

Synthesis of C₄-Modified Acanthoic Acid Analogs and Their Biological Evaluation as Nitric Oxide Inhibitors

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Synthesis and biological evaluation of a series of C₄-modified acanthoic acid analogs are reported. Among them, the analog **7** and **10** exhibit potent cellular inhibitory activity in NO inhibition assay.

Key words: Acanthoic acid analog, NO inhibition

INTRODUCTION

Nitric Oxide (NO) which is produced by L-arginine oxidation, catalyzed by nitric oxide synthase (NOS), is an important bioregulator and ubiquitous biomessenger existing in a variety of organisms (Palmer *et al.*, 1987; Pfeiffer *et al.*, 1999). NOS bring about a five-electron oxidation of L-arginine to produce nitric oxide and L-citrulline as shown in Fig. 1. To date three NOS isoforms have been discovered and two of them, endothelial NOS and neuronal NOS, are known to be constitutive and involved in neurotransmission and vasodilatation, respectively (Rees *et al.*, 2002; Bhme *et al.*, 1993). The third isoform, which is induced by inflammatory cytokines, is concerned with host defense (White and Marletta, 1992).

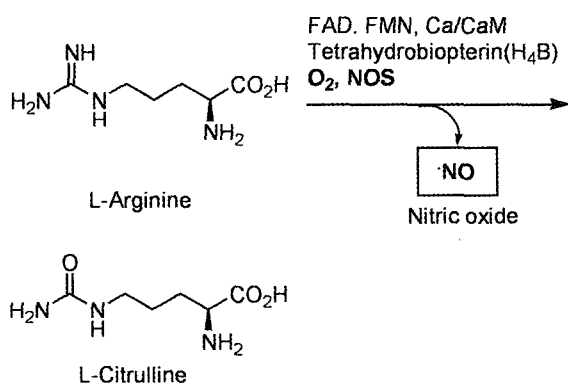


Fig. 1. Biosynthetic pathway of nitric oxide

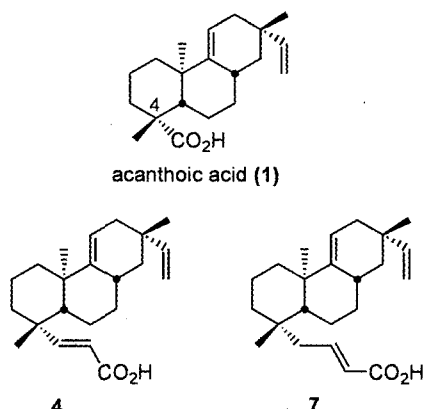
An excessive production of nitric oxide by iNOS leads to inflammatory diseases such as arthritis and inflammatory bowel disease (Kilbourn *et al.*, 1990; Jang *et al.*, 1997; Miller *et al.*, 1993). Thus, a variety of structural analogs of natural products have been synthesized and evaluated for the treatment of inflammatory diseases in animal models and in clinical trials. Acanthoic acid, isolated from the root bark of *Acanthopanax koreanum* Nakai, has recently been reported to suppress Tumor Necrosis Factor- α (TNF- α) and Interleukin-1 β (IL-1 β) selectively (Kang *et al.*, 1996). More recently, we have also reported anti-inflammatory effects of acanthoic acid analogs, which exhibit excellent COX-2 and NO inhibitory activities (Suh *et al.*, 2001, 2004). In particular, our previous work revealed that the length of the linker between C₄ and the terminal carboxyl group plays an important role for the anti-inflammatory effects of the acanthoic acid analogs.

As an extension of studies on acanthoic acid analogs, we have focused on the linker optimization of the C₄-chain by investigating the effect of the linker length and the terminal functional groups on the NO inhibitory effects. We have diversified the C₄-chain on the structural basis of the analogs **4** and **7**, which exhibited NO inhibitory activities of 45.1 and 77.9 inhibition percent at 40 μ M concentration, respectively (Fig. 2). We herein describe synthesis of the C₄-chain modified acanthoic acid analogs as well as evaluation of their inhibitory activities in NO generation in Raw 264.7 cells.

