

Recent Advances in the Use of Enzymes for Environment-Friendly Swine Diets

Hong Jong-Wook · Kim In-Ho*
Dept. of Animal Resource and Science, Dankook University

환경친화성양돈사료를 위한 효소제 사용의 최근 경향

홍종욱 · 김인호*
단국대학교 동물자원학과

(CONTENTS)

국문요약	III. Conclusion
I. Introduction	Literature cited
II. Exogenous enzyme	

국문요약

본 논문의 목적을 위한 외인성 효소 즉 phytase, β -glucanase, pentosanase는 전 세계적으로 양돈사료에 첨가제로서 광범위하게 사용하고 있다. 이러한 효소의 화학적 효과는 이해가 잘 되고 있다. 하지만 돼지에서 이러한 효소들의 효과에 대해서는 아직까지 논란의 여지가 있다. Phytase는 곡류내 존재하는 피틴태 인의 이용성을 증가시킬 수 있어 배설되는 분 중 인의 오염도를 낮출 수 있고 사료내 사용하는 무기태 인의 양을 감소시킬 수 있다. 또한, 보리와 귀리에 존재하는 β -glucanase와 호밀과 밀에 존재하는 용해성 pentosans과 같은 효소들은 양돈사료에서 찾을 수 있는 항영양소 인자들을 분해하는 효과가 있다. 그래서 비전분과당류들의 소화율을 증가시키는 결과를 초래한다. 앞으로 이 분야의 연구는 현재 효소들의 효율적인 사용, 효과적인 다른 생산품의 개발 그리고 열 안정성 효소들의 개발들을 포함하고 있다.

색인어: 돼지, phytase, β -glucanase, pentosanase

I. Introduction

In the past, the aim of feeding program and diet formulations were maximizing production performance without special concern for nutrient oversupply. However, with the increased public concerns on environment, animal production has to minimize excessive N and P output in animal excreta. Therefore, animal nutrition has recently focused on not only improving of animal productivity, but also developing low-pollution diets.

N and P are the most important contributors to pollution from animal manure. Excretion of N and P in swine and poultry manure can be substantially reduced by several nutritional strategies. One of effective ways to reduce pollutants from animal manure is to use exogenous enzyme and yucca extract supplementation.

Endogenous enzymes break down the carbohydrates, proteins and fats contained in the diets into a form that can be utilized by the pigs(Corring, 1982). Enzymes are also involved in activating and hastening the many chemical reactions which take place in the animal's body. Therefore, it is possible, under conditions where enzyme production may be limiting that pig performance could be improved by the addition of enzyme to the diets. However, the expense of the enzyme products limits its practical use.

Many publications have demonstrated performance benefits of enzymes when added to barley(Classen et al., 1985 ; Elwinger and Saterby, 1987 ; Marquardt et al., 1994), wheat(Classen et al., 1995 ; Bedford and Morgan, 1996) and more corn based diets (Wyatt et al., 1997, 1999 ; Steinfeldt et al., 1998).

This paper is a review of the effects of exogenous enzyme supplementation for development of low-pollution swine diets.

II. Exogenous enzyme

(1) Phytase

1) Environmental concerns

Environmental concerns as a results of agricultural production can be divided into those related to the soil, to the surface and ground water and to the air. Farrell(1997) estimates the current world-wide feed requirement for pigs and poultry at 632 million

tons ; which is based on both industrially feedmilled and home-mixed rations. This requirement is forecast to increase to 758 million tons in 2002. However, large percentage of the P in pig diets is not utilized, much of the dietary P is excreted in the manure. Large amounts of P in manure pose an environmental concern, especially in parts of the world where land and water resources are scarce and animals are density populated. Of the 1 million tons of P excreted annually by farm animals, approximately 200,000 and 120,000 tons of P come from swine and poultry, respectively.

2) Phytate phosphorus

P is an essential component of animal body, which plays an important role in the development and maturation of the skeletal system, as well as in numerous other metabolic pathways. However, the requirement has not been determined precisely because of the variable bioavailabilities of feedstuffs from plant origin. The bioavailability of P in various feedstuffs of plant origin varies from 10 to 60%. Digestibility and availability of the P differed considerably depending on the feedstuffs. A large proportion(over 55%) of the P found in cereals and oilseeds is bound in phytate(Jongbloed et al., 1991). The digestive tract of monogastric animals dose not contain sufficient amounts of phytase. Thus, P in the phytate is poorly available to monogastric animals(Peeler, 1972).

