# The Synthesis of 1,4,7-Triazacyclononane Conjugated Amyloid-phillic Compound and Its Binding Affinity to the $\boldsymbol{\beta}$-Amyloid Fibril 

Jongmin Kang* and Jee Hye Jo<br>Department of Applied Chemistry, Sejong linversity, Seoul 143-747, Korea<br>Received./uly 7, 2003

Key Words: Alzheimers disease, $\beta$-Amyloid fibril, Amyloid-philic molecule

The pathological hallmark of $A D$ is a deposition of anyloid plaques in the brains of patients. $\beta$-anyloid protein is a major protein component of Alzheimer's plagues. When aggregated into amyloid fibrils, the peptide is toxic to neuronal cells. ${ }^{1.2}$ The neurotoxicity of $A \beta$ fibril is closely related to the $\beta$-sheet conformation and aggregation of $\mathrm{A} \beta$ peptide. ${ }^{3}$ Various compounds which have affinity for $\beta$ anyloid fibril have shown ability of preventing neurotoxicity of $\beta$-amyloid fibril and inhibiting aggregation of $\beta$-amyloid fibril." Therefore, the development of new compounds which have affinity for the $\beta$-anyloid fibril would lead to the new compounds that could have therapeutic effects on AD. Previously, we generated new anyloid-phillic amide derivative of Chrysamine G I and found that this compound protect human astrocyte cells against $\mathrm{A} \beta$-induced toxicity. ${ }^{5.6}$ As conjugation of amyloid-philic molecules with suitable metal chelating ligands could lead to new diagnostic molecules for in vivo quantification of anyloid deposition ${ }^{7.8}$ and new probes for amyloid structure. ${ }^{9,10}$ we designed the compound 2, which was conjugate of 1,4,7-triazacyclononane and the amyloid-philic compound $I$. Here, we would like to report the synthesis of compound 2 and its binding property of $\beta$-anyloid tibril. The synthesis of compound 2 was achieved by combining three fragments the biphenyl amine 9 , isophthalic acid 5 and 1,4, 7-triazacyclononane $\mathbf{2 0}$.


The synthesis of isophthalic acid part 5 commenced with esterfication of isophthalic acid in methanol with HCl to produce the monoester 4 in $38 \%$ yield. Treatment of the monoester 4 with oxalyl chloride afforded the acyl chloride 5 in $95 \%$ yield (Scheme 1).

The synthesis of biphenyl amine part 9 began with the reaction between 4,4-dinitro-2-biphenylamine 6 and 6bromohexanoyl chloride 7 in dried methylene chloride to afford compound 8 in $92 \%$ yield. Hydrogenation of the compound 8 at elevated pressure with $\mathrm{Pd} / \mathrm{C}$ gave the biphenyl amine 9 in $99 \%$ yield (Scheme 2).

The synthesis of 1,4 -bis(tert-butylcarbonate)-1,4,7-triazacyclononane $\mathbf{2 0}$ started from tosylation of diethylenetriamine 12. Therefore, the reaction between diethylenetriamine and tosylchloride gave the tosylated diethylenetriamine 13 in 93 $\%$ yield. Then tosylated diethylenetriamine 13 was refluxed

Scheme 1


6


Scheme 2


Scheme 3
in absolute ethanol with sodium methoxide to give sodium metallated compound $\mathbf{1 4}$ in $96 \%$ yield. Metallated compound 14 was reacted with tosylated ethylene glycol 11 in hot and dried DMF to give tritosylated-1,4,7-triazacyclononane 15 in $90 \%$ yield. Detosylation of the compound 15 was performed in hot sulfuric acid for 48 hours to give the 1,4,7-triazacyclononane 16 in $60 \%$ yield. The 1.4.7-triazacyclononane 16 was reacted with $\mathrm{N}, \mathrm{N}$ '-dimethylformamide dimethylacetal to give the protected 1,4.7-triazacyclononane 17 in $85 \%$ yield. The protected 1,4.7-triazacyclononane 17 was reacted with benzyl bromide in dried THF to give the 1-benzyl-1,4,7-triazacyclononane 18 in quantitative yield. Then the compound 18 was reacted with dibutyldicarbonate in methylene chloride to give the compound 19 in
quantitative yield. Debenzylation of the compound 19 with $10 \% \mathrm{Pd} / \mathrm{C}$ in acetic acid gave the desired 1,4 -bis(tert-butylcarbonate)-1.4,7-triazacyclononane 20 in $81 \%$ yield (Scheme 3).

The coupling of three fragments was accomplished by the reaction of the biphenyl compound 9 and the acyl chloride 5 to afford the molecule 21 in $30 \%$ yield. Refluxing the compound 20 and 21 in acetonitrile gave the compound 22 in $71 \%$ yield. Deprotection reaction of 22 in 4 M HCl dioxane and neutralization of the deprotected compound with 6 N LiOH gave the final compound $\mathbf{2}$ as a lithiated form (Scheme 4).

As lithiated salt form, the compound 2 was readily soluble in water. For the binding study, $80 \mu \mathrm{~L}$ of various concen-


Scheme 4


Figure 1. The Scatchard analysis of the compound 2 for the $\beta$ amyloid fibril 1-40.
trations of the compound $\mathbf{2}$ in water $(5-100 \mu \mathrm{M})$ were added to the $20 \mu \mathrm{~L}$ of $100 \mu \mathrm{M} \beta$-amyloid 1-40 fibril. After 2 hours at $37{ }^{\circ} \mathrm{C}$, the incubating eppendorf tubes were centrifuged ( $16,000 \mathrm{rpm} .20 \mathrm{~min}$ ) to spin down the $\beta$-amyloid fibril and the compound bound to the $\beta$-amyloid fibril. Then the concentrations of unbound compound in the solution were measured by UV spectrometer at 290 nm . Figure 1. shows Scatchard analysis of the binding of 2 to the $\beta$-amyloid 1-40 fibril. Like compound $\mathbf{1 , 2}$ has two binding sites. The higher affinity binding site appears to have a $\mathrm{K}_{\mathrm{d}}$ of $2.98 \mu \mathrm{M}$ and a $\mathrm{B}_{\text {nuis }}$ of 0.58 moles per mole of anyloid 1-40 peptide. The lower affinity binding site appears to have a $\mathrm{K}_{\mathrm{d}}$ of $88.50 \mu \mathrm{M}$ and a $\mathrm{B}_{\text {max }}$ of 5.60 moles per mole of $\beta$-amyloid 1-40 peptide. As the compound 1 showed $\mathrm{K}_{\mathrm{d}}=0.13, \mathrm{~B}_{\text {tuax }}=0.36$ at higher affinity binding site and $\mathrm{K}_{\mathrm{d}}=10.11 \mathrm{~B}_{\text {nax }}=0.60$ at lower affinity binding site, the compound 2 has comparable affinity with the compound 1 or Chrysamine G..$^{56}$ The coupling of 1,4,7-triazacyclononane to the compound $\mathbf{1}$ did not affect the binding affinity of the molecule 2 . The molecule 2 could be used to as a new diagnostic molecule for quantification of amyloid deposition or new probes for amyloid structure after metal chelation. The possibilities are currently under investigation.

## Experimental Section

Monomethyl isophthalate 4. To a solution of 2 g isophthalic acid in 60 mL of methyl alcohol was added 3 drops of concentrated HCl and refluxed for 2.5 hours. After evaporation of methyl alcohol the reaction mixture was chromatographed on silicagel with $7 \% \mathrm{MeOH}-\mathrm{MC}$ to afford 839 mg product 4 in $38 \%$ yield. ${ }^{\prime} \mathrm{H}$ NMR ( $200 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $8.8(\mathrm{t}, \mathrm{IH}, J-1.6) 8.3$ (dd, 2H, $J-8.0, J-1.6) 7.6(\mathrm{t}, \mathrm{IH}, J$ $-8.0) 4.0(\mathrm{~s}, 3 \mathrm{H}) \mathrm{MS}$ calculated for $\mathrm{C}_{9} \mathrm{H}_{8} \mathrm{O}_{4} 180 . \mathrm{I}$ found for $179.0(\mathrm{M}-\mathrm{H})$.
Monomethylester isophthaloyl chloride 5. To a solution of 730 mg 4 in 15 mL of dichloromethane was added 3.5 mL of oxalyl chloride and $150 \mu \mathrm{~L}$ of $5 \%$ DMF-MC. After stirring an hour, evaporation of all solvent and oxalyl chloride at reduced pressure produced the compound 5 in 95 \% yield. ${ }^{1} \mathrm{H}$ NMR ( $200 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) 8.6 (t. $\mathrm{IH}, J-\mathrm{I} .6$ ) 8.2 $(\mathrm{m}, 2 \mathrm{H}) 7.5(\mathrm{t} .1 \mathrm{H}, J-7.8) 3.9(\mathrm{~s}, 3 \mathrm{H})$.
Compound 8. To a solution of 2 g 4.4-dinitro-2-
biphenylaniline and 1.3 mL triethylamine in 80 mL of dried dichloromethane was added 1.4 mL 6-bromohexanoyl chloride and stirred for 4 hours. Then the reaction mixture was extracted with 100 mL of water three times. The organic layer was dried with $\mathrm{MgSO}_{4}$ and filtered. Evaporation of filtrate and silicagel chromatography (hexane: dichloromethane $=4: 1$ ) gave 550 mg of product 8 in $92 \%$ yield. ${ }^{1} \mathrm{H}$ NMR ( $200 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $9 . \mathrm{I}(\mathrm{d} . \mathrm{IH}, J=2.2) 8.4(\mathrm{~d}, 2 \mathrm{H}, J=$ 6.8) $8.1(\mathrm{dd}, \mathrm{IH}, J=8.4, J=2.2) 7.6(\mathrm{~d}, 2 \mathrm{H}, J=6.8) 7.4(\mathrm{~d}$, $1 \mathrm{H}, J=8.4) 7.0(\mathrm{~s}, 1 \mathrm{H}) 3.4(\mathrm{t}, 2 \mathrm{H}, J=6.5) 2.3(\mathrm{t}, 2 \mathrm{H}, J=7 . \mathrm{I})$ 1.8 (quin, $2 \mathrm{H}, J=7.0$ ) 1.7 (quin, $2 \mathrm{H}, J=8.0$ ) 1.5 (quin, 2 H , $J=6.6$ ) MS calculated for $\mathrm{C}_{18} \mathrm{H}_{18} \mathrm{BrN}_{3} \mathrm{O}_{5} 435.0$ found for 434.8.

Compound 9. A solution of $\lg$ compound 8 and 250 mg $10 \% \mathrm{Pd} / \mathrm{C}$ in $20 \%$ methanol in dichloromethane was stirred with $\mathrm{H}_{2}$ gas at elevated pressure for 2 hours. The solution was filtered through celite and the filtrate was evaporated to afford 862 mg product 9 in $99 \%$ yield. 'H NMR ( 200 MHz , $\left.5 \% \mathrm{CD}_{3} \mathrm{OD}-\mathrm{CDCl}_{3}\right) 7.5(\mathrm{~d}, \mathrm{IH}, J=2.3) 7.0(\mathrm{~d}, 2 \mathrm{H}, J=8.3)$ $6.9(\mathrm{~d}, \mathrm{IH}, J=8.2) 6.7(\mathrm{~d}, 2 \mathrm{H}, J=8.3) 6.5(\mathrm{dd}, 1 \mathrm{H}, J=8.2, J$ $=2.3) 3.3(\mathrm{t}, 2 \mathrm{H}, J=5.7) 2.1(\mathrm{t}, 2 \mathrm{H}, J=7.0) 1.7(q u i n, 2 \mathrm{H}, J$ $=6.6$ ) 1.5 (quin, $2 \mathrm{H}, J=7.3$ ) 1.3 (quin, $2 \mathrm{H}, J=6.4$ ) MS calculated for $\mathrm{C}_{18} \mathrm{H}_{22} \mathrm{BrN}_{3} \mathrm{O} 375.1$ found for 296.1 (M-HBr).

Ditosyl ethylene glycol 11. To a solution of 29 g triethylamine and 55 g tosyl chloride in 200 mL of dichloromethane at $0{ }^{\circ} \mathrm{C}$ was added 8 g ethylene glycol for an hour. Then the reaction mixture was stirred for 24 hours. The reaction mixture was poured into 2 L water and extracted with 100 mL of dichloromethane 5 times. The organic layer was dried with $\mathrm{MgSO}_{4}$ and filtered. Evaporation of dichloromethane produced 47 g of compound 11 in $97 \%$ yield. ${ }^{1} \mathrm{H}$ NMR ( $200 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) 7.7 (d, $4 \mathrm{H}, J=8.2$ ) $7.3(\mathrm{~d}, 4 \mathrm{H}, J=$ 8.2) $4.2(\mathrm{~s}, 4 \mathrm{H}) 2.5(\mathrm{~s}, 6 \mathrm{H})$.

Tritosyl diethylenetriamine 13. To a solution of 25.2 g triethylamine and 8 g diethylenetriamine in 350 mL dichloromethane was added 45.8 g tosylchloride at $0^{\circ} \mathrm{C}$. Then the reaction mixture was stirred for 12 hours. Evaporation of the solvent and recrystallization in methanol gave 41.4 g of product 13 in $93 \%$ yield. ${ }^{~} \mathrm{H}$ NMR $(200 \mathrm{MHz}$, $\left.\mathrm{CDCl}_{3}\right) 7.6(\mathrm{~d}, 4 \mathrm{H}, j-8.2) 7.3(\mathrm{~d}, 4 \mathrm{H}, J-8.2) 5.1(\mathrm{t}, 2 \mathrm{H})$ 3.1 (t. $8 \mathrm{H} . J-6.8) 2.4$ (s. 9 H ).

Compound 14. To a solution of 32 g of the compound 13 in 310 mL ethanol was added 23 g of $28 \% \mathrm{MaOMe}$ and refluxed for 2 hours. Filtration of the solid afforded the compound 14 in $96 \%$ yield. 'H NMR ( $200 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) 7.46 (d, 6H, $J-8.0$ ) 7.28 (d. $2 \mathrm{H}, J-8.0$ ) 7.1 I (d. $4 \mathrm{H}, J-$ $8.0) 2.83(\mathrm{~m}, 4 \mathrm{H}) 2.62(\mathrm{~m}, 4 \mathrm{H}) 2.83(\mathrm{~m}, 4 \mathrm{H}) 2.62(\mathrm{~m}, 4 \mathrm{H})$ 2.34 ( $\mathrm{s}, 3 \mathrm{H}$ ).

Tritosyl 1,4,7-triazacyclononane 15. To a solution of 28 g of the compound 14 in 180 mL dried DMF at $100^{\circ} \mathrm{C}$ was added 17 g of the compound 11 in 150 mL dried DMF for 2 hours. Then the reaction mixture was stirred for 2 hours more at $100^{\circ} \mathrm{C}$. Evaporation of the DMF at reduced pressure and recrystallization in water gave the 24.5 g of the compound 15 in $90 \%$ yield. 'H NMR ( $200 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ) $7.65(\mathrm{~d}, 6 \mathrm{H}, J-8.0) 7.28(\mathrm{~d}, 6 \mathrm{H}, J-8.0) 3.40(\mathrm{~s}, 12 \mathrm{H}) 2.4 \mathrm{I}$ ( s .9 H ).
$1,4,7$-Triazacyclononane 16 . The solution of 25.6 g of the compound 15 in 55 mL concentrated $\mathrm{H}_{2} \mathrm{SO}_{4}$ was heated at $95^{\circ} \mathrm{C}$ for 48 hours. Then the reaction misture was slowly poured into 300 mL of cold diethyl ether and ethanol (1:1) The solid was filtered and dissolved in water and pH was adjusted until 8 with 6 N KOH . Then all of the water was evaporated and the residue was dissolved in small amount of methanol. The undissolved KOH was removed by filtration. This method gave the product 16 in $60 \%$ yield. ${ }^{1} \mathrm{H}$ NMR ( $200 \mathrm{MHz} . \mathrm{CDCl}_{3}$ ) 2.7 (s. 12 H ) 1.9 (s. lH ).

Compound 17. To a solution of 0.2 g compound 16 in 18 mL dried acetonitrile was added $11 \mathrm{~mL} \mathrm{~N}, \mathrm{~N}$ '-dimethy lformamide dimethyl actal. The reaction mixture was stirred for 3 hours at $85^{\circ} \mathrm{C}$. Evaporation of the solid gave 0.17 g of the product $17 \mathrm{in} 85 \%$ yield. ${ }^{1} \mathrm{H}$ NMR ( $200 \mathrm{MHz} . \mathrm{CDCl}_{3}$ ) 5.0 (s. $1 \mathrm{H}) 2.6-3.0(\mathrm{~m}, 12 \mathrm{H})$.
1-Benzyl-1,4,7-triazacyclononane 18. To a solution of 6.6 g of the compound 17 in 65 mL dried THF was added 6.7 mL benzyl bromide. Then the reaction mixture was stirred for an overnight. After the solvent was evaporated. 55 mL of water and 2 g NaOH was added to the reaction misture and refluxed for 2 days. The reaction misture was extracted with chloroform and the organic layer was dried with $\mathrm{MgSO}_{4}$. Filtration and evaporation of the filtrate gave the product 18 in quantitative yield. The product obtained in this stage was crude. Without further purification this material was used for the next reaction.

1,4-Bis(tert-butylcarbonate)-7-benzyl-1,4,7-triazacyclononane 19. To a solution of 8.9 g of the compound 18 in 250 mL dichloromethane was added 35 g dibutyldicarbonate. Evaporation of the solvent and chromatography (hexane ethyl acetate $=8: 1$ ) on silicagel gave product 19 in quantitative yield. ${ }^{1} \mathrm{H}-\mathrm{NMR}\left(200 \mathrm{MHz}, \mathrm{CDCl}_{3}\right) 7.2(\mathrm{~m} .5 \mathrm{H})$ 3.63 (s. 2H) 3.43 (t. 4 H ) $3.14(\mathrm{~m} .4 \mathrm{H}) 2.61(\mathrm{~m}, 4 \mathrm{H}) 1.38(\mathrm{~s}$. 9 H ).

1,4-Bis(tert-butylcarbonate)-1,4,7-triazacyclononane 20. To a solution of Ig of the compound 19 in 25 mL acetic acid was added $0.1 \mathrm{~g} 10 \% \mathrm{Pd} / \mathrm{C}$ and stirred under $\mathrm{H}_{2}$ gas atmosphere for 3 hours. The reaction mixture was filtered through celite and the filtrate was evaporated. The residue was dissolved in small amount of water and pH was adjusted until 12 with 1 N NaOH . The water layer was extracted with 50 mL chloroform 3 times. The organic layer was dried with $\mathrm{MgSO}_{4}$. Filtration and evaporation of the filtrate gave 0.6 g of the compound 20 in $81 \%$ yield. ${ }^{l} \mathrm{H}-\mathrm{NMR}$ ( 200 MHz . $\left.\mathrm{CDCl}_{3}\right) 3.47(\mathrm{t} .4 \mathrm{H}) 3.32(\mathrm{~b} .4 \mathrm{H}) 2.97(\mathrm{~m} .4 \mathrm{H}) 1.47(\mathrm{~s} .9 \mathrm{H})$.

