

Effects of dopamine on angiotensin II-induced stimulation of Na⁺ uptake in primary cultured rabbit renal proximal tubule cells

Hyun-ju Koh, Soo-hyun Park, Ho-jae Han

Department of Veterinary Physiology, College of Veterinary Medicine, Hormone Research Center,
Chonnam National University

(Received May 17, 1998)

초대배양한 신장 근위세뇨관세포에서 ANG II의 Na⁺ uptake 촉진효과에 대한 dopamine의 효과

고현주 · 박수현 · 한호재

전남대학교 수의과대학, 호르몬연구센터
(1998년 5월 17일 접수)

초 록 : 신장 근위세뇨관세포들은 사구체에서 여과된 물질의 재흡수, 분비 및 대사에 관여하는 여러 호르몬들의 수용체들을 가지고 있다. 이들중에서 dopamine(DA)과 angiotensin II (ANG II)가 Na⁺/H⁺ 상호운반체 조절에 중요한 역할을 하고 있다. 본 연구는 초대배양한 토끼 신장 근위세뇨관세포의 Na⁺ uptake에 있어서 DA와 ANG II의 상호관계를 알아보고자 실시하였다.

DA은 농도의존적으로 Na⁺ uptake를 유의성 있게 억제하였다(10^{-6} M ; $83.2 \pm 7.2\%$, 10^{-3} M ; $67.2 \pm 3.8\%$ vs. control)($p < 0.05$). DA₁ 작용제(SKF 38393, 10^{-6} M)는 대조군의 $81.4 \pm 6.7\%$ 까지 Na⁺ uptake를 유의성 있게 억제하였으나($p < 0.05$) DA₂ 작용제는 영향을 미치지 않았다. DA₁ 길항제(SCH 23390, 10^{-6} M)에 의해 DA의 Na⁺ uptake 억제효과는 차단되었으나 DA₂ 길항제(spiperone, 10^{-6} M)에 의해서는 영향을 받지 않았다. DA와 대조적으로 10^{-11} M ANG II는 AT₁ 수용체를 통하여 대조군의 $120.7 \pm 4.9\%$ 까지 Na⁺ uptake를 유의성 있게 촉진하였다($p < 0.05$). DA 및 10^{-11} M ANG II를 병합처리하였을 때 DA은 농도의존적으로 ANG II에 유도된 Na⁺ uptake 촉진효과를 유의성 있게 차단하였다($p < 0.05$). 한편 ANG II에 의해 유도된 Na⁺ uptake 촉진작용은 DA₁ 또는 DA₂ 작용제에 의해 차단되었으나 DA에 의한 차단 효과는 DA₁ 및 DA₂ 길항제를 병합처리하였을 때만 반전되었다.

This study was supported by grants awarded to Ho-jae Han from Korea Science and Engineering Foundation(KOSEF 961-0606-057-1, HRC 97-0401-0401).

Address reprint requests to Dr. Ho-jae Han, Department of Veterinary Physiology, College of Veterinary Medicine, Chonnam National University, Kwangju 500-757, Republic of Korea.

결론적으로 DA은 DA₁ 수용체를 통하여 Na⁺ uptake를 억제하였으나 ANG II에 의한 Na⁺ uptake 촉진작용의 억제에는 DA₁ 및 DA₂ 수용체 모두가 관여하였다.

Key words : kindey, dopamine, angiotensin II, Na⁺/H⁺ antiport.

Introduction

The renal proximal tubule cells contain a variety of cellular membrane receptors which, upon binding of their selective agonists or endogenous hormones, lead to alteration in reabsorption, secretion, and metabolism. There are a number of hormones that have been shown to modulate proximal tubule Na⁺ reabsorption through their effects on the luminal Na⁺/H⁺ antiporter activity^{1,2}. Among these hormones, DA and ANG II represent two powerful hormone systems in regulation of Na⁺/H⁺ antiporter activity and have been involved in hypertension in the kidney³⁻⁵.

DA receptors which are classified into the DA₁ and DA₂, are located at various regions within the kidney including renal vasculature, juxtaglomerular cells, and the renal tubules^{6,7}. DA has been reported to inhibit Na⁺ transport⁸. But it is not yet documented which receptor is related to the regulation of Na⁺ transport in the proximal tubule cells(PTCs).