Phytate has the potential of binding Ca and trace minerals, resulting in a decreased absorption of these minerals(Oberleas 1973). Phytate reportedly has the highest binding affinity for Cu and Zn. Specially phytic acid forms complex with other minerals, also reducing their availability. But phytase has been shown to increase the availability and retention of Ca(Simons et al, 1990) and improve the absorption of Mg, Cu, Fe and Zn(Lei et al., 1993b).

Microbial phytase supplementation improves the availability P in feedstuffs of plant origin(Simons et al., 1990 ; Denbow et al., 1995) and reduce excretion of P in the manure of pigs(Harper et al., 1997) and poultry(Um et al., 1998). Thus, a reduction of P excretion in this way will have favorable impact on the environment. Recently, a microbial phytase from *Asperigillus* has been used to improve the availability of phytate P in corn-soybean meal diets(Simons et al., 1990 ; Cromwell et al., 1993 ; Lei et al., 1993a). Supplemental phytase in swine diets has resulted in improved pig performance and bone mineralization by increasing the digestibility and retention of P and Ca and has resulted in decreased excretion of both P and Ca(Simons et al., 1990 ; Jongbloed et

al., 1993 ; Lei et al., 1993c).

3) Effects of phytase for swine

Dietary supplementation with phytase improves growth performance and phytate P availability in growing-finishing pigs. Young et al.(1993) found that performance and bone ash of weanling pigs were maintained compared to the control diet when dietary P decreased by 0.17% and 500 PTU was fed. Addition of 1,000 PTU did not further improve growth or percent bone ash, however, did restore bone ash weight which is a more sensitive indicator of an improvement in bone strength than bone ash percent(Cromwell et al., 1995). Response of growth performance of weanling pigs to increasing exogenous phytase levels diminishes at phytase levels greater than 700 PTU(Yi et al., 1996). The report of Young et al.(1993) supports this finding. The coefficient of digestibility of P in weanling pigs fed a corn-soybean meal diet has been improved from 0.27 to 0.68 when a low-P diet was fed with 1,200 PTU of exogenous phytase(Lei et al., 1993c).

Other studies with low-P diets observed an increase in the coefficient of digestibility from 0.46 to 0.69 with 750 PTU(Lei et al., 1993a) or from 0.32 to 0.55 with 800 PTU(Mroz et al., 1994). When higher levels of P were fed the coefficient of P digestibility increased from 0.60 to 0.65 or 0.63 to 0.74 when 1,500 or 1,000 PTU, respectively, were fed(Young et al., 1993 ; Adeola et al., 1995). Digestibility is typically higher in pigs fed diets containing P at or above the NRC(1998) estimated requirement for 10-20kg pigs than when P is supplied at half the estimated requirement. Therefore, the response to exogenous phytase would be expected to be diminished when P is fed near the nutrient requirement. Beers and Jongbloed(1992) fed 1,450 PTU exogenous phytase to pigs consuming a corn-barley-soybean meal-potato protein diet. The coefficient of P digestibility increased from 0.39 to 0.60 in 25kg pigs when exogenous phytase was added.

Yi et al.(1996) suggested that adding 1,050 U of phytase to the basal diet increased P apparent absorption by 23%, decreased fecal P excretion by 10%, increased average daily gain by 30%, increased average daily feed intake by 24% and increased gain/feed by 11%. Harper et al.(1997) reported that apparent digestibility of P was linearly improved with addition of 250 and 500 U/kg microbial phytase to the low P diet. On another trial conducted by Veum et al.(1996) using a grain sorghum-canola meal diet, performance including average daily gain, average daily feed intake and gain/feed ratio

were similar to feeds a low P diet with 200 to 800 U/kg of added phytase and pigs fed on adequate P diet.

