

## The Development of Miniaturized Extraction Tool for the Gene Biosensor

Haemyun Kwon, Hyojung Kim, and Yongseong Kim\*

Department of Chemistry, Kyungnam University, Masan 631-701, Korea. \*E-mail: kimys@kyungnam.ac.kr

Received August 17, 2007

**Key Words :** DNA, Extraction, PCR, Capillary, Photopolymerization

Analysis of DNA has represented the deep insight of diagnostic and clinical information in individuals for detecting and treating genetic diseases related to loss of heterozygosity, mutation, gene copy number, and genotype.<sup>1-3</sup> Traditional DNA assays involve extraction of DNA from whole blood, amplification by the polymerase chain reaction,<sup>4</sup> and subsequent separation using slab gel electrophoresis.<sup>5</sup> Although this process is known to be reasonably successful, it is generally time-consuming and unsuitable for high-throughput analysis.<sup>6</sup> A smaller reaction volume would be beneficial to reduce the cost of reagents and the amount of waste generated. Extraction of DNA from cellular constituents is also a key for most nucleic acid-based drug discovery and clinical diagnostic applications.<sup>7</sup> It is important that inhibitors or interfering substances from the cells and extracellular environment should be removed prior to PCR analysis.<sup>8</sup> Traditionally, the organic-based extraction and ethanol precipitation have been used to isolate and concentrate DNA prior to digestion and amplification. Although this procedure could handle about 10 to 20  $\mu\text{L}$  of blood, it costs time and efforts due to many steps including cells lysis, several ultracentrifuges and precipitations.<sup>9,10</sup> Commercial kits such as Qiagen DNA Stool Mini Kit and QIAamp DNA kit have provided relatively good extraction efficiency, however, drawbacks are increasing number of steps (more than 20 steps) for blood treatment and large sample volume (more than 250  $\mu\text{L}$ ).<sup>11,12</sup>

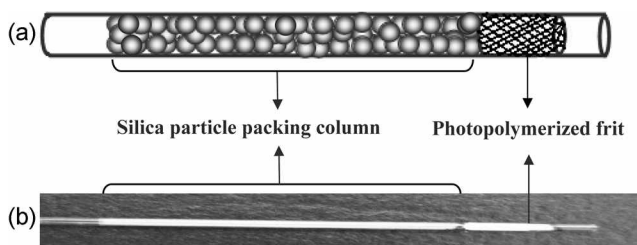
Several papers have been reported in order to overcome the disadvantages of the extraction of DNA from whole blood. For example, dendrimer-modified bacterial magnetic particles were employed based on the interaction of negatively-charged DNA and cations on dendrimer.<sup>13-15</sup> This electrostatic interaction was also utilized on amino silane coated-polydimethylsiloxane (PDMS) microchip with the blood volume of less than 20  $\mu\text{L}$ .<sup>15,16</sup> Glass microchip with heat-treated frit was used for the extraction and detection of DNA from spores of the vaccine strain. Although somewhat effective extraction was achieved with magnetic particles or microchips, many shortcomings such as long modification time, intricate control of coating, and high cost for microchip production should be considered before the experiment. Recently, DNA purification process with adsorption of DNA onto a solid surface *via* hydrogen bonding to silica and *via* electrostatic interactions has been developed.<sup>17,18</sup> This approach allows inhibiting species to be removed before the purified DNA is eluted from the surface and affords the

opportunity for miniaturization.

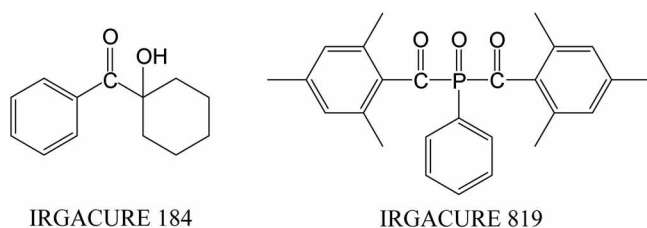
In this paper, a capillary-based DNA extraction tool was developed for the blood volume of less than 3  $\mu\text{L}$ . Only 3 steps were required before PCR amplification of extracted DNA. Glutathione-S-transferase genes related to bladder and breast cancer was tested with our extraction tool. The conditions for frit formation by photopolymerization including the ratios of photoinitiators, the frit length, UV exposure time, and the size of the silica bead particles were examined and optimized.