CHEMISTRY

The syntheses of acanthoic acid analogs are outlined in Schemes, 1-3. The syntheses of the analogs **4** and **7** are

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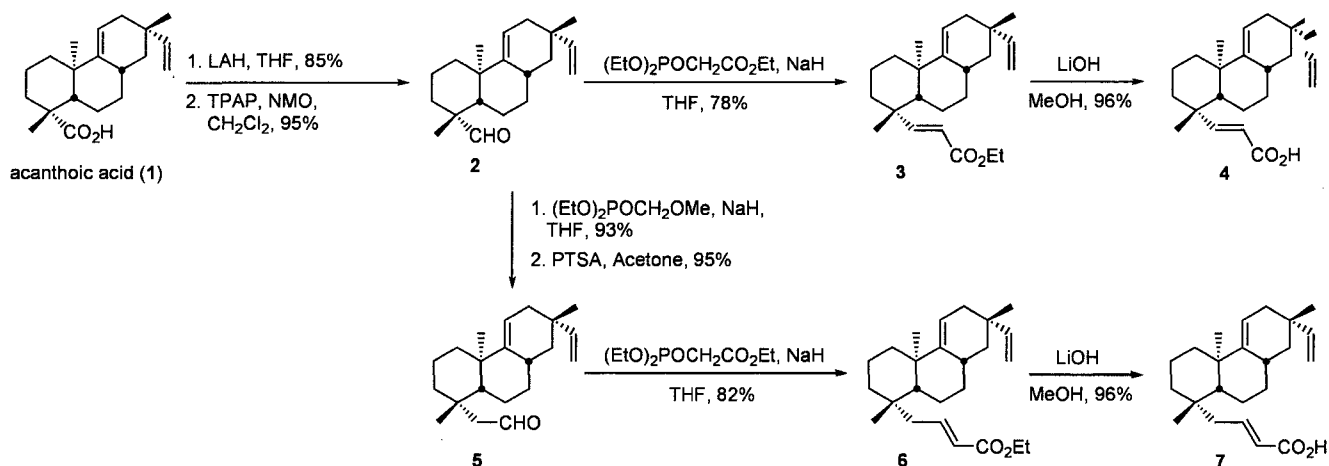
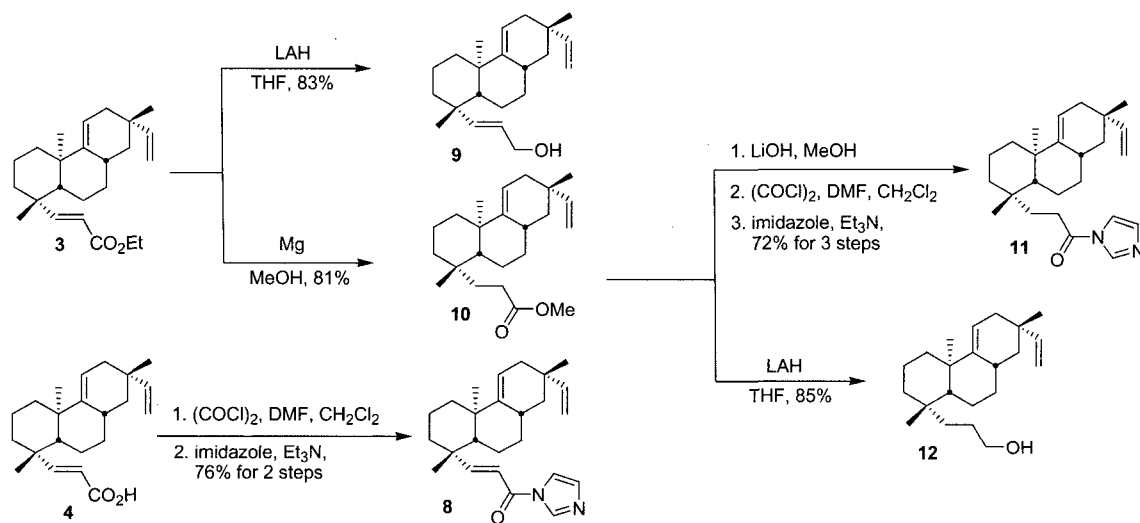