Use of cereal phytase of various feeds may be a more practical alternative than the use of microbial phytase. Pointillart et al.(1984, 1987) showed positive effects of cereal phytase of wheat and triticale on dietary phytate P utilization by pigs, but their experiments lasted for approximately 6 wk, and their diets contained too much(over 80%) wheat or triticale to be applicable in the swine industry. Also, Han et al.(1997) suggested that cereal phytase in the commonly used dietary levels of wheat bran was also shown to be almost as effective as microbial phytase in improving phytate P utilization for body weight, but not for bone mineralization in weaning through finishing.

(2) β -glucanase

The mixed-linked(1 \rightarrow 3)(1 \rightarrow 4)- β -D-glucans which are frequently present in the endosperm cell walls of barley(Aman and Graham, 1987) may interfere with digestion and absorption of nutrients and energy. Although there physiological reasons for augmenting the digestive capacity of pigs with supplementation of β -glucanase, the responses to supplementation have been inconsistent.

Most studies on β -glucanase supplementation to barley-based diets have been carried with starter pigs(Inbarr and Ogle, 1988 ; Bedford et al., 1992 ; Thacker et al., 1992 ; Inbarr et al., 1993 ; Officer, 1995 ; Baidoo et al., 1998 ; Jensen et al., 1998 ; Defa Li et al., 1999) and growing and finishing pigs(Graham et al., 1986 ; Graham et al., 1989 ; Thacker et al., 1992). With supplementation of β -glucanase, there is an increase in the breakdown of endosperm cell wall components, resulting in more complete digestion of starch and protein in the small intestine(Hesselman and Aman, 1986).

The mechanisms by which β -glucans interfere with digestion and absorption are closely related to their physicochemical properties. β -glucans differ from cellulose in that approximately 30% of the linkages between glucose units are in the form of β (1 \rightarrow 3) and 70% in the form of β (1 \rightarrow 4)(Fleming and Kawakami, 1977). This branched structure prevents compact folding of the molecules and increases the water-holding capacity which results in its characteristic viscosity and gelling properties. The viscosity and gelling properties tend to hinder intestinal motility(Holt et al., 1979) thereby decreasing the mixing of digesta, digestive enzymes and other necessary components required for digestion and absorption(Vahouny and Cassidy, 1985). These

properties may delay or decrease the digestion and absorption of nutrients by increasing the unstirred fluid layer, creating a physical barrier at the absorption surface on the microvilli (Johnson and Gee, 1981).

Graham et al. (1986) found that there was no effect of β -glucanase on nutrient digestibility in 30 to 50kg of pigs fed barley-based diets. Dietary β -glucanase improved numerically ileal digestibility of starch and β -glucan by 1.7 % and 1.4 %, respectively in 80kg of pigs (Graham et al. 1989). Some β -glucan degradation also appears to occur in the stomach, although it has not been resolved whether this reflects enzymic or acid hydrolysis. Several researches conducted with pigs showed no or little improvement on growth performance. Some improvement in growth and feed conversion as well as digestible energy and protein have been reported in pig fed barley diet (Newman et al., 1983; Bedford et al., 1992) in response to enzyme supplementation. However, the entire field of research was open to the criticism of poorly defined enzyme sources, in some cases at least β -glucanase source successful for poultry had little effect on swine (Thacker et al., 1988, 1989, 1991).

(3) Pentosanase

Rye also has been evaluated in diets for growing-finishing pig with and without pentosanase. Thacker et al. (1991) reported a significant reduction in growth rate of pigs fed rye-based diets compared with barely-based diets, whenever diets were supplemented with pentosanase. Bedford et al. (1992) reported that supplementation of pentosanase to a rye-based diet tended to increase viscosity of the small intestine digesta in baby pigs, suggesting pentosan solubilization was occurring at a greater rate than xylan hydrolysis. Supplementation of pentosanase to rye-based diets tended to improve average daily gain and gain/feed of growing-finishing pigs, but there was no effect in barley-based diets (Thacker and Baas, 1996). In studies with weanling pigs fed barley, both daily gain and feed conversion were improved by enzyme supplementation (Thacker et al., 1992) and a reduction in post-weaning diarrhea was reported by Inborr and Ogle (1988).