Figure 1 shows our miniaturized extraction tool using fused-silica capillary tube (520  $\mu\text{m}$  i.d.). As shown in Figure 1(a) and (b), the tool was built with two parts: the region for silica particles and the frit for packing of the particles. Packed silica particles provide the isolation and extraction of genomic DNA while the frit holds the silica particles. Compared to the sleeve type packing with glass fiber,<sup>17</sup> our tool is simpler in construction, and easier to fill the particles in the capillary tube.

For the construction of the frit, the choice of the photoinitiator is important. Bis acyl phosphine or  $\alpha$ -hydroxyketone is a well-known chemical class that absorbs UV energy from the light source and initiates the radical polymerization that converts the liquid formulation to a solid, cured film. In our experiment, it was found that the ratio of the IRGACURE solutions played a critical role in the formation of the frit. When either IRGACURE 184 ( $\alpha$ -hydroxyketone) or IRGACURE 819 (bis acyl phosphine) as shown in Figure 2 was tested as the curing agent, the frit formation was failed. Since the manufacturer recommended the use of IRGACURE 184 in combination with IRGACURE



**Figure 1.** Schematic of the miniaturized DNA extraction tool (a) and the real picture (b). Capillary tube; 520  $\mu\text{m}$ , i.d., total length of the capillary; 10 cm, the size of the silica bead; 15  $\mu\text{m}$ , the length of the bead-packed region; 3.0 cm, the length of the photopolymerized region; 3.0 mm, the ratio of IRGACURE 184 to IRGACURE 819; 4:1.



**Figure 2.** Chemical structure of IRGACURE 184 (1-hydroxy-cyclohexyl-phenyl ketone) and IRGACURE 819 (phenyl bis(2,4,6-trimethyl benzoyl) phosphine oxide).

819 for good surface hardness, we have tried many different ratios of both curing agents. It was observed that the ratios other than 4:1 of IRGACURE 184 and IRGACURE 819 did not provide the solid formation of the stable frit. Although the reason was not clear yet, we believe that the wavelength and the power of the UV lamp might be related to the activity of the curing agents.

During photopolymerization process, the effect of UV exposure time was investigated by obtaining the image of scanning electron micrograph. As shown in Figure 3(b), UV exposure time between 15 to 20 min produced the stable frit successfully. When the exposure time was relatively short (Figure 3(a)), the frit was not properly formed. When the exposure time was more than 20 min, the packing of the silica particles became strenuous due to high back pressure against the frit. It was observed that the length of the frit was also significant. As shown in Table 1, the stability of the frit was determined by the length of the photopolymerized frit. For the frit length of shorter than 3.0 mm, the stability of the frit was weak, resulting in the breakage of the frit structure.

