Inhibition of Monoamine Oxidase by Palmatine

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Monoamine oxidase (EC 1.4.3.4; MAO), present in the outer mitochondrial membrane, plays an important role in the catabolism of catecholamines and serotonin, and regulation of their levels in the brain (Tipton *et al.*, 1987). In addition, MAO confers protection by inactiva-ting potentially toxic exogenous amines.

Many MAO inhibitors have beem reported belonging to various chemical classes: isoquinolines and tetrahydroisoquinolines (Bembenek et al., 1990), 4-(2-benzofuranyl)piperidines (Strolin-Benedetti and Dostert, 1987), oxadiazoles (Mazouz et al., 1990), phenoxathin-10,10-dioxides (White and Scates, 1992), and natural xanthones (Suzuki et al., 1981). In addition, oxygen-containing compounds such as coumarin, dibenzofuran, flavone, xanthene, thioxanthone and acridone also exert an inhibitory action against MAO (Thull and Testa, 1994). Among the isoquinoline derivatives, salsolinol, tetrahydropapaveroline, 1-methylisoquinoline, N-methylisoquinolinium ion and tetrahydroisoquinolines inhibit MAO activity (Bembenek et al., 1990; Giovine et al., 1976; Naoi et al., 1988, 1989).

Recently, we investigated the inhibitory effects of bioactive fractions from *Coptis japonica* on MAO activity and were able to prove that their bioactive fractions mainly contained the protoberberine isoquinoline alkaloids (data not shown). Palmatine belongs to a group of the protoberberine alkaloids and has been demonstrated to carry antiarrhythmic (Ma et al., 1985), analgesic (Chen et al., 1984), antimalarial (Vennerstrom and Klayman, 1988) and sedative (Hsieh et al., 1993) effects. In this study, the inhibitory effects of palmatine on MAO activity were examined using kynuramine as a substrate.

Palmatine chloride, kynuramine, 4-hydroxyquinoline, zinc sulfate, iproniazid and sucrose were purchased from

Sigma Chemical Co. (St. Louis, MO, USA). All other chemicals were of reagent grade.

Mice (ICR, male, 20-25 g) were obtained from Samyook Animal Center (Soowon, Korea). Mouse brain mitochondria was isolated by the method of Thull and Testa (1994) with a slight modification (Lee $et\ al.$, 1999). MAO activity was adjusted to 0.320 ± 0.012 nmol/min/mg protein for the experiments. Protein amounts were determined by the method of Lowry $et\ al.$ (1951) using bovine serum albumin as a standard.

MAO activity was measured fluorimetrically using kynuramine as a substrate according to Kraml's method (1965). Level of 5-hydroxyquinoline formed by enzyme reaction for 30 min at 37° C was measured using fluorospectrophotometer (λ ex., 315 nm/ λ em., 380 nm, Hitachi Model F-3000, Tokyo, Japan).

The values of the Michaelis constant (K_m) and the maximum velocity (V_{max}) were obtained by Lineweaver-Burk's plot using various concentrations of kynuramine. Results were presented means \pm SEM from five experiments. Statistical significances were determined at the P<0.05 using Student's t-test.

Previously, the bioactive fractions containing the protoberberine isoquinoline alkaloids were found to inhibit MAO activity (data not shown). In addition, isoquinoline compounds such as salsolinol, tetrahydropapaveroline, N-methylisoquinoline, 1-methylisoquinoline, tetrahydroisoquinolines and higenamine have been found to inhibit MAO activity (Bembenek *et al.*, 1990; Giovine *et al.*, 1976; Naoi *et al.*, 1988, 1989; Lee *et al.*, 1999). Hence, we investigated the effects of palmatine on MAO activity.

In this experiment, palmatine displayed a mild inhibitory effect on MAO (40.0% inhibition at concentration of 50 μ M) (Table I). The IC₅₀ value of palmatine was 90.6 μ M. However, pamatine's activity against MAO was less than iproniazid, a selective MAO inhibitor (Table I).

