가

١. Dragoo 5 가 Anderegg 3 가 Randall 가 가 가 Goon . Herbert 가 Dragoo . Andreasen

. Hany 가 Rud 가 가 Bollen, Quirynen 가 가 Ra=0.2μm(Ra : average surface roughness) Pitaru in vitro Nishimura . Cogen 가 . Khatiblou 가 Sommerman in vitro 가 , Kan SEM 가 가 . Caughman II. Bruce 1.

		20	. 2
1) 1 :			
2) 2 :		4 :	silicone
(GC Fuji II	LC)	mold	,
3) 3 :	(3M.		20
Restorative Z100)		•	
4) 4 : (Cavex	. Holland)		
		3.	
2.			
4) 0 . 0 . 0		40/	
1) : 3 × 3 × 2mm 2)			penicillin, streptomycin rial solution(Gibco . USA),
3)			co . USA), DMEM(Gibco .
1 :		USA)	3
		3371)	•
(Gracey curette, Hu - Frie	edy co.)	5	
		9	5% , 37
PBS (gauze	95%, 5	5% CO ₂ .
250 270	15		0.25% trypsin - EDTA 3
		Mℓ	•
2 :	_		
5		4.	
	20		
. mylar stri 가	р		24 microwell
71			well 가 1
3 : 50		× 10 ⁵ /M <i>Q</i>	0.59/
J . JU		A I U / IVIE	. 95%,

Table 1. Experimental Groups

		Sample size	
	Cell counting	SEM	
1 :	12	2	4
2 :	12	2	4
3 :	12	2	4
4 :	12	2	4

a) NRD: Normal Root Dentin

b) R - M GIC: Resin - Modified Glass Ionomer cement

c) CR : Composite Resin d) DA : Dental Amalgam

37 , 95%	, 5% CO	3	Hobson, England) . Ra(average surface roughness) Rt(maximal peak - to - valley height)
5.			ANOVA,
			multiple comparion
3	PBS		Person correlatison
		microwell	가 .
0.25	% trypsin - EDTA		
		_	III.
	pan blue	hemocy -	
tometer	· A	as colling la	1.
ANOV	A	multiple	
comparison	•		1
•			, , (Table 1).
6. SEM			16.7 ± 4.41(×
O. OLIVI			10 ³) $8.13 \pm 3.63 (\times 10^3)$,
			$0.72 \pm 3.33 \times 10^{3}$
			(P<0.05)
		2	14.0 ± 4.15 (×
PBS			10 ³) 1
2.5% Glutara	aldehyde 1	25%	(P<0.05).
			2
critical point	t dryer		가
aluminum	n stub		. 4
gold coating			가
7.			
			2. Scanning Electron Microscopy
4	Talana (//D	. d. T	Examination
For	m Talysurf(Rar	id Tayler -	

Table 2. The number of attached $cell(\times 10^3)$

1	2	3	4	5	6	7	8	9	10	11	12	Mean ± SD
Teeth 20.8	9.6	23.2	9.6	18.8	12.8	19.6	19.2	14.4	16.8	20.4	15.6	16.7 ± 4.41
RM - GI 8.8	9.6	11.6	18.4	13.2	18.8	14.4	19.6	8.8	10.4	16.0	18.4	14.0 ± 4.15
Resin 4.4	5.2	9.2	5.6	4.0	9.2	8.8	4.8	9.6	8.0	16.0	12.8	8.13 ± 3.63
Amalgam0.8	8.0	0.8	8.0	0.4	8.0	0.8	0.0	8.0	1.2	1.2	0.4	0.72 ± 3.33

Group 1,2,3,4 : See Table 1

Table 3. Multiple comparisons of attached cell number

	1	2	3	4
1			*	*
2			*	*
3	*	*		*
4	*	*	*	

^{*.} The mean difference is significant at the 0.05

level

.

Table 4. The value of mean surface roughness

		Mean surface roughness					
		Ra(μm)	Rt(μm)				
1 ()	0.6972 ± 0.104	5.233 ±				
1.673							
2 (RM - GI)		0.0822 ± 0.009	0.798 ±				
0.106							
3 (Resin)		0.0875 ± 0.005	$0.999 \pm$				
0.145							
4 (Amalgam))	4.2145 ± 0.985	32.29 ± 7.772				

Group 1,2,3,4 : See Table 1

Table 5. Multiple comparison of surface roughness(Ra, Rt)

	1	2	3	4
1				*
2				*
3				*
4	*	*	*	

^{*.} The mean difference is significant at the 0.05

가

(2) 2 (

1

(3) 3 ()

.

(4) 4 ()

3.

Ra(average surface rough -Rt(maximal peak to valley height) ness) (Table 3), Ra Rt .(perason correlation: 0.676) $Ra = 0.69 \pm 0.10 \mu m$, $Rt = 5.23 \pm 1.67$ 1 2 $Ra = 0.08 \pm 0.009 \mu m$, $Rt = 0.79 \pm 0.009 \mu m$ μm, $Ra = 0.08 \pm 0.005 \mu m$, $0.10 \mu m$, 3 가 $Rt = 0.99 \pm 0.14 \mu m$ $Ra = 4.21 \pm 0.98 \mu m$

 $Rt = 32.29 \pm 7.72 \mu m$

, Ra pearson correlation - 0.593, Rt pearson correlation - 0.580 .