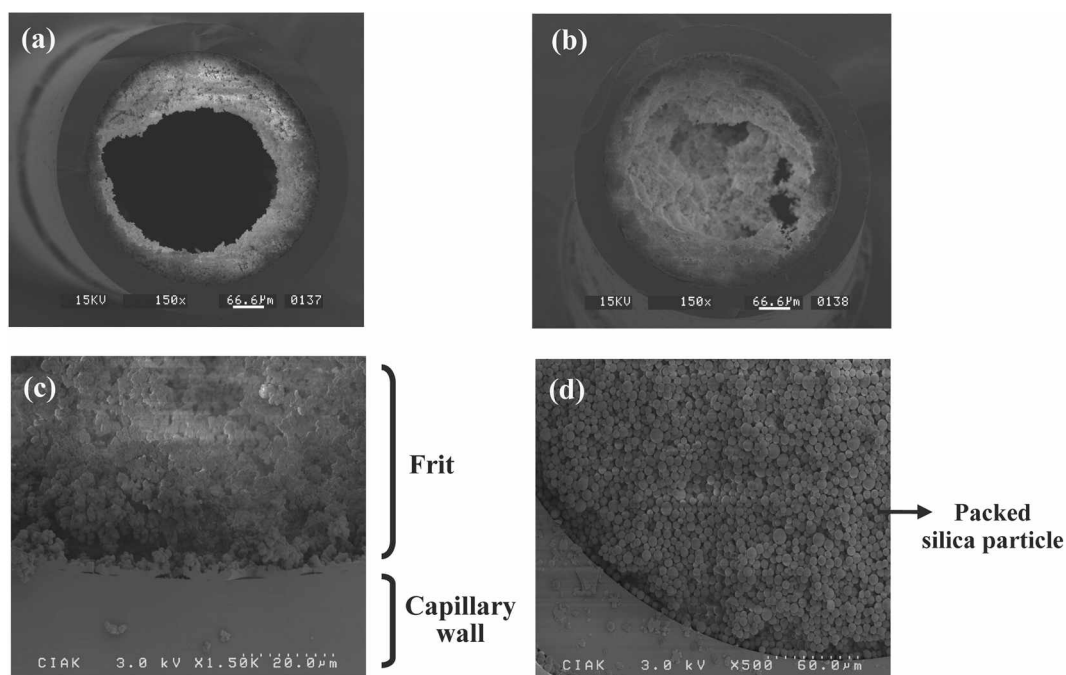
**Table 1.** Stability of photopolymerized frit length

Photopolymerized frit length (mm)	Stability of photopolymerized frit
1.0	very weak
2.0	weak
3.0	stable
4.0	low back pressure
5.0	high back pressure

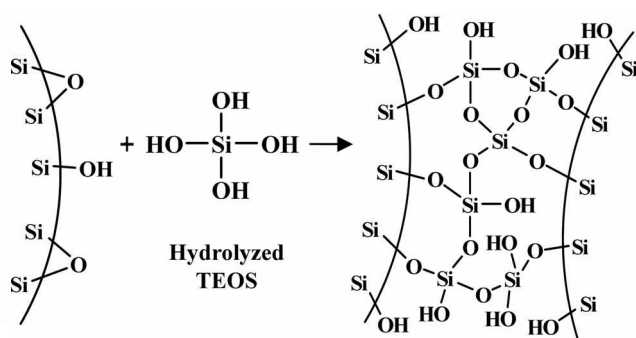
When the length was larger than 3.0 mm, the back pressure became stronger, which made the packing of the silica particles problematic.

It seemed that the size of the silica particles did not cause much variation in the yield of DNA extraction. We have employed the silica particles of 5.0 or 15  $\mu\text{m}$  diameter for the same volume of the capillary tube, and it turned out that the results of PCR amplification were similar. However, the fixation of the silica particles in the capillary tube was important. When only silica slurry was introduced and dried, the packing of the silica bead particles became less dense, resulting in irreproducible DNA extraction. As shown in Figure 4, hydrolyzed tetraethoxysilane (TEOS) not only provides more hydroxyl groups for DNA binding,<sup>5,18</sup> but also fixes silica beads in place generating more dense packing (see Figure 3(d)).

In order to confirm the applicability of our capillary-based miniaturized tool for DNA extraction to the real blood sample, two genes (GSTM1 and GSTT1) from glutathione-S-transferase (GST) were selected as the test DNA. It is known that deletion of GSTM1 or GSTT1 may cause the cancer due to malfunction of glutathione-S-transferase, a



**Figure 3.** Scanning Electron Microscopy (SEM) images of the frit and the silica bead packing. (a) UV exposure time 5 min to 10 min, 150 X magnification, (b) UV exposure time 15 min to 20 min, 150 X magnification (c) 1500X magnification of (b), (d) silica bead packing, 500 X magnification. SEM acceleration voltage; 10-30 kV.