Fig. 2. Structures of acanthonic acid and acanthonic acid analogs

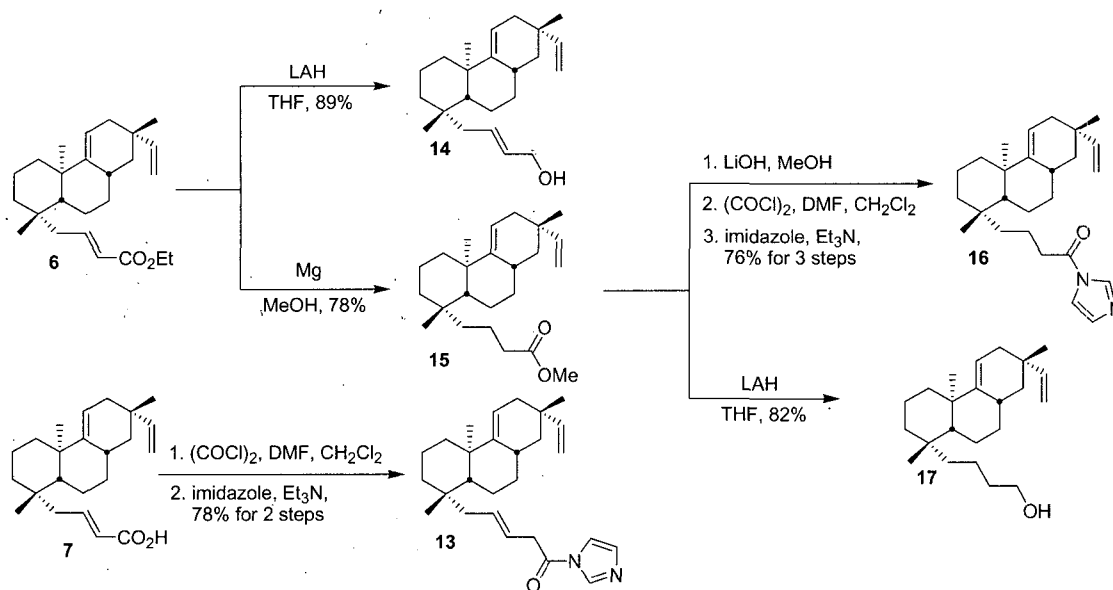
presented in Scheme 1. LAH reduction of acanthonic acid (1) and TPAP oxidation of the resulting alcohol provided the aldehyde 2, which were transformed into the analog 4

by olefination and ester hydrolysis. One carbon homologation of the aldehyde 2 and hydrolysis of the resulting enol ether provided the aldehyde 5, which were transformed into the analog 7 by the same procedures for the analog 4.

The analogs 4 and 7 were transformed into the corresponding alcohol or amide and then reduced to the analogs 11, 12, 16, and 17 by selective olefin reduction with magnesium as shown in Scheme 2 and 3. Treatment of the analog 4 with oxalyl chloride, followed by amidation with imidazole provided the analog 8. LAH reduction of the analog 3 provided the analog 9 while conjugated reduction with magnesium of the analog 3 provided the analog 10, which was transformed into the analog 11 by sequential ester hydrolysis and amidation. Direct LAH reduction of the analog 10 afforded the analog 12.

The analogs 13-17 were prepared from the analogs 6 and 7 as shown in Scheme 3. The synthetic pathways for


Scheme 1. Synthesis of the analogs 4 and 7

Scheme 2. Synthesis of the analogs 8-12



Scheme 3. Synthesis of the analogs, 13-17

the analogs **13-17** are same as those for the analogs **8-12**.