(4) α -galactosidase

Swine feed compounders can improve the metabolizable energy and N digestibility of soybean meal by means of a feed enzyme that targets the less-well-known anti-

nutritional factors called α -galactosides. Soybean meal is undoubtedly the feed industry's most widely used vegetable protein source. However, it is not an ideal ingredient because it contains high amounts of anti-nutritional factors. The offending compounds include trypsin inhibitors that interfere with protein digestion, as well as non-starch polysaccharides, including arabinoxylans, pectins β -glucans and many more. particularly troublesome are oligosaccharides from the α -galactosides series. The α -galactosides consist of one sucrose unit(fructose-glucose) to which is attached a chain of variable length formed by several galactose units, linked by α -1,6 bonds. Nutritionists have known about these polysaccharides for a long time, however, recently have begun to consider them of significant importance.

Like other anti-nutritional substances, α -galactosides cannot be digested by the pigs endogenous enzymes and no α -1,6 galactosidase activity has been found in the intestinal mucosa(Gitzelamnn and Auricchio, 1965). Only when the micro organisms of the gastrointestinal tract ferment these oligosaccharides can the pigs obtain usable energy from them. However, α -galactosides are only changed into volatile fatty acids. This means their contribution to the total net energy supply of the animal is lower than if they were converted directly into monosaccharides. Moreover, an immediate consequence of this fermentation process is the decrease in the metabolizable energy value of diet that is rich in these oligosaccharides.

Use of feed enzyme clearly show activity over their specific substrates such as like β -glucans or xylans, with best results in the animals that are least able to digest these polysaccharides. Therefore, the use of specific enzymes, capable to act upon the α -galactosides should as an alternative or a complement to physical processing. The use of the α -galactosidase enzyme against the raffinose series of oligosaccharides would allow hydrolysis of the compound to minimize the anti-nutritional effects of these oligosaccharides.

Using α -galactosides as a supplement in conventional corn-soybean meal diet for pigs, there is a clear improvement in the metabolizable energy and N digestibilities of the soybean meal.

III. Conclusion

Exogenous enzymes which, for the purpose of this paper, include phytase, β -glucanase, pentosanase and α -galactosidase, are now extensively used throughout the world as additives in swine diets. The chemical effects of these enzymes are well understand. However, the manner in which their benefits to the swine are brought about is still under debate. Phytase was to increase the availability of plant phytate phosphorus, which reduces phosphorus pollution and allows reductions in the amount of inorganic phosphate used. Also, enzymes have been discovered which have the potential to break down deleterious compounds commonly found in swine rations such as β -glucanase contained in barley and oats and the soluble pentosans found in rye and wheat thus increasing the digestibility of these non-starch polysaccharides. Future research in these area will allow for more efficient use of the current enzymes, development of more efficient future products and development of more thermotolerant enzymes.

Key words : 색인어 : 돼지, phytase, β -glucanase, pentosanase

Literature cited

1. Adeola, O., B. V. Lawrence, A. L. Sutton and T. R. Cline. 1995. Phytase-induced changes in mineral utilization in zinc-supplemented diets for pigs. *J. Anim. Sci.* 73 : 3384-3391.
2. Aman, P. and H. Graham. 1987. Analysis of total and insoluble mixed-linked(1 \rightarrow 3),(1 \rightarrow 4)- β -D-glucan in barley and oats. *J. Agric. Food Chem.* 35 : 704-709.
3. Baidoo, S. K., Y. G. Liu and D. Yungblut. 1998. Effect of microbial enzyme supplementation on energy, amino acid digestibility and performance of pigs fed hullless barley based diets. *Can. J. Anim. Sci.* 78 : 625-631.
4. Bedford, M. R. and A. J. Morgan. 1996. The use of enzymes in poultry diets. *World Poult. Sci.* 52 : 61-68.
5. Bedford, M. R., J. F. Patience, H. L. Classen and J. Inbarr. 1992. The effect of dietary enzyme supplementation of Rye- and barley-based diets on digestion and subsequent performance in weanling pigs. *Can. J. Anim. Sci.* 72 : 97-105.