#### Silica bead

**Figure 4.** Schematic of the fixation of silica beads by hydrolyzed tetraethoxysilane (TEOS).

xenobiotic metabolizing enzyme that deactivates carcinogens or procarcinogenic compounds or protects cells from oxidative stress.<sup>19,20</sup> Figure 5 shows the analysis of GSTM1 and GSTT1 from 3  $\mu$ L of blood.  $\beta$ -Globin was used as the internal standard. It was obvious that GSTT1 was deleted for one individual (Figure 5(a)), while both GSTM1 and GSTT1 were missed for the other individual (Figure 5(b)). For the individual in Figure 5(c), it was clearly seen that GSTM1 was missed. Figure 5 shows that a genetic polymorphism for glutathione-S-transferase could be verified with our lab-on-a-chip system for different individuals.

Our capillary-based miniaturized DNA extraction tool was successfully applied to the analysis of genes from human blood. Since the blood volume of less than 3  $\mu$ L was required, the potential for reducing pain for the patients, decreasing waste disposal, and shortening the process time for DNA extraction would be excellent. Compared to the polyethylene sleeve type extraction tool reported by Tian's group,<sup>17</sup> our miniaturized tool turned out to be simpler to construct and more efficient to perform DNA extraction due to reduced instability of microparticle bead bed using TEOS silica "glue". Also, our capillary-based system is much cheaper to prepare than a microchip-based one<sup>5,18</sup> since the photolithographic process for the microchip production is generally more expensive. Integration of this miniaturized extraction tool to genetic disease diagnosis using lab-on-a-chip is under progress in our laboratory.

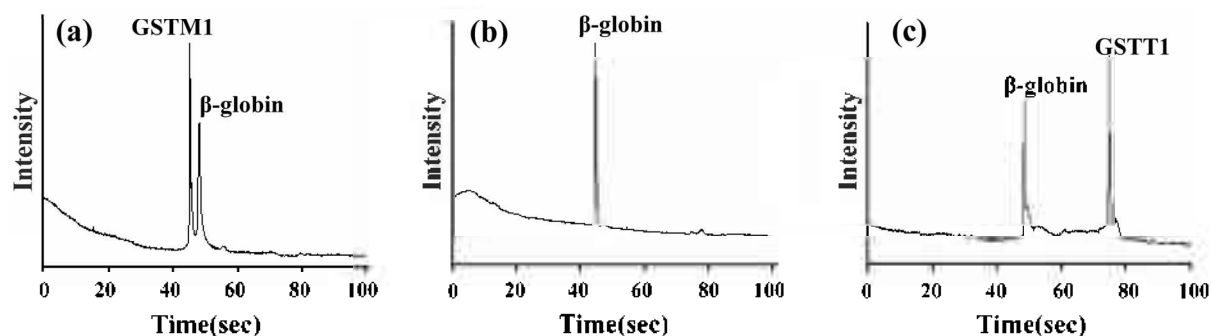
#### Experimental Section

**Chemicals.** Tetraethoxysilane (TEOS), guanidine hydrochloride (GuHCl), Triton-X 100, Tris(hydroxymethyl)aminomethane, boric acid, 3-(trimethoxysilyl)propyl methacrylate (TMSPM), ethylenediaminetetraacetate (EDTA), ethidium bromide (EB), poly(ethylene oxide) (PEO,  $M_w = 8,000,000$ ), toluene, 2-propanol, hydrochloric acid, nitric acid were obtained from Sigma-Aldrich, Co. (MO, USA). A monomeric dsDNA intercalating dye (EB) is mutagenic and carcinogenic so that a pair of lab-glove should be used during treatment. For DNA extraction, either TE (10 mM Tris, 1 mM EDTA) or 1x TBE (89 mM Tris, 89 mM borate, 2.0 mM EDTA) were employed.

Bare fused silica capillary with 520  $\mu$ m i.d. was purchased from Polymicro Technologies (Phoenix, AZ, USA). Glass capillary tubes with 1.1 mm i.d. were from Scientific Glass Inc. (Rockwood, USA). IRGACURE 184 and 819 as a photoinitiator were acquired from Ciba Specialty Chemicals Inc. (Basel, Switzerland). Silica particles with either 15  $\mu$ m or 5  $\mu$ m diameter were obtained from Daiso Co. (Osaka, Japan). PCR premix and glutathione-S-transferase (GST) were from Bioneer, Co. (Daejeon, Korea). Deionized water (Mili-Q reagent water system, MA, USA) was used throughout the experiment.

**Miniaturized extraction tool.** The sol-gel solution was made up of 750  $\mu$ L of 3-(trimethoxysilyl)propyl methacrylate, 22.5  $\mu$ L of 0.12 M hydrochloric acid, and 225  $\mu$ L of water. This solution was stirred for 30 min in the dark at room temperature. A 170  $\mu$ L of toluene was added to 30  $\mu$ L of the sol-gel solution and stirred for 30 min at room temperature. A 1.78 mg of IRGACURE 819 and a 7.12 mg of IRGACURE 184 were added to the toluene mixture. Then, this IRGACURE solution was stirred for 1 h at room temperature and stored at dark for later use.

A 3.0 mm window was made at the end of the 520  $\mu$ m i.d. polyimide-coated capillary tube using a torch. After IRGACURE solution was filled inside the capillary tube, both ends of the capillary were sealed with parafilm prior to UV exposure for 15 min by Deuterium lamp (E8039, Hamamatsu Photonics K. K., Shizuoka-Ken, Japan). When the frit at one end of the capillary tube was formed, it was



**Figure 5.** Analysis of PCR products of GSTM1,  $\beta$ -globin, and GSTT1 by using PDMS microchip. (a) GSTM1 (215 bp),  $\beta$ -globin (268 bp), (b)  $\beta$ -globin, (c)  $\beta$ -globin, GSTT1 (480 bp). Electric field strength: 250 V/cm, separation channel length: 3.5 cm, channel size: 80  $\mu$ m width  $\times$  50  $\mu$ m depth, running buffer: 1.2% PEO containing 3.0  $\mu$ g/mL of ethidium bromide in 1X TBE.

rinsed with ethanol by positive pressure with a syringe to remove the unreacted IRGACURE solution.

The capillary tube was filled with silica beads by drawing a bead/water slurry toward the frit by using the syringe. Then, the column was rinsed with 1 M HCl to hydrolyze the surface of the silica and the capillary wall. Hydrolyzed TEOS solution was prepared by adding 0.1% (v/v) of HNO<sub>3</sub> to a 27% solution of TEOS in water, and heating to 60 °C for 10 min and then 80 °C for 60 min with stirring at 200 rpm.

**DNA extraction procedure.** New miniaturized extraction tool was conditioned with MeOH for 30 min. For each extraction, extraction tool was rinsed with elution buffer (TE) for 30 min, followed by GuHCl load solution. The extraction procedure itself consisted of load, wash and elution steps. Before the load step, whole blood (3.0 μL) was lysed by mixing with 220 μL of 10% Triton-X 100 and 20 μL of proteinase K and then water bath at 55 °C for 25 min. A 50 μL of the lysate was passed through the extraction tool. Protein and possible PCR inhibitors that were adsorbed onto the silica during the load step were removed by passing 70 μL of the wash buffer (2-propanol/H<sub>2</sub>O, 80%) through the extraction tool. Finally, DNA was eluted in a low ionic strength TE buffer. After elution of DNA from whole blood, the extraction tool was rinsed with load solution for 5 min to prepare the bead surface for a subsequent extraction.

**Polymerase chain reaction (PCR) for GSTM1 and GSTT1.** The primers for GSTT1, GSTM1, and β-globin were as follows: GSTT1-forward: TCACCGGATCATGGCCAGCA, GSTT1-reverse: TTCCTTACTGGTCCATC-ATC-TC, GSTM1-forward: GAAGTCCCTGAAAAGCTA-AAGC, GSTM1-reverse: GTTG-GGCTCAAATATACGGT-GG β-globin-forward: CAACTTCATCCACGTTCCACC, β-globin-reverse: GAAGAGCCAAG GACAGGTAC. PCR was performed with thermal cycler system (T-CY) using the following protocol; preheating (95 °C for 15 min), 30 cycles at 94 °C for 1 min, 61 °C for 1 min, and 72 °C for 1 min, with a final extension at 72 °C for 1 min. The PCR products were confirmed by the slab gