lisuride therapy on striatal dopamine D2 receptors. *Neurology* 1994;44: 1325-1329
- 91) Heinonen EH, Anttila MI, Lammintausta RA: Pharmacokinetic aspects of 1-deprenyl (Selegiline) and its metabolites. *Clinical Pharmacology & Therapeutics* 1994;56:742-749
- 92) Engberg G, Elebring T, Nissbrandt H: Deprenyl (selegiline), a selective MAO-B inhibitor with active metabolites; Effects on locomotor activity, dopaminergic neurotransmission and firing rate of nigral dopamine neurons. *J Pharm & Experimental Therapeutics* 1991;259:841-847
- 93) Okuda C, Segal DS, Kuczenski R: Deprenyl alters behavior and caudate dopamine through an amphetamine-like action. *Pharmacology, Biochem & Behavior* 1992;43:1075-1080
- 94) Fang J, Yu PH: Effect of L-deprenyl, its structural analogues and some monoamine oxidase inhibitors on dopamine uptake. *Neuropharmacology* 1994;33:763-768
- 95) Rajput A, Rodzilsky B, Rajput A: Accuracy of clinical diagnosis in parkinsonism—a prospective study. *Can J Neurol Sci* 1991;18:275-278