ANG II has been reported to have a biphasic effect on Na⁺ uptake in the proximal tubules: low dosages of ANG II stimulate Na⁺ uptake while high dosages of ANG II inhibit^{9,10}. ANG II receptors, which are classified into the ANG II type I and II, are located in the renal tissues including proximal tubules, juxtaglomerular cells¹¹. Especially, low dosages of ANG II are closely involved in renal hypertension development and directly stimulate Na⁺ reabsorption through ANG II receptor in the PTCs. Since alterations in the balance of responsiveness of proximal tubule to DA and ANG II have important implications for net sodium and water reabsorption, we aimed to study the interrelation between DA and ANG II.

The PTCs utilized in this study, have been observed to re-

tain a number of differentiated typical functions of the renal proximal tubules, including Na⁺-dependent alpha methyl-D-glucoside uptake, Na⁺-dependent phosphate uptake, and parathyroid hormone-sensitive cAMP production^{12,13}. Therefore this primary culture system in hormonally defined, serum free culture conditions is a powerful tool for studying interaction between DA and ANG II in the Na⁺/H⁺ antiport system. In the present study, we examined the interaction between DA and ANG II on Na⁺ uptake in primary cultured rabbit renal proximal tubule cells.

Materials and Methods

Materials : Male New Zealand White rabbits (1.5~2.0kg) were used for these experiments. Dulbecco's Modified Eagle's Medium, Nutrient Mixture F-12 (Ham), Class IV collagenase and soybean trypsin inhibitor were purchased from Life Technologies (Grand Island, NY). Angiotensin II (ANG II), dopamine (DA), spiperone (DA₂ antagonist), ouabain, insulin, hydrocortisone, transferrin, and BSA fraction V were obtained from Sigma Chemical (St. Louis, MO). PD123319 was purchased from Parke-Davis. Losartan (DuP 753) and ²²Na⁺ were purchased from DuPont/New England Nuclear. SKF 38393 (DA₁ agonist), SCH 23390 (DA₁ antagonist), and quinpirole (DA₂ agonist) were purchased from Research Biochemicals (Natick, MA). All other reagents were of the highest purity commercially available. Liquiscint was obtained from National Diagnostics (Parsippany, NY). Iron oxide was prepared by the method of Cook and Pickering¹⁴. Stock solutions of iron oxide in 0.9% NaCl were sterilized using an autoclave and diluted with phosphate buffered saline prior to use.

Isolation of rabbit renal proximal tubules and culture

conditions : Primary rabbit renal proximal tubule cell cultures were prepared by a modification of the method of Chung *et al*¹². The basal medium, D-MEM/F-12, pH 7.4, was a 50 : 50 mixture of Dulbecco's Modified Eagle's Medium and Ham's F-12. The basal medium was further supplemented with 15mM HEPES buffer (pH 7.4) and 20mM sodium bicarbonate. Immediately prior to the used of the medium, three growth supplements (5µg/ml insulin, 5µg/ml transferrin, and 5×10^{-8} M hydrocortisone) were added. Water utilized in medium preparation was purified by means of a Milli Q deionization system. Kidneys of a male New Zealand White rabbit were perfused via the renal artery, first with PBS, and subsequently with D-MEM/F-12 containing 0.5% iron oxide (wt/vol) until the kidney turned grey-black in color. Renal cortical slices were prepared by cutting the renal cortex and then homogenized with 4 strokes of a sterile glass homogenizer. The homogenate was poured first through a 253µm and then a 83µm mesh filter. Tubules and glomeruli on top of the 83µm filter were transferred into sterile D-MEM/F-12 medium containing a magnetic stirring bar. Glomeruli (containing iron oxide) were removed with a magnetic stirring bar. The remaining proximal tubules were briefly incubated in D-MEM/F-12 containing 80µg/ml collagenase (Class IV) and 0.025% soybean trypsin inhibitor. The dissociated tubules were then washed by centrifugation, resuspended in D-MEM/F-12 containing the three supplements, and transferred into tissue culture dishes. PTCs were maintained at 37°C, in a 5% CO₂-humidified environment in D-MEM/F-12 medium containing the three supplements. Medium was changed one day after plating and every three days thereafter.