Spectral data for representative analogs

Spectral data for the analog **9**

IR (neat) 3331, 2924, 1621, 1455, 1374, 1114 cm^{-1} ; $^1\text{H-NMR}$ (CDCl_3 , 300 MHz) δ 5.85 (d, 1H, $J = 16.3$ Hz), 5.75 (dd, 1H, $J = 17.5, 10.7$ Hz), 5.44 (m, 1H), 5.28 (m, 1H), 4.86 (dd, 1H, $J = 17.5, 1.2$ Hz), 4.79 (dd, 1H, $J = 10.7, 1.2$ Hz), 0.95-1.98 (m, 18H), 0.90 (s, 3H), 0.87 (s, 3H), 0.85 (s, 3H); HR-MS (EI): calcd for $\text{C}_{22}\text{H}_{34}\text{O}$ [M]: 314.2610; found: 314.2615.

Spectral data for the analog **12**

IR (neat) 3321, 2921, 1456, 1372, 1058; $^1\text{H-NMR}$ (CDCl_3 , 300 MHz) δ 5.75 (dd, 1H, $J = 17.5, 10.7$ Hz), 5.27 (m, 1H), 4.86 (dd, 1H, $J = 17.5, 1.2$ Hz), 4.79 (dd, 1H, $J = 10.7, 1.2$ Hz), 3.54 (t, 2H, $J = 6.5$ Hz), 0.90-1.99 (m, 20H), 1.02 (s, 3H), 0.90 (s, 3H), 0.80 (s, 3H); HR-MS (EI): calcd for $\text{C}_{22}\text{H}_{36}\text{O}$ [M]: 316.2766; found: 316.2758.

BIOLOGICAL ASSAYS

Inhibition of NO production in Raw 264.7 cells was assayed by measuring the accumulation of nitrite in the culture medium by the Griess reaction. Raw 264.7 cells were transferred in 96 well plates at a density of 1×10^5 cells/well. After 3 h incubation, the cells were stimulated with LPS (1 $\mu\text{g}/\text{mL}$) for 24 h in the absence or presence of the acanthoic acid analogs tested. As a parameter of NO synthesis, nitrite concentration was measured in the supernatant of Raw 264.7 cells by Griess reaction (Jin *et al.*, 2002).

Table I. The *in vitro* data of NO inhibition of acanthoic acid analogs

entry	Structure (R)	analog	NO inhibition IC_{50} , μM
1	$\text{CH}=\text{CHCO}_2\text{H}$	4	>10
2	$\text{CH}=\text{CHCOim}^a$	8	1.02
3	$\text{CH}=\text{CHCH}_2\text{OH}$	9	0.05
4	$\text{CH}_2\text{CH}_2\text{COim}$	11	19.4
5	$\text{CH}_2\text{CH}_2\text{CH}_2\text{OH}$	12	0.21
6	$\text{CH}_2\text{CH}=\text{CHCO}_2\text{H}$	7	>10
7	$\text{CH}_2\text{CH}=\text{CHCOim}$	13	>10
8	$\text{CH}_2\text{CH}=\text{CHCH}_2\text{OH}$	14	>10
9	$\text{CH}_2\text{CH}_2\text{CH}_2\text{COim}$	16	>10
10	$\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{OH}$	17	>10

^aim: imidazole.

RESULTS AND DISCUSSION

The *in vitro* data of NO inhibition of the synthesized compounds are summarized in Table I.

For the extension of the C_4 -linker, the two-carbon homologation turned out to be optimal for the increased NO inhibitory activity. In case of the terminal functional groups (the analogs **4**, **8**, **9**, **11**, and **12**), the analogs bearing alcohol and amide exhibited the better NO inhibitory activities. In addition, the alcohol analogs, **9** and **12**, showed the higher potencies compared to the amide analogs, **8** and **11**. In particular, the alcohol analog **9**

exhibited the most potent activity with an IC₅₀ of 50 nM. This also partly support that the presence of double bond in the C₄-chain is beneficial for the higher NO inhibitory activity.