6. Beers, S. and A. W. Jongbloed. 1992. Effect of supplementary *Aspergillus niger* phytase in diets piglets on their performance and apparent digestibility of phosphorus. *Anim. Prod.* 55 : 425-430.
7. Classen, H. L., T. A. Scott, G. G. Irish P. Huck, M. Swift and M. R. Bedford. 1995. The relationship of chemical and physical measurements to the apparent metabolisable energy of wheat when fed to broiler chickens with and without a wheat enzyme source. In: *Processings of the second european symposium on feed enzymes.* p. 65-69.
8. Classen, H. L., G. L. Campbell, B. G. Rassnagel, R. S. Bhatta and R. D. Reichert. 1985. Studies on the use of hulless barley in chick diets : deleterious effects and methods of alleviation. *Can. J. Anim. Sci.* 65 : 725-733.
9. Corring, T. 1982. Enzyme digestion in the proximal digestive tract of the pig. A review. *Livest. Prod. Sci.* 9 : 581-590.
10. Cromwell, G. L. 1989. Requirements biological availability of calcium, phosphorus for swine evaluated. *Feedstuffs.* 61 : 16-20.
11. Cromwell, G. L., R. D. Coffey and H. J. Monegue. 1993. Phytase(Natuphos®) improves phytase phosphorus utilization in corn-soybean meal for pigs. *J. Anim. Sci.* 71(Suppl. 1) : 165(Abstr.).
12. Cromwell, G. L., R. D. Coffey, G. R. Parker, H. J. Monegue and J. H. Randolph. 1995. Efficacy of a recombinant-derived phytase in improving the bioavailability of phosphorus in corn-soybean meal diets for pigs. *J. Anim. Sci.* 73 : 2000-2208.
13. Defa Li, S. D. Liu, S. Y. Qiao, G. F. Yi, C. Liang and P. A. Thacker. 1999. Effect of feeding organic acid with or without enzyme on intestinal microflora, intestinal enzyme activity and performance of weaned pigs. *Asian-Aus. J. Anim. Sci.* 12 : 411-416.
14. Denbow, D. M., V. Ravindran, E. T., Kornegay, Z. Yi and R. M. Hulet. 1995. Improving phosphorus availability in soybean meal for broilers by supplemental phytase. *Poultry Sci.* 74 : 1831-1842.
15. Elwinger, K. and B. Saterby. 1987. The use of β -glucanase in practical broiler diets containing barley or oats. Effect of enzyme level, type and quality of grain. *Swedish J. Agric. Res.* 17 : 133-140.
16. Farrell, D. J. 1997. Where in the world will we find the ingredients to feed our livestock by the year 2007. *Recent advances in animal nutrition in australia UNE.* pp.136-145.

17. Fleming, M. and K. Kawakami. 1977. Studies of the fine structure of β -glucans of barley extracted at different temperatures. *Carbohydr. Res.* 57 : 15-23.
18. Gitzelmann, R. and S. Auricchio. 1965. The handling of soy alpha galactosides by a normal galctosemic child. *Pediatrics.* 36 : 231-232.
19. Graham, H., J. G. Fadel, C. W. Newman and R. K. Newman. 1989. Effect of pelleting and beta-glucanase supplementation on the ileal and fecal digestibility of a barley-based diet in the pigs. *J. Anim. Sci.* 67 : 1293-1298.
20. Graham, H., K. Hesselman, E. Jonsson and P. Aman. 1986. Influence of beta-glucanase supplementation on digestion of barley-based diet in the pig gastrointestinal tract. *Nutr. Rep. Int.* 34 : 1089-1096.
21. Han, Y. M., F. Yang, A. G. Zhou, E. R. Miller, P. K. Ku, M. G. Hogberg and X. G. Lei. 1997. Supplementation phytases of microbial and cereal sources improve dietary phytate phosphorus utilization by pigs from weaning through finishing. *J. Anim. Sci.* 75 : 1017-1025.
22. Harper A. F., E. T. Kornegay and T. C. Schell. 1997. Phytase supplementation of low-phosphorus growing-finishing pig diets improves performance, phosphorus digestibility, and bone mineralization and reduces phosphorus excretion. *J. Anim. Sci.* 75 : 3174-3186.
23. Hesselman, K. and P. Aman. 1986. The effect of β -glucanase on the utilization of starch and nitrogen by broiler chickens fed on barley of low- or high-viscosity. *Anim. Feed Sci. Technol.* 15 : 83-93.
24. Holt, S., R. C. Heading, D. C. Carter, L. F. Prescott and P. Tothill. 1979. Effect of gel-fibre on gastric emptying and absorption of glucose and paracetamol. *Lancet.* 1 : 636-639.
25. Inbarr, J. and R. B. Ogle. 1988. Effect of enzyme treatment of piglet feeds on performance and post-weaning diarrhoea. *Swed. J. Agric. Res.* 18 : 129-133.
26. Inbarr, J., M. Schmitz and F. Ahrens. 1993. Effect of adding fibre and starch degrading enzymes to a barley/wheat based diet on performance and nutrient digestibility in different segments of the small intestine of early-weaned pigs. *Anim. Feed Sci. Technol.* 44 : 113-127.
27. Jensen, M. S., K. E. Bach Kundsén, I. Inbarr and K. Jakobsen. 1998. Effect of β -glucanase supplementation on pancreatic activity and nutrient digestibility in piglets fed diets based on hulled and hullless barley varieties. *Anim. Feed Sci. Technol.* 72 : 329-345.

28. Johnson, I. T. and J. M. Gee. 1981. Effect of gel-forming gums on the intestinal unstirred layer and sugar transport in vitro. *Gut* 22 : 398-403.
29. Jongbloed, A. W., H. Everts and P. A. Kemme. 1991. Phosphorus availability and requirement in pigs. *Rec. Adv. Anim. Nutr.* 65-80.
30. Jongbloed, A. W., Z. Mooz, P. A. Kemme, C. Geerse and Y. van Der Honing. 1993. The effect of dietary calcium levels on microbial phytase efficacy in growing pigs. *J. Anim. Sci.* 71(Suppl. 1) : 166(Abstr).
31. Lei, X. G., P. K. Ku, E. R. Miller and M. T. Yokoyama. 1993a. Supplementing corn-soybean meal diets with microbial phytase linearly improves phytate phosphorus utilization by weanling pigs. *J. Anim. Sci.* 71 : 3359-3367.
32. Lei, X. G., P. K. Ku, E. R. Miller, D. E. Ullrey, and M. T. Yokoyama. 1993b. Supplemental microbial phytase improves bioavailability of dietary zinc to weanling pigs. *J. Nutr.* 123 : 1117-1124.
33. Lei, X. G., P. K. KU, E. R. Miller, M. T. Yokoyama and D. E. Ullrey. 1993c. Supplementing corn-soybean meal diets with microbial phytase maximizes phytase phosphorus utilization by weanling pigs. *J. Anim. Sci.* 71 : 3368-3375.
34. Marquardt, R., D. Boros, W. Guenter and G. Crow. 1994. The nutritive value of barley, rye, wheat and corn for young chicks as affected by use of a trichoderma reesei enzyme preparation. *Anim. Feed Sci. Technol.* 45 : 363-378.
35. Mroz, Z., A. W. Jongbloed and P. A. Kemme. 1994. Apparent digestibility and retention of nutrients bound to phytate complexes as influenced by microbial phytase and feeding regimen in pigs. *J. Anim. Sci.* 72 : 126-132.
36. Newman, C. W., R. F. Eslick and A. M. El-Negoumy. 1983. Bacterial diastase effect on the feed value of two hullless barleys for pigs. *Nutr. Rep. Int.* 28 : 139-146
37. NRC. 1998. Nutrient requirement of pigs. National Research Council, Academy Press. Washington, D. C.
38. Oberleas, D. 1973. Phytates, in toxicants occurring naturally in foods. p 363. National academy of sciences, Washington, DC.
39. Officer, D. I. 1995. Effect of multi-enzyme supplements on the growth performance of piglets during the pre- and post-weaning periods. *Anim. Feed Sci. Technol.* 56 : 55-65.
40. Peeler, H. T. 1972. Biological availability of nutrients in feeds ; Availability of major mineral ions. *J. Anim. Sci.* 35 : 695-699.