Na⁺ uptake experiment : The confluent monolayers were incubated with DA or/and 10⁻¹¹M ANG II with and without their receptor agonists and antagonists for 4 hrs before Na⁺ uptake experiments. Uptake experiment was conducted as described by the method of Rindler *et al*¹⁵. For Na⁺ uptake studies, the medium was removed by aspiration. Before the uptake period, the monolayers were washed twice with 100mM Tris-HCl buffer, pH 7.4. Na⁺ uptake was measured at 37°C for 30 mins in an uptake buffer (10mM Tris, 1mM CaCl₂, 1mM MgCl₂, 140mM Choline chloride) containing 0.25µCi/

ml Na⁺ and 5×10^{-5} M ouabain (pH 7.4). At the end of the incubation period, the monolayers were gently washed three times with ice cold 100mM Tris-HCl buffer, pH 7.4, and the cells were solubilized with 1 ml of 0.1% SDS. To determine the Na⁺ incorporated intracellularly, 900µl of each sample was removed and counted in a liquid scintillation counter (Beckmann Co.). The remainder of each sample was used for protein determination¹⁶. The radioactivity counts in each sample were then normalized with respect to protein and were corrected for zero-time uptake per mg protein. All uptake measurements were made in triplicate.

Statistical analysis : Results were expressed as means ± standard error (S.E.). The difference between two mean values was analyzed by Student's *t*-test. The difference was considered statistically significant when $p < 0.05$.

Results

Dose response effects of DA on Na⁺ uptake : Fig 1 shows the dose response effects of DA on Na⁺ uptake. DA at concentrations of 10⁻⁹~10⁻³M was applied to PTCs for 4 hrs. Addition of DA to PTCs inhibited Na⁺ uptake in a dose-dependent manner. 10⁻⁶M and 10⁻³M DA inhibited Na⁺ uptake (10⁻⁶M DA : 83.2 ± 7.2% control, 10⁻³M DA : 67.2 ± 3.8% control, respectively ; $p < 0.05$). 10⁻⁶M DA close to physiological concentration was used to this study.

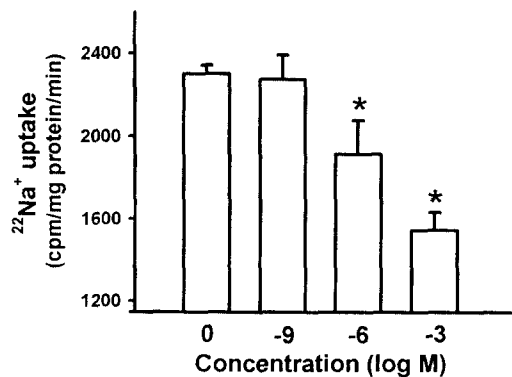


Fig 1. Dose response effects of dopamine (DA) on Na⁺ uptake. PTCs were incubated with DA (10⁻⁹~10⁻³M) for 4 hrs. Values are the means ± S.E. of 12 experiments performed on 4 different cultures. * $p < 0.05$ vs. control.

Effects of DA receptor agonists and antagonists : To determine the receptor subtype mediating the DA-induced inhibition of Na⁺ uptake, PTCs were exposed to the DA₁ agonist (SKF 38393, 10⁻⁶M), DA₂ agonist (quinpirole, 10⁻⁶M), DA₁ antagonist (SCH 23390, 10⁻⁶M), and DA₂ antagonist (spiperone, 10⁻⁶M). As shown in Fig 2, the inhibitory effect of DA on Na⁺ uptake was mediated via the DA₁ receptor because DA₁ agonist but not DA₂ agonist inhibited Na⁺ uptake (DA₁: 81.4±6.7% control; p < 0.05, DA₂: 103.0±7.6% control; p = NS), and the inhibitory effect of DA was also abolished by DA₁ antagonist but not by DA₂ antagonist.

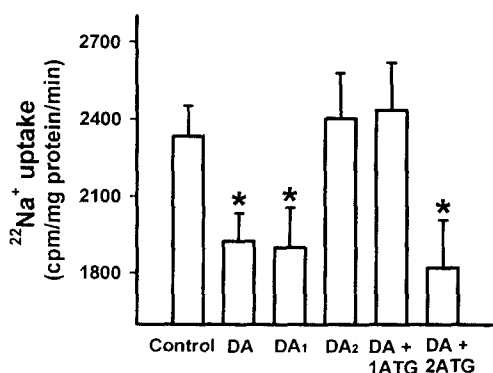


Fig 2. Effects of DA₁, DA agonists, and antagonists on Na⁺ uptake. DA (DA, 10⁻⁶M), DA₁ agonist SKF 38393 (DA₁, 10⁻⁶M), DA₂ agonist quinpirole (DA₂, 10⁻⁶M), DA₁ antagonist SCH 23390 (1ATG, 10⁻⁶M), and DA₂ antagonist spiperone (2ATG, 10⁻⁶M) were treated to the PTCs for 4 hrs. Values are the means±S.E. of 9 experiments performed on 3 different cultures. *p < 0.05 vs. control.