In conclusion, we have established the partial structure-activity relationship (SAR) of the C₄-chain length of the acanthoic acid analogs although our study does not provide clear SAR yet. In addition, our study confirmed that the C₄-chain length plays an important role for the NO inhibitory activity of the acanthoic acid analogs. Moreover, we have identified an acanthoic acid analog, which possesses an excellent inhibitory activity in NO production in Raw 264.7 cells through the SAR studies. The enzyme assay in three NOS isozymes are currently being explored to validate the selectivity for iNOS and the successful results will be reported in due courses.

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REFERENCES

- Palmer, R. M. J., Ferrige, A. G., and Moncada, S., Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. *Nature*, 327, 524-526 (1987).
- Pfeiffer, S., Mayer, B., and Hemmens, B., Nitric Oxide: Chemical Puzzles Posed by a Biological Messenger. *Angew. Chem., Int. Ed.*, 38, 1714-1731 (1999).
- Rees, D. D., Palmer, R. M. J., and Moncada, S., Role of Endothelium-Derived Nitric Oxide in the Regulation of Blood Pressure. *Proc. Natl. Acad. Sci. U.S.A.*, 86, 3375-3378 (1989).
- Böhme, G. A., Bon, C., Lemaire, M., Reibaud, M., Piot, O., Stutzmann, J. M., Doble, A., and Blanchard, J. C., Altered Synaptic Plasticity and Memory Formation in Nitric Oxide Synthase Inhibitor-Treated Rats. *Proc. Natl. Acad. Sci. U.S.A.*, 90, 9191-9194 (1993).
- White, K. A. and Marletta, M. A., Nitric oxide synthase is a cytochrome P-450 type hemoprotein. *Biochemistry*, 31, 6627-6631 (1992).
- Kilbourn, R. G., Jubran, A., Gross, S. S., Griffith, O. W., Levi, R., Adams, J., and Lodato, R. F., Reversal of endotoxin-mediated shock by N^G-methyl-L-arginine, an inhibitor of nitric oxide synthesis. *Biochem. Biophys. Res. Commun.*, 172, 1132-1138 (1990).
- Jang, Daniel, Murrell, and George A. C., Nitric oxide in arthritis. *Free Radical Biology & Medicine*, 24, 1511-1519 (1998).
- Miller, M. J. S., Sadowska-Krowicka, H., Chotinaruemol, S., Kakkis, J. L., and Clark, D. A., Amelioration of chronic ileitis by nitric oxide synthase inhibition. *J. Pharmacol. Exp. Ther.*, 264, 11-16 (1993).
- Kang, H.-S., Kim, Y.-H., Lee, C.-S., Lee, J.-J., Choi, I., and Pyun, K.-H., Suppression of interleukin-1 and tumor necrosis factor- α production by acanthoic acid, (-)-pimarane-9(11),15-dien-19-oic acid, and its antifibrotic effects *in vivo*. *Cell Immunol.*, 170, 212-221 (1996).
- Suh, Y.-G., Kim, Y.-H., Park, M.-H., Choi, Y.-H., Lee, H.-K., Moon, J.-Y., Min, K.-H., Shin, D.-Y., Jung, J.-K., Park, O.-H., Jeon, R.-O., Park, H.-S., and Kang, S.-A., Pimarane cyclooxygenase 2 (COX-2) inhibitor and its structure-activity relationship. *Bioorg. Med. Chem. Lett.*, 11, 559-562 (2001).
- Suh, Y.-G., Lee, K.-O., Moon, S.-H., Seo, S.-Y., Lee, Y.-S., Kim, S.-H., Paek, S.-M., Kim, Y.-H., Lee, Y.-S., Jeong, J. M., Lee, S. J., and Kim, S. G., Synthesis and anti-inflammatory effects of novel pimarane diterpenoid analogs. *Bioorg. Med. Chem. Lett.*, 14, 3487-3490 (2004).
- Jin, J. Z., Hwang, B. H., Kim, H. S., Lee, J. H., Kim, Y. H., and Lee, J., Antiinflammatory Constituents of *Celastrus orbiculatus* Inhibit the NF- κ B Activation and NO Production. *J. Nat. Prod.*, 65, 89-91 (2002).