41. Pointillart, A., A. Fourdin and N. Fontaine. 1987. Importance of cereal phytase activity for phytate phosphorus utilization by growing pigs fed diets containing triticale or corn. *J. Nutr.* 117 : 907-913.
42. Pointillart, A., N. Fontaine and M. Thomasset. 1984. Phytate phosphorus utilization and intestinal phosphatases in pigs fed low phosphorus : wheat or corn diets. *Nutr. Rep. Int.* 29 : 473-483.
43. Simons, P. C. M., H. A. J. Versteegen, A. W. Jongbloed, P. A. Kemme, P. Slump, K. D. Bos, M. G. E. Wolters, R. F. Beudeker and G. J. Verschoor. 1990. Improvement of phosphorus availability by microbial phytase in broilers and pigs. *Br. J. Nutr.* 64 : 525-540.
44. Steinfeldt, S., A. Mullertz and J. F. Jensen. 1998. Enzyme supplementation of wheat-based diets for broilers. 1. Effect on growth performance and intestinal viscosity. *Anim. Feed Sci. Technol.* 75 : 27-43.
45. Thacker, P. A. and T. C. Baas. 1996. Effects of gastric pH on the activity of exogenous pentosanase and the effect of pentosanase of growing-finishing pigs. *Anim. Feed Sci. Technol.* 63 : 187-200.
46. Thacker, P. A., G. L. Campbell and J. W. D. GrootWassink 1992. The effect of salinomycin and enzyme supplementation on nutrient digestibility and performance of pigs fed barley - or rye-based diets. *Can. J. Anim. Sci.* 72 : 117-125.
47. Thacker, P. A., G. L. Campbell and J. W. D. GrootWassink. 1988. The effect of beta-glucanase supplementation on the performance of pigs fed hullless barley. *Nutr. Rep. Int.* 38 : 91-99.
48. Thacker, P. A., G. L. Campbell and J. W. D. GrootWassink. 1991. The effect of enzyme supplementation on the nutritive value of rye-based diets for swine. *Can. J. Anim. Sci.* 71 : 489-496.
49. Thacker, P. A., G. L. Campbell and J. W. D. GrootWassink. 1989. The effect of sodium bentonite on the performance of pigs fed barley-based diets supplemented with beta-glucanase. *Nutr. Rep. Int.* 40 : 613-619.
50. Um J. S., I. K. Paik, M. B. Chang and B. H. Lee. 1998. Effects of microbial phytase supplementation to diets with low non-phytase phosphorus levels on the performance and bioavailability of nutrients in laying hens. *Asian-Aus. J. Anim. Sci.* 12 : 203-208.
51. Vahouny, G. V. and M. M. Cassidy. 1985. Dietary fibre and absorption of nutrients. *Proc. Soc. Exp. Biol. Med.* 180 : 432-446.

52. Veum, T. L. 1996. Use of microbial phytase in corn-soybean meal and grain sorghum-canola meal diets for growing-finishing swine diets. In : M. B. Coehlo and E. T. Kornegay(Ed.). Phytase in animal nutrition and waste management. P. 365-380. BASF Corp., Mount Olive, NJ.
53. Wyatt, C. L., E. Morgan and M. R. Bedford. 1997. Utilizing feed enzymes to enhance the nutritional value of corn-based broiler diet. *Poult. Sci.* 76 : 39-39.
54. Wyatt, C. L., M. R. Bedford and L. A. Waldron. 1999. Role of enzymes in reducing variability in nutritive value of maize using the ileal digestibility method. *Proc. Aust. Poult. Sci.* 11 : 108-111.
55. Yi, Z., E. T. Kornegay, V. Ravindran, V. M. D. Lindemann and J. H. Wilson. 1996. Effectiveness of Natuphos phytase in proving the bioavailability of phosphorus and nutrients in soybean meal-based semipurified diets for young pigs. *J. Anim. Sci.* 74 : 1601-1611.
56. Young, L. G., M. Leunissen and J. L. Atkinson. 1993. Addition of microbial phytase to diets of young pigs. *J. Anim. Sci.* 71 : 2147-2150.