Effects of ANG II and ANG II receptor antagonists on Na⁺ uptake : 10⁻¹¹M ANG II was treated to PTCs for 4 hrs before uptake experiment. 10⁻¹¹M ANG II stimulated Na⁺ uptake by 120.7±4.9% control. To determine the receptor subtype mediating the ANG II-induced stimulation of Na⁺ uptake, cells were exposed to the specific receptor antagonist losartan [ANG II receptor subtype 1 (AT₁) antagonist, 10⁻⁸M] or PD123319 [ANG II receptor subtype 2 (AT₂) antagonist, 10⁻⁸M] in the absence or presence of 10⁻¹¹M ANG II. As shown in Fig 3, losartan prevented 10⁻¹¹M ANG II-induced stimulation of Na⁺ uptake but PD123319 did not (10⁻¹¹M ANG II : 120.7±4.9% control vs. ANG II

plus losartan : 101.1±6.4% control ; p < 0.05).

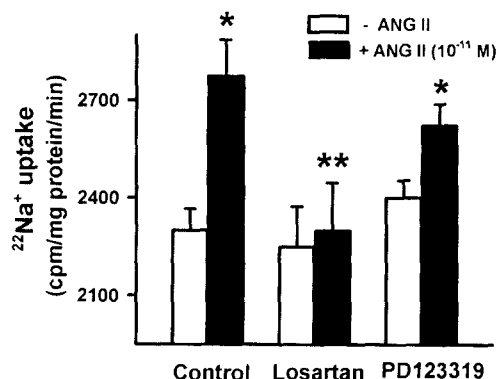


Fig 3. Effects of angiotensin II (ANG II) receptor antagonists on Na⁺ uptake. PTCs were treated with ANG II (10⁻¹¹M) alone or together with losartan (AT₁ receptor antagonist, 10⁻⁸M), or PD123319 (AT₂ receptor antagonist, 10⁻⁸M). Values are the means±S.E. of 18 experiments performed on 6 different cultures. *p < 0.05 vs. control. **p < 0.05 vs. ANG II alone.

Interaction between the DA and ANG II on Na⁺ uptake : When DA (10⁻⁹~10⁻³M) was added simultaneously, the stimulatory effects of ANG II were abolished in a DA dose-dependent manner (Fig 4). To determine which receptor subtype mediate the antagonistic effect of ANG II-induced stimulation of Na⁺ uptake, PTCs were incubated with ANG II (10⁻¹¹M) alone or together with DA, DA₁ agonist,

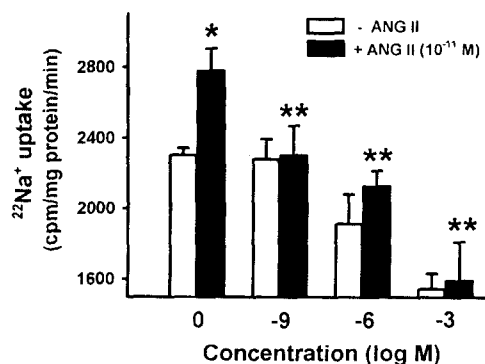


Fig 4. Effects of DA on ANG II-induced stimulation of Na⁺ uptake. PTCs were incubated with ANG II (10⁻¹¹M) alone or together with DA (DA, 10⁻⁹~10⁻³M) for 4 hrs. Values are the means±S.E. of 12 experiments performed on 4 different cultures. *p < 0.05 vs. ANG II alone.

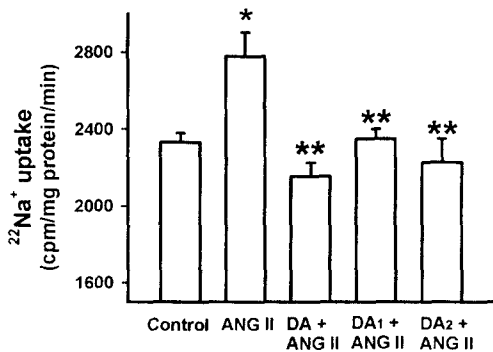


Fig 5. Effects of DA and DA agonists on ANG II-induced stimulation of Na⁺ uptake. PTCs were incubated with ANG II (10⁻¹¹M) alone or together with DA (10⁻⁶M), DA₁ agonist SKF 38393 (10⁻⁶M), or DA₂ quinpirole (10⁻⁶M) for 4 hrs. Values are the means ± S.E. of 9 experiments performed on 3 different cultures. *p < 0.05 vs. control, **p < 0.05 vs. ANG II.

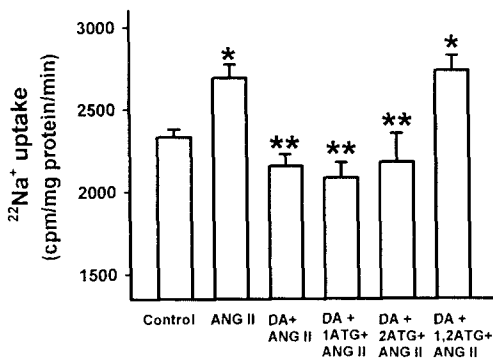


Fig 6. Effects of DA antagonists on ANG II-induced stimulation of Na⁺ uptake. PTCs were incubated with ANG II (10⁻¹¹M) alone or together with DA (10⁻⁶M) and/or with DA₁ antagonist SCH 23390 (1ATG, 10⁻⁶M), DA₂ antagonist spiperone (2ATG, 10⁻⁶M), or both antagonists for 4 hrs. Values are the means ± S.E. of 9 experiments performed on 3 different cultures. *p < 0.05 vs. control, **p < 0.05 vs. ANG II.

and DA₂ agonist. Both DA₁ and DA₂ agonists exhibited the effect of DA (Fig 5). The antagonistic effect of DA was not reversed by DA₁ or DA₂ antagonists; however, a complete reversion required the combination of both DA₁ and DA₂ antagonists (Fig 6). These results indicate that both DA₁ and DA₂ receptors mediated the inhibitory effect of DA on ANG II-induced stimulation of Na⁺ uptake.

Discussion

The kidney plays an important role in the regulation of blood pressure and body fluid homeostasis¹⁷. The proximal tubules are a major site of salt and water reabsorption in the mammalian nephron. The Na⁺ reabsorption of proximal tubule are under the control of various hormones including ANG II, DA^{18,19}. DA was active at a concentration of 10⁻⁶M; plasma and peritubular concentrations of DA are lower than 10⁻⁶M, but DA is produced locally in the kidney, and the intracellular concentration of DA may be in the micromolar range^{20,21}. Few data are available on the effects of DA in the PTCs in hormonally defined, serum free culture conditions, but in the present study, we showed that DA inhibited Na⁺ uptake in a dose-dependent manner in the PTCs (Fig 1). The results of this study corroborate inhibition of the Na⁺/H⁺ antiporter demonstrated in rat proximal nephron segments and in renal brush-border membrane^{22,23}.

Kidney has two types of DA receptors: DA₁ and DA₂. In the present study, the results summarized in Fig 2 demonstrate that the inhibition of Na⁺ uptake by DA in the PTCs was reproduced by the DA₁ agonist SKF 38393, whereas the DA₂ agonist quinpirole had no effect on Na⁺ uptake. It was completely abolished by DA₁ antagonist SCH 23390 but not by DA₂ antagonist spiperone. That this action of DA is mediated via its DA₁ receptor is further supported by several other results^{7,22} that the DA₁ but not DA₂ receptor is mainly associated with inhibition of renal Na⁺ transport. In rat renal cortical brush-border membrane vesicles DA and the DA₁ agonist, SKF 82526, but not DA₂ agonist, LY 171555, decreased Na⁺ uptake apparently by inhibiting Na⁺/H⁺ antiport activity. This effect was blocked by the DA₁ antagonist, SCH 23390²⁴. Indeed, DA₁ receptors are predominantly located in proximal tubules, but DA₂ receptors in glomeruli and mesangial cells^{6,25,26}. But Bello Reuss *et al*²⁷ reported that DA and DA agonists had no effect on fluid transport in the isolated perfused proximal straight tubule of the rabbit. In contrast, it was reported that DA increased luminal uptake of Na⁺ by activation of the Na⁺/H⁺ antiport in isolated rabbit proximal tubule cells²⁸. Whether these ap-

parently conflict reports are related to differences in experimental model system or receptor subtypes remains to be determined.

Although the effects of DA have previously been assessed singularly, this agent has not been investigated in terms of the interactions that this may have with other hormones on Na⁺ uptake in the PTCs. In contrast to DA, ANG II stimulated Na⁺ uptake via a AT₁ receptor (Fig 3). The ANG II -induced stimulation of Na⁺ uptake was markedly attenuated by DA (Fig 4). This effect of DA was mediated via both DA₁ and DA₂ receptors, because DA₁ and DA₂ agonists exerted similar effects (Fig 5), and the effect of DA was prevented only by the combination of DA₁ and DA₂ antagonists (Fig 6). This result is also supported by the report that DA₂ receptors are involved in the antagonistic effect of DA on ANG II stimulation of Na⁺ uptake, and thus provides an additional role of DA₂ receptors in the inhibitory effect of DA on proximal tubule Na⁺ reabsorption²⁰. Bertorello *et al*²⁹ also reported the role of DA₁ and DA₂ receptors in the results that sodium pump activation may require simultaneous activation of both DA₁ and DA₂ receptors in the proximal tubule. Cheng *et al*³⁰ reported that DA, acting through DA₁, acting DA₁ receptors, decrease AT₁ receptor expression in proximal tubule, suggesting the interaction between AT₁ receptor and DA₁ receptor. However, in the present study, in addition to DA₁, DA₂ had effect on the inhibition of ANG II -induced stimulation of Na⁺ uptake. Further studies about the these signal pathways are needed.

In summary, DA inhibits ANG II -induced stimulation on Na⁺ uptake by DA₁ but not DA₂ receptors mediate the effects of DA to inhibit Na⁺ uptake while both DA₁ and DA₂ receptors mediated the antagonistic effect of DA on ANG II stimulation of Na⁺ uptake.

Conclusion

The present study examined the interaction between DA and ANG II on Na⁺ uptake in PTCs. DA caused a dose-dependent inhibition of Na⁺ uptake. DA₁ agonist but not DA₂ agonist inhibited Na⁺ uptake, and the inhibitory effect of DA was abolished by DA₁ antagonist but not by DA₂ an-

tagonist. In contrast to DA, ANG II stimulated Na⁺ uptake via AT₁ receptor. When DA and ANG II were added together, DA blocked ANG II -induced stimulation of Na⁺ uptake in a dose-dependent manner. Both DA₁ and DA₂ agonists inhibited stimulatory effect of ANG II on Na⁺ uptake. This inhibitory effect of DA was reversed only by the combination of both DA₁ and DA₂ antagonists. In conclusion, DA inhibits Na⁺ uptake through DA₁ receptors, while both DA₁ and DA₂ receptors mediate the inhibitory effect of DA on ANG II -induced stimulation of Na⁺ uptake.

Reference

1. Grinstein S, Rothstein A. Mechanisms of regulation of the Na⁺/H⁺ exchanger. *J Membr Biol*, 90:1-12, 1996.
2. Mahnensmith RL, Aronson PS. The plasma membrane sodium-hydrogen exchanger and its role in physiological and pathophysiological process. *Circ Res*, 56:773-788, 1985.
3. Pelayo JC, Filder RD, Eisner GM, *et al*. Effects of dopamine blockade on renal sodium excretion. *Am J Physiol*, 245:F247-F253, 1983.
4. Schuster VL, Kokko JP, Jacobson HR. Angiotensin II directly stimulates sodium transport in rabbit proximal convoluted tubules. *J Clin Invest*, 73:507-515, 1984.
5. Weinman EJ, Dubinsky W, Shenolikar S. Regulation of the renal Na⁺/H⁺ exchanger by protein phosphorylation. *Kidney Int*, 36:519-525, 1989.
6. Felder CC, McKelvey AM, Gitler MS, *et al*. Dopamine receptor subtype in renal brush border and basolateral membranes. *Kidney Int*, 36:183-193, 1989.
7. Felder RA, Felder CC, Eisner GM, *et al*. The dopamine receptor in adult and maturing kidney. *Am J Physiol*, 257:F315-F327, 1989.
8. Felder CC, Campbell T, Albrecht F, *et al*. Dopamine inhibits Na⁺/H⁺ exchanger activity in renal BBMVs by stimulation of adenylate cyclase. *Am J Physiol*, 259:F297-F303, 1990.
9. Han HJ, Koh HJ, Park SH. Regulatory mechanisms of angiotensin II on the Na⁺/H⁺ antiport system in rabbit renal proximal tubule cells. I. Stimulatory effects of