IV.

가 가 , Pitaru in vitro . Cogen 가 가 가 가 가 (Sommerman). 가 가 Isidor, 가 Aukhil Sommerman 가 Hou Rompen 가 가 가 . Aleo 가 가 가 가 가 in vitro 가 가 in vitro

가 (Caughman et al 1990)		, ,	,	,	,
Mylar strip	가	가			가
Green, Rosenberg		가 .			
, , ,	가 가		가		가
(p<0.05).		·	,	1,2	
	en 1993). 가				٠
(Fardal 1990).		가			·
. 2		Brunette (contact guidance)			가
가 (p<0.05). Kan		,			
Peltola, Bruce		, Leirskar 24			
	•	24			가 48,
(HEMA) 가		72 Peltola		·	
(Kan, Hany - Anwar M, Cau Dragoo	ghman).	,	,		, 가

가 가 in vivo 가 가 가 in vivo Khatiblou 1mm notch 가 ٧. Nishimura 가 가 가 1. 가 Pellen - Mussi (p<0.05). 1, 2, 2. 3 가 가 3. 가 (p<0.05) 가 4. 가

가가 , 가 가 가 가 . VI.

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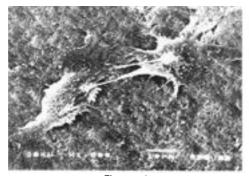
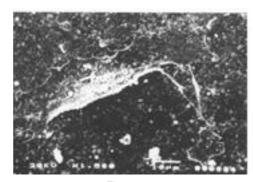




Figure 1

Figure 2



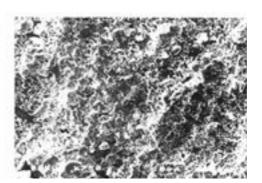


Figure 3 Figure 4

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Figure 1. Control root dentin specimen(SEM ×1,500)

Surface was covered by smear layer, therefore, showed a relatively irregular topography. Cell morphology was stellate, flat with the many surface bleb, filopodia, lamellopodia, and attached to root surface, adjacent cells intimately. Some cells are extremely flat.

Figure 2. Resin modified glass ionomer specimen(SEM ×1,500)

Surface of specimen was very smooth appearance. Cells were spindle, flat shape and exhibited the attachment apparatus. Relatively firm attachment was examined.

Figure 3. Resin specimen (SEM \times 1,500)

Cells showed a relative few attachment apparatus than root dentin, resin modified glass ionomer specimens, and cell shape was somewhat round, long.

Figure 4. Amalgam specimen(SEM ×750)

Attched cell was not observed. Only considerable rough surface was seen.

- Abstract -

Attachment of Human Gin gival Fibroblast to Various Subgingival Restorations : A Comparative Study in Vitro

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When mucoperiosteal flaps are positioned and sutured to desirable position, the wound contains several interface between tissues which differ fundamentally in composition & biological reaction. Thus the C - T surface of the flap will, on one hand, oppose another vascularized surface, and on the other, the avascular dental material for example, when root resoptions, fractured root, endodontic perforation, deep root carious lesions were filled with amalgam, glass ionomer, resin etc.

Recently, a number of case report described the successful treatment of a subgingival root lesion with restorative material & free gingival graft, open flap surgery, but more objective research was needed.

Most of study on restorative materials were concerned for cytotoxicity not for actual healing event on that materials and its influencing factors such as biocompati -

bility, surface wettability, surface topography.

The aim of this in vitro study was to evaluate the effect of amalgam, resin modified glass ionomer, composite resin per se, and their surface roughness on the growth of human gingival fibroblast.

The cells were obtained and placed on culture flask and incubated for 3 days with the prepared test materials. Then count the attached cell number with hemocytome - ter,(n=12) and 2 samples were examined with SEM about attachment cell morpholo - gy.

Another 4 samples were evaluated on their surface roughness with Talysurf and average surface roughness value(Ra) were obtained.

Statistical difference in attached cell number, roughness value were analyzed using ANOVA.

The number of attached cell was as follows, for root dentin specimen 16.7 ± 4.41 , resin modified glass ionomer 14.0 ± 4.15 , resin 8.13 ± 3.63 , amalgam 0.72 ± 3.33 ($\times 10^3$). Between root dentin and resin-modified glass ionomer, no significant difference was observed, but resin, amalgam showed a significant less cell numbers than for root dentin, resin modified glass ionomer cement.

SEM examination expressed many cell surface attachment apparatus in root dentin and resin modified glass ionomer specimens. For resin specimen, cell attachment was observed but exposed less appratus.

The average surface roughness value are following results. Dentin specimen 0.6972 ± 0.104, resin modified glass ionomer 0.0822

 $\pm\,0.009$, resin 0.0875 ± 0.005 , amalgam 4.2145 ± 0.985 (μ m). Between root dentin, resin - modified glass ionomer, and resin, no significant difference was observed, but amalgam showed a significant more rough surface than other groups.

When evaluated the interrelationship between cell attachment and surface roughness, therefore, there was weak reverse correlation. (pearson correlation: -0.593)

These results suggest that resin modified glass ionomer have the favorable healing potential when used for subgingival restoration. And for relationship between cell attachment and surface characteristics, further investigations were needed.