The values of K_m and V_{max} of the mouse brain MAO using kynuramine as a substrate were $78.2 \pm 4.0 \mu M$

Table I. Inhibitory effects of palmatine on mouse brain monoamine oxidase (MAO).

Inhibitor	MAO activity (nmol/min/mg protein) (% of control)
Control	0.320±0.012 (100)
lproniazid, 10 μM	0.163±0.031 (50.9)**
Palmatine, 25 μM	0.241±0.016 (75.3)*
50 μΜ	0.192±0.015 (60.0)**
100 μΜ	0.152±0.007 (47.5)***

Control level of MAO activity was taken as 0.320 nmol/min/mg protein. Iproniazid was used as a positive control. The data are presented as means \pm SEM for five experiments. Significantly differenct from the control value: *, P<0.05; **, P<0.01; ***, P<0.001 (Student's t-test).

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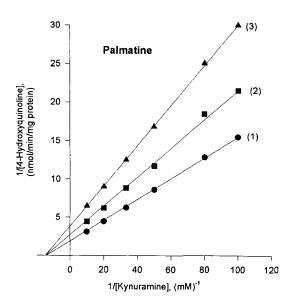


Fig. 1. Inhibition of monoamine oxidase (MAO) by palmatine added in the enzyme reaction mixture. The reciprocal of MAO activity was plotted against the reciprocal of substrate concentration. Palmatine concentration: 1, 0 μ M; 2, 25 μ M; 3, 80 μ M.

(n=5) and 0.65 ± 0.05 nmol/min/mg protein (n=5), respectively. Furthermore, the kinetic analysis by a Lineweaver-Burk reciprocal plot indicates that palmatine is a noncompetitive inhibitor of MAO (Fig. 1).

Palmatine exerts a sedative effect. The mechanism of action may be related to decrease in the levels of catecholamine and serotonin in the cortex (Hsieh et al., 1993). Palmatine was found to decrease dopamine content by reducing tyrosine hydroxylase activity in PC12 cells (Lee and Kim, 1996). Palmatine also inhibits competitively bovine adrenal tyrosine hydroxylase (Lee et al., 1997). Our present study indicates that palmatine noncompetitively inhibits MAO activity. Palmatine interacts with DNA, but does not induce apoptosis in human leukemic HL-60 cells (Kuo et al., 1995). Palmatine also interacts with the HIV-I reverse transcriptase and inhibits the enzyme activity (Gudima et al., 1994). Despite palmatine's interaction with DNA, above results suggest that palmatine may be involved in the regulation of catecholamine levels at the biologically active sites. The In vivo physiological functions of palmatine need to be studied further.

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REFERENCES CITED

Bembenek, M. E., Abell, C. W., Chrisey, L. A., Rozwadowska, M. D., Gessner, W. and Brossi, A., Inhibition of monoamine oxidases A and B by simple isoquinoline alkaloids: racemic and optically active 1,2,3,4-tetrahydro-, 3,4-dihydro- and fully aromatic isoquinolines. J. Med. Chem., 33, 147-152 (1990).

Chen, S. H. and Sha, N. H., Dose dependent relationship of analgesic effect of Corydalis app. (Papaveraceae) in mice. *Chin. J. Pharm. Anal.*, 4, 162-163 (1984).

Giovine, A., Renis, M. and Bertolino, A., In vivo and in vitro studies on the effect of tetrahydropa paveroline and salsolinol on COMT and MAO activity in rat brain. *Pharmacol.*, 14, 86-94 (1976).

Gudima, S. O., Memelova, L. V., Borodulin, V. B., Pokholok, D. K., Mednikov, B. M., Tolkachev, O. N. and Kochetkov, S. N., Kinetic analysis of interaction of human immunodeficiency virus reverse transcriptase with alkaloids. *Molekul. Biolog. (Russia)*, 28, 1308-1314 (1994).