- ANG II on Na⁺ uptake. *Korean J Physiol Pharmacol*, 1:413-423, 1997.
10. Han HJ, Park SH, Koh HJ. Regulatory mechanisms of angiotensin II on the Na⁺/H⁺ antiport system in rabbit renal proximal tubule cells. II. Inhibitory effects of ANG II on Na⁺ uptake. *Korean J Physiol Pharmacol*, 1:425-434, 1997.
 11. Timmermans PBM, Wong P, Chiu AT, *et al.* Angiotensin II receptors and angiotensin II receptor antagonists. *Pharmacol*, 45:205-251, 1993.
 12. Chung SD, Alavi N, Livingston D, *et al.* Characterization of primary rabbit kidney cultures that express proximal tubule functions in a hormonally defined medium. *J Cell Biol*, 95:118-126, 1982.
 13. Han HJ, Kang JW, Park KM, *et al.* Functional characterization of primary culture cells grown in hormonally defined, serum-free medium and serum-supplemented medium. *Korean J Vet Res*, 36:551-563, 1996.
 14. Cook WF, Pickering GW. A rapid method for separating glomeruli from rabbit kidney. *Nature*, 182:1103-1104, 1958.
 15. Rindler MJ, Taub M, Saier MH Jr. Uptake of ²²Na⁺ by cultured Dog Kidney Cells(MDCK). *J Biol Chem*, 254:111431-111439, 1979.
 16. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Bio-chemistry*, 72:248-254, 1976.
 17. Smith MC, Dunn MJ. in *The kidney*(Brenner BM, Receptor FC, eds). pp.1221-1251, 1986.
 18. Cogan MG. Angiotensin II : A powerful controller of sodium transport in the early proximal tubule. *Hypertension*, 15:451-458, 1990.
 19. Aperia A. Dopamine action and metabolism in the kidney. *Curr Opin Nephrol Hypertens*, 3:39-45, 1994.
 20. Hagege J, Richet G. Proximal tubule dopamine histofluorescence in renal slices incubated with L-dopa. *Kidney Int*, 27:3-8, 1985.
 21. Hayashi M, Yamaji Y, Kitajima W, *et al.* Aromatic L-amino acid decarboxylase activity along the rat nephron. *Am J Physiol*, 258:F28-F33, 1990.
 22. Sheikh-Hamad D, Wang YP, Jo OD, *et al.* Dopamine antagonizes the actions of angiotensin II in renal brush-border membrane. *Am J Physiol*, 264:F737-F743, 1993.
 23. Gesek FA, Schoolwerth AC. Hormonal interactions with the proximal Na⁺-H⁺ exchanger. *Am J Physiol*, 258:F514-F521, 1990.
 24. Bello-Reuss E, Higashi Y, Kaneda Y. Dopamine decrease fluid reabsorption in straight portions of rabbit proximal tubule. *Am J Physiol*, 242:F634-F640, 1982.
 25. Missale CM, Memo M, Liberini P, *et al.* Dopamine selectively inhibits angiotensin II -induced aldosterone secretion by interaction with D-2 receptors. *J Pharmacol Exp Ther*, 246:1137-1143, 1988.
 26. Lokhandwala MF, Amenta F. Anatomical distribution and function of dopamine receptors in the kidney. *FASEB J*, 5:3023-3030, 1991.
 27. Felder CC, Campbell T, Jose PA. Role of cAMP on dopamine-1 (DA-1) receptor regulated Na⁺-H⁺ antiport in renal tubular brush border membrane vesicles (BBMV) (Abstract). *Kidney Int*, 35:172, 1989.
 28. Mochida S, Kobayashi H, Libet B. Stimulation of adenylate cyclase in relation to dopamine-induced long-term enhancement (LTE) of muscarinic depolarization in the rabbit superior cervical ganglion. *J Neurosci*, 7: 311-318, 1987.
 29. Bertorello A, Hokfelt T, Goldstein M, *et al.* Proximal tubule Na⁺-K⁺-ATPase activity is inhibited during high-salt diet; evidence for DA-mediated effect. *Am J Physiol*, 254:F795-F801, 1988.
 30. Cheng HF, Becker BN, Burns KD, *et al.* Angiotensin II upregulates type-1 angiotensin II receptors in renal proximal tubule. *J Clin Invest*, 95:2012-2019, 1995.