Compound 21. To a solution of 1.0 g of the compound 9 and 1.1 mL triethylamine in 10 mL chloroform was added compound 5 . The reaction mixture was stirred for an hour. Evaporation of the solvent and chromatography on the silicagel (methanol: dichloromethane $=7: 493$ ) gave 578 mg of the product 21 in $30 \%$ yield. ${ }^{1} \mathrm{H}$ NMR ( $10 \% \mathrm{CD}_{3} \mathrm{OD}$ $\mathrm{CDCl}_{3}$ ) 8.5 (s.2H) $8.0(\mathrm{td} .4 \mathrm{H} . J=7.8 \mathrm{~J}=1.3) 7.9$ (d. $1 \mathrm{H} . J$ $=2.0) 7.8(\mathrm{dd} .1 \mathrm{H} . J=8.4, J=2.0) 7.7(\mathrm{~d} .2 \mathrm{H} . J=8.5) 7.5$ (td. $2 \mathrm{H} . J=7.8 . J=1.3$ ) 7.3 (d. $2 \mathrm{H} . J=8.5$ ) 7.2 (d. $1 \mathrm{H} . J=$ $8.4) 3.8$ (s. 6 H ) 3.3 (t. $2 \mathrm{H} . J=6.7$ ) 2.2 (t. $2 \mathrm{H} . J=7.1$ ) 1.8 (quin. $2 \mathrm{H} . J=7.3$ ) 1.5 (quin. $2 \mathrm{H} . J=7.3$ ) 1.3 (quin. $2 \mathrm{H} . J=$
5.9) MS calculated for $\mathrm{C}_{36} \mathrm{H}_{34} \mathrm{BrN}_{3} \mathrm{O}_{7} 699.1$ found for 619.9 (M-Br).

Compound 22. To a solution of 550 mg of the compound 21 and 0.3 mL triethylamine in 20 mL of actonitrile was added 259 mg of the compound $\mathbf{2 0}$. The reaction mixture was refluxed for 45 hours. Evaporation of the solvent and chromatography on the silicagel ( $4 \%$ methanol in dichloromethane) afforded 113 mg of the compound 22 in $71 \%$ yield. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) 8.5$ (d. $\left.2 \mathrm{H}, J=1.6\right) 8.2$ (s. 1 H$) 8.1$ (m. 4H) 7.9 (d, $2 \mathrm{H} . J=8.2$ ) 7.5 (td, $2 \mathrm{H} . J=7.9 . J=1.6$ ) 7.4 (d. IH. $J=7.8$ ) 7.3 (d. $2 \mathrm{H} . J=8.2$ ) 7.2 (d. IH. $J=8.4$ ) 3.9 (s, $6 \mathrm{H}) 3.4(\mathrm{~s}, \mathrm{br} .4 \mathrm{H}) 3.1(\mathrm{~s}, 4 \mathrm{H}) 2.5(\mathrm{~s}$, br. 4 H$) 2.4(\mathrm{~m}, 2 \mathrm{H}) 2.2$ (m. 2 H ) 1.5 (m. 6 H ) $1.4(\mathrm{~m}, 18 \mathrm{H})$ MS calculated for $\mathrm{C}_{52} \mathrm{H}_{64} \mathrm{~N}_{6} \mathrm{O}_{11} 949.1$ found for $950.1\left(\mathrm{M}+\mathrm{H}^{+}\right)$.

Compound 23.30 mg of the compound 22 was dissolved in 4 M HCl in dioxane ( 0.4 mL ) and stirred for an hour. Evaporation of the solvent gave 26 mg of the product 23 in $99 \%$ yield. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CD}_{3} \mathrm{OD}\right) 8.6(\mathrm{t} .2 \mathrm{H} . J=1.5) 8.2(\mathrm{t} .4 \mathrm{H}$, $J=7.7) 8.0(\mathrm{~d}, 1 \mathrm{H} . J=2.0) 7.8(\mathrm{~d}, 2 \mathrm{H}, J=8.6) 7.7(\mathrm{td}, 2 \mathrm{H} . J$ $=7.7 . J=1.5) 7.6(\mathrm{~d}, 1 \mathrm{H}, J=2.1) 7.5(\mathrm{~d} .2 \mathrm{H} . J=8.6) 7.4(\mathrm{~d}$, $1 \mathrm{H} . J=4.4) 4.0(\mathrm{~d}, 6 \mathrm{H}, J=0.8) 3.6(\mathrm{~s} .4 \mathrm{H}) 3.3(4 \mathrm{H}) 3.0(\mathrm{~m}$. $4 \mathrm{H}) 2.8(\mathrm{t} .2 \mathrm{H} . J=7.9) 2.4(\mathrm{t} .2 \mathrm{H} . J=6.9) 1.7(\mathrm{~m}, 4 \mathrm{H}) 1.3$ (m. 2 H )

