Jour. Korean For. Soc. 89(4): 522 - 526. 2000 韓國林學會誌 89(4): 522 - 526. 2000

Differentiation of Winter Buds of *Prunus yedoensis*Matsumura from Jeju Island Depending on the Collection Time and Media Conditions¹

Eun Ju Cheong², Chan Soo Kim² and Jae Seon Yi³

ABSTRACT

We tried to mass-propagate *Prunus yedoensis* from Jeju through tissue culture. We investigated the effect of bud collection time, the concentration of NH₄NO₃ in media and plant growth regulators(BAP, GA₃ and IBA) on the differentiation of winter buds. Buds, taken in February, flushed well regardless of various *in vitro* conditions. Bud flushing rate was significantly different depending on the collection time. BAP appeared to be effective on bud flushing. Sixty percent of buds taken in October flushed on the media containing 3.0 mg/ ℓ BAP. No buds flushed on the medium supplemented with IBA. Buds, after flushing in BAP media, grew as foliated shoots and showed a rosette of leaves. When GA₃ supplemented to the BAP-containing media as a higher concentration than that of BAP, shoots elongated and developed into normal shoots. The combination of BAP 1.0 mg/ ℓ and GA₃ 2.0 mg/ ℓ is most recommendable for shoot elongation.

Key words: BAP, flushing rate, GA3, IBA, Prunus yedoensis, winter bud

要 約

경제적으로 중요한 가치가 있는 제주 자생 왕벛나무를 조직배양을 통하여 대량 증식하고자 동아를 이용할 때 분화 및 생장에 미치는 기내 조건을 구명하고자 하였다. 동아의 채취시기, 배지의 NH4NO3의 농도, 생장조절물질의 종류 (BAP, GA3, IBA) 및 농도에 따른 영향을 조사하였다. 동아의 채취시기는 2월이 적합하였으며 그 중 1,200 mg/ ℓ NH4NO3의 농도에서 잎 전개율이 70%로 가장 높았다. 생장조절물질 처리구 중 BAP 3.0 mg/ ℓ 첨가 배지에서 채취시기에 관계없이 잎의 전개율이 높았다. 반면 IBA 처리구에서는 전혀 잎이 전개되지 않아 동아의 전개를 저해하는 것으로 나타났다. BAP 첨가 배지에서 전개된 동아는 줄기로 자라지 못하고 다발 모양을 이루었다. BAP와 함께 GA3를 처리한 결과 GA3의 농도가 BAP보다 높을 때 줄기 신장에 효과가 있는 것으로 나타났으며, BAP 1.0 mg/ ℓ 와 GA3 2.0 mg/ ℓ 혼합 처리구에서 줄기의 생장이 가장 양호하였다.

¹ Received on August 16, 2000.

² Jeju Forest Experiment Station, Korea Forest Research Institute, 1253 Sanghyo-dong Seogwipo-city 697-050, Republic of Korea 임업연구원 제주임업시험장

³ Division of Forest Science, College of Forest Science, Kangwon National University, Chunchon 200-701, Republic of Korea 강원대학교 산림자원학부.

INTRODUCTION

Nineteen species of Prunus grow in Korea and most of them have economic value in addition to ornamental importance for their beautiful flowers. Prunus yedoensis grow at the altitude of 450m to 850m in Mt. Halla. This species exhibits high variability in flower and tree form. The timber was valuable for furniture and ship in the past. Prunus yedoensis trees in Mt. Halla are distinguished geneti cally from those trees planted along the road in Korea and other countries (Kim, 1997) even though they own the same scientific name. In these days the demand for P. yedoensis is rapidly increasing due to beautiful spring flowers. Although P. vedoensis in Jeju is considered to be much valuable in many aspects, it is very difficult to propagate asexually such as cutting and grafting, possibly because of aging. In addition, seed germination rate is very low. Tissue culture technique can be applied to this species (Bonga, 1980; Ahuja and Muhs, 1985).

In Europe most *Prunus* species have been cultivated to produce cherries. Many researchers tried to produce virus-free plants through apical meristem culture of cultivars (Boxus and Quoirin, 1974, 1977) or transferred virus-resistant gene into *Prunus* species (Machado *et al.*, 1992; Tiziana *et al.*, 1995). However, there are few studies on *P. yedoensis* (Kim *et al.*, 1993; Koh *et al.*, 1997, 1998).

This study was performed to investigate the proper conditions to *in vitro* mass-propagate *P. yedoensis* using winter buds.

MATERIALS AND METHODS

One tree (ca. 40 years old) was chosen at the altitude of 600m in Mt. Halla. The winter buds were taken from current-year branches of the crown every month from October in 1997 to February in 1998. Apical and/or lateral buds were used for culture except flower buds. Small scions were prepared as long as two centimeters and surface-

sterilized in 70% ethanol for a minute and then in a 2% sodium hypochlorite solution for 20 minutes followed by three rinses with sterilized, distilled water. Before plated on media, scions were in flame in a short time.

In the first experiment, NH₄NO₃ concentrations of macro- and micro-nutrients were increased twice (800 mg/ ℓ) and three times(1,200 mg/ ℓ) as high as that of the basal one (400 mg/ ℓ) based on Woody Plant Medium(WPM, Lloyd and McCown, 1984). Sucrose(30 g/ ℓ) and agar(7 g/ ℓ) were supplemented in all media. pH was adjusted to 5.7 before autoclaving in 121°C at 1.5 kg/cm² for 15 minutes. Ten milliliters medium was dispensed in a test-tube and capped with aluminium foil. Bud flushing rate only was investigated.

In the second experiment, buds collected in February were used. 1.0, 2.0 and 3.0 mg/ ℓ 6-benzylamino purine (BAP), indole-3-butyric acid (IBA) and gibbellic acid (GA₃) were added to the basal WPM media, respectively. In the other experiment 1.0 mg/ ℓ BAP was added in combination with 0.5, 1.0 or 2.0 mg/ ℓ GA₃. Bud flushing rate, shoot formation rate, callus formation rate, shoot length and fresh weight were investigated. Each treatment consisted of two replications with 10 explants.

RESULTS AND DISCUSSION

Flushing rate of buds depending on collection time and media containing various concentrations of NH₄NO₃

Within 5 days after plating, explants became brown in test-tubes and scales were separated from buds. Small green leaves appeared after a week. In the first culture stage, most buds were contaminated with fungi and had to be transferred to fresh media.

Bud flushing rate was various depending on the collection time. February buds, followed by January buds, showed the highest rate regardless of NH₄NO₃ concentration. However, there were no differences

medium.

among the others, i. e., October, November and December buds. Treatment of 1,200 mg/ ℓ NH₄NO₃ seemed to be effective on flushing. Kim *et al.* (1993) reported that winter buds of *P. yedoensis* taken from Jeju in February flushed well in three different kinds of media and showed the highest flushing percentage in WPM. For successful flushing of buds, collection time appeared to be more important than the concentration of NH₄NO₃(Table 1).

Table 1. Differences of bud flushing rate of *P. yedoensis* depending on the concentrations of NH₄NO₃ and collection time¹.

Concentration of NH ₄ NO ₃ ² (mg/ ℓ)	Collection Time				
	Oct.	Nov.	Dec.	Jan.	Feb.
400	30	30	30	40	60
800	30	30	40	40	50
1,200	40	40	40	50	70

¹ Culture period was 3 weeks under 16-hour photoperiod.
² Other nutrients and vitamins were based on WPM

All buds flushed grew into foliated shoots but leaves formed a rosette form. The leaves turned out to be yellow and fell off within a few days. It is supposed that other nutrients and/or growth regulators may be required. In the case of white birch, buds were dormant during the winter from October to February and were activated in March or April after winter(Rinne et al., 1994). Hammerschlag (1982) suggested that a certain amount of chilling is needed to get the *in vitro* shoots from buds. *P. yedoensis* seemed to have the same habit on bud dormancy as other species.

2. BAP, GA₃ and IBA treatment in culture media

Bud flushing rate on the different culture media appeared to be similar among the growth regulators except IBA. No buds flushed in the media containing IBA(Fig. 1). In most treatments one bud established two normally-shaped leaves, no matter how many small leaves were formed on the medium containing BAP(Fig. 2).

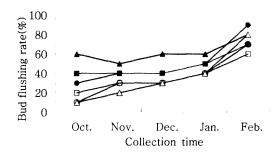


Fig. 1. Differences of bud flushing rate of *P. yedoensis* depending on the concentrations of BAP and GA₃, and collection time. Culture period was 2 weeks under 16 hours/day photoperiod.

● : BAP 1.0, ■ : BAP 2.0, ▲ : BAP 3.0, ○ : GA₃ 1.0, □ : GA₃ 2.0, △ : GA₃ 3.0

Fig. 2. The response of winter buds on the different plant growth regulators.

A: Control, B: IBA, C: GA₃, D: BAP

Even 60% of October buds flushed on the media containing 3.0 mg/ ℓ BAP. It is suggested that BAP may break the dormancy of winter buds in this species. There are some reports that BAP stimulates bud flushing in Prunus spp(Snir, 1982; Sakatani, 1989; Kim et al., 1993). Bud flushing rate was very similar between hormone-free media and GA3containing ones. However, buds in GA3 media flushed and developed into shoots when they were collected in February when buds dormancy break in spring. Considering our results, February buds may have more cytokinins than others so that flushed well in all media and could developed shoots when the GA₃ was added exogenously. Bonga(1997) observed that chilling treated buds of Larix spp. had flushed in vitro but no shoot and showed embryo-like structure. On the other hands, buds that had taken after winter developed into shoots. This report is very similar to our result. Most buds in IBA media did not flush and showed callus formation at the cut ends. It suggests that IBA works as inhibitor in bud flushing.

3. Shoot differentiation by treatment BAP+GA₃

Even though BAP was effective on bud flushing, a plantlet established still looked like a bundle of leaves and main shoot was too short to use further culture. It was required to elongate the internode of stems produced in BAP-containing media. On the other hand, GA₃ appeared to stimulate shoot elongation. For shoot development BAP-containing media were supplemented with GA₃. As the result of the treatment of GA₃, buds were elongated into shoots. GA₃, however, was affected by BAP. Three kinds of concentrations of GA₃(0.5, 1.0 and 2.0 mg/ ℓ) were combined together with 1.0 mg/ ℓ BAP (Fig. 3).

There was no difference in bud flushing rate following the ratio of BAP/GA₃. However, callus formation rate decreased depending on the ratio of BAP/GA₃ (Fig. 3). As BAP/GA₃ ratio became lower, shoot formation rate increased and shoot length

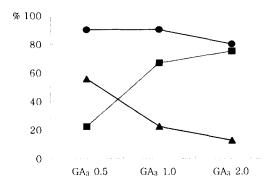


Fig. 3. Bud flushing rate, shoot formation rate and callus formation rate of winter buds of P. yedoensis on the media containing 1.0 mg/ ℓ BAP, when supplemented with 0.5, 1.0 and 2.0 mg/ ℓ GA3, respectively. Basal media was WPM and culture period was 3 weeks.

■ : Bud flushing rate,■ : Shoot formation rate,▲ : Callus formation rate

become longer and longer(Table 2). This suggests that shoot development was affected by the ratio of BAP/GA₃ rather than absolute concentration of GA₃, and vice versa.

Table 2. Shoot length and fresh weight of bud differentiated on the media supplemented with BAP and GA3.

Concentration (mg/ ℓ)	Shoot Length (mm)	Fresh Weight (mg)	
BAP 1.0+GA ₃ 0.5	2.5 ± 0.7 b*	167.0 ± 30.4 b	
BAP $1.0 + GA_3$ 1.0	$2.8 \pm 1.3 b$	$204.8 \pm 67.9 a$	
BAP 1.0+GA ₃ 2.0	5.0 ± 2.2 a	$190.3 \pm 63.0 a$	

* Means with the same letter are not significantly different at the α =0.05 following Duncan's multiple range test.

It is known that gibberellin has a positive effect on shoot elongation. When the winter buds of P. avium were cultured, $0.1 \text{ mg/} \ell$ GA₃-NH₄ accelerated the differentiation rate(Jones and Hopgood, 1979). GA₃ was also reported to give an effect on the shoot elongation of the winter buds of P. yedoensis. As the protocol mentioned above, we could get many shoots with long stem internodes by adding GA₃ and BAP to the media at the first step.