Hsieh, M.-T., Su, S.-H., Tsai, H.-Y., Peng, W.-T., Hsieh, C.-C. and Chen, C.-F., Effects of palmatine on motor activity and the concentration of central monoamines and its metabolites in rats. *Japan J. Pharmacol.*, 61, 1-5 (1993).

Kraml M., A rapid microfluorimetric determination of monoamine oxidase. *Biochem. Pharmacol.*, 14, 1684-1685 (1965).

Kuo, C. L., Chou, C. C. and Yung, B. Y. M., Berberine complexes with DNA in the berberine-induced apoptosis in human leukemic HL-60 cells. Cancer Lett., 93, 193-200 (1995).

Lee, M. K. and Kim, H. S., Inhibitory effects of the protoberberine alkaloids from the root of Coptis japonica on catecholamine biosynthesis in PC12 cells. *Planta Med.*, 62, 31-34 (1996).

Lee, M. K., Zhang, Y. H. and Kim, H. S., Inhibition of tyrosine hydroxylase by palmatine. *Arch. Pharm. Res.*, 19, 258-260 (1996).

Lee, S. S., Yoon-Choi, H. S., Kim, E. I. and Lee, M. K., Inhibition of monoamine oxidase by higenamine. *Med. Sci. Res.,* 27, 71-72 (1999).

Lowry, O. H., Rosebrough, N. J., Farr, A. L. and Randall, R. J., Protein measurement with the Folin phenol reagent. *J. Biol. Chem.*, 193, 265-275 (1951).

Ma, S. S., Chen, K. G., Ma, Y. L. and Bau, S. F., Screening test of antiarrythmia effect of Corydalis turtschaninovii (Papaveraceae) in rats. *Bull. Chin. Materia Med.*, 10, 41-42 (1985).

Mazouz, F., Gueddari, S., Burstein, C., Mansuy, D. and Milcent, R., 5-(4-Benzyloxyphenyl)-1,3,4-oxadiazol-2 (3H)-one derivatives and related analogues: new reversible, highly potent, and selective monoamine oxidase type B inhibitors. *J. Med. Chem.*, 36, 1157-1167 (1993).

Naoi, M. and Nagatsu, T., Inhibition of type A mono-

- amine oxidase by methylquinolines and structurally related compounds. *J. Neurochem.*, 50, 1105-1110 (1988).
- Naoi, M., Takahashi, T., Darrez, H., Kabeya, R. and Taguchi, E., N-Methylisoquinolinim ion as an inhibitor of tyrosine hydroxylase, aromatic L-amino acid decarboxylase and monoamine oxidase. *Neurochem. Int.*, 15, 315-320 (1989).
- Strolin-Benedetti, M. and Dostert, P. L., Overview of the present state of MAO inhibitors. *J. Neural Transm.* (Suppl), 23, 103-119 (1987).
- Suzuki, O., Katsumata, Y., Oya, M., Chari, V. M., Vermes, B., Wagner, H. and Hostettmann, K., Inhibition of type A and type B monoamine oxidases by naturally

- occurring xanthones. *Planta Med.*, 42, 17-21 (1981). Thull, U. and Testa, B., Screening of unsubstituted cyclic compounds as inhibitors of monoamine oxidases. *Biochem. Pharmacol.*, 47, 2307-2310 (1994).
- Tipton, K. F., O'Carroll, A. M. and McCrodden, J. M., The catalytic behavior of monoamine oxidase. J. Neural Transm. (Suppl), 23, 25-35 (1987).
- Vennerstrom, J. L. and Klayman, D. L., Protoberberine alkaloids as antimalarials. *J. Med. Chem.*, 31, 1084-1087 (1988).
- White, H. L. and Scates, P., Mechanism of monoamine oxidase-A inhibition by BW 1370U87. *Drug Dev. Res.*, 25, 191-199 (1992).