Compound 2. To a solution of 20 mg of the compound 23 was added 60 mL of 6 N LiOH and stirred for an overnight. Filtration of the precipitated solid afforded 17 mg of lithiated product 2 in $96 \%$ yield. ${ }^{1} \mathrm{H}$ NMR ( $\left.\mathrm{CD}_{3} \mathrm{OD}\right) 8.5$ (m, 2H) 8.2 (d. $2 \mathrm{H} . J=7.6$ ) 8.1 (d. $2 \mathrm{H} . J=7.6$ ) $8.0(\mathrm{~s} .1 \mathrm{H}) 7.8$ (d. $2 \mathrm{H} . J=$ $8.5) 7.7(\mathrm{~d} .1 \mathrm{H}, J=8.8) 7.5(\mathrm{t}, 2 \mathrm{H} . J=7.7) 7.45(\mathrm{~d}, 2 \mathrm{H}, J=$ $6.6) 7.4(\mathrm{~d} .1 \mathrm{H} . J=6.4) 2.6(\mathrm{~m} .12 \mathrm{H}) 2.3(\mathrm{t}, 2 \mathrm{H}, J=7.3) 1.5$ (m. 4 H ) $1.3(\mathrm{~m} .2 \mathrm{H})$ MS calculated for $\mathrm{C}_{40} \mathrm{H}_{42} \mathrm{Li}_{2} \mathrm{~N}_{6} \mathrm{O}_{7} 732.3$ found for $733.3\left(\mathrm{M}+\mathrm{H}^{+}\right)$.

Acknowledgement. This study was supported by a grant of the Korea Health 21 R\&D Project, Ministry of Health \& Welfare, Republic of Korea. (01-PJ1-PG3-21500-0017)

## References

1. Harrigan, M. R;; Kunkel. D. D.; Nguyen. L. B.; Malouf, A. T. Newobiol. Aging 1995. 16.779.
2. Pike. C. T.: Walencewicz-wassermant. A. T.: Komoski. J.: Cribbs. D. H.: Glabe. C. G.: Cotman. C. W. J. Nemochen. 1995. 64. 253.
3. Simmons. L. D.: May, P. C.: Tomaselli, K. T.: Rydel, R. E.: Fuson. K. S.: Brigham, E. F.; Wright, S.: Lieberburg. I.; Becher, G. W.; Brems. D. N.: Li, W. Y. Mol. Pharmacol. 1994, 45. 373.
4. Schenk. D. B.: Rydel. R. E.: May. P:Little. S.: Panetta. T.: Lieberburg. I.: Sinha. S. J. Med. Chem. 1995. 38 (21). 4141 .
5. Kang. J.: Han. K. Bull. Korean Chent. Soc. 2001. 22 (10). 1065.
6. Kang, J.: Park, J. E.; Jo, S. Bull. Korean Chem. Soc. 2002, 23(3). 363.
7. Han. H., Cho, C -G.: Lansbury. P. T.. Jr J. Am. Chem. Soc. 1996. 118. 4506.
8. Dezutter. N. A.: Dom. R. T.: De Groot. T. J.: Bormans. G. M.: Verbruggen. A. M. Eur J. Nucl. Med 1999. 11. 1392.
9. Klunk. W. E.; Debnath. M. L.; Pettergrew. J. W. Nemobiology of Aging 1994. 15(6). 691.
10. Ashbum, T. T:; Han. H.: McGuinness, B. F.: Lansbury, P. T., Jr. Chemistry \& Biology 1996. 3. 351.