LITERATURE CITED

- Ahuja M. R. and H. J. Muhs. 1985. In vitro techniques in clonal propagation of forest tree species. In vitro techniques propagation and long term storage. Schäfer-Menuhr A(Ed). Martinus Nijhoff /Dr. W. Junk publishers, pp. 41~49.
- Bonga J.M. 1980. Plant propagation through tissue culture, emphasizing woody species. Plant Cell Culture; Results and perspectives. F. Sala, B. Parisi, R. Cella and O. Ciferri (Ed) Elsevier/ North-Holland Biomedical Press pp. 253~264.
- Bonga J.M. 1997. The effect of collection date and frozen storage on the formation of embryo -like structures and elongating shoots from explants from mature *Larix decidua* and *L. eurolepis*. Plant Cell Tissue and Organ culture

- $51(3):195\sim200.$
- 4. Boxus P. and M. Quoirin. 1974. La culture de meristemes apicaux de quelques especes de *Prunus.* Bull. Soc. R. Bot. Belg 107: 91 ~ 101.
- 5. Boxus P. and M. Quoirin. 1977. Comportement en pepiniere d'arbres fruitieres issus de culture in vitro. Acta Hortic. 78: 373~379.
- 6. Druart P. 1992. In vitro culture and micropropagation of Plum (Prunus spp.) In: Biothechnology in Agriculture and Forest 18.
- 7. Hammerschlag F. A. 1982. Factors affecting establishment and growth of peach shoots in vitro. HortScience 17:85~86.
- 8. Jones O.P. and M.E. Hopgood. 1979. The successful propagation in vitro of two rootstocks of Prunus. The plum rootstock Pixy(P. institia) and the cherry F12/1(P. avium). J. Hortic. Sci. $54:63\sim66.$
- 9. Kim C.S. 1997. Distribution and taxonomic study of Prunus yedoensis Matsumura (Rosaceae). Ph. D. Dissertation of Cheju Natl. Univ.
- 10. Kim C.S., J.K. Koh, and R.M. Cho. 1993. Effects of media, growth regulators and dark treatment on in vitro propagation using vegetative buds of Prunus yedoensis Matsumura. Kor. Soc. Plant Tissue Culture 20: 213~219.
- 11. Koh J.K., Y.C. Park, D.Y. Yang, E.S. Kim, M.Y. Oh, and S.C. Koh. 1997. Plant regeneration and somatic embryogenesis from zygotic embryoderived callus of native Prunus yedoensis in Mt. Halla, Kor. Soc. Plant Tissue Culture 24: 345 $\sim 349.$
- 12. Koh J.K., S.J. Oh, E.S. Kim, M.H. Kim and

- S.C. Koh. 1998. Plant regeneration through direct somatic embryogenesis from immature zygotic embryo of Prunus yedoensis in Mt. Halla. Kor. J. Plant Reports 11:9~14.
- 13. Lloyd G.B. and H. McCown. 1981. Commercially feasible micropropagation of mountain laurel(Kalmia latifolia) by use of shoot tip culture. Comb. Proc Intl. Plant Prop. Soc. 30: $421 \sim 427$.
- 14. Machado M.L., A.C. Machado, V. Hanzer, H. Weiss, F. Regner, H. Steinkerller, D. Mattanovich, R. Plail, E. Knapp, B. Kalthoff and H. Katinger. 1992. Regeneration of transgenic plants of Prunus armeniaca, containing the coat protein gene of Plum pox virus. Plant Cell Reports 11: $25 \sim 29$.
- 15. Marino G., P. Rossati and F. Sagrati. 1985. Storage of in vitro culture of Prunus rootstock. Plant Cell Tissue Organ Culture 5(1): $73 \sim 78$.
- 16. Rinne P., H. Tuominen, and O. Juntilla. 1994. Seasonal changes in bud dormancy in relation to bud morphoology, water and starch content, and abscisic acid concentration in adult trees of Butula pubescens. Tree Physiology 14: 549 $\sim 561.$
- 17. Sakatani S. 1989. Sakura, Propagation and Breeding of Woody Plant pp. 125~151.
- 18. Snir I. 1982. In vitro propagation of sweet cherry cultivars Hortic Science 17: 192 ~ 193.
- 19. Tiziana A., P. Lauri and C. Damiano. 1995. Agrobacterium-mediated transformation almond leaf pieces. Plant Cell Reports 14: $267 \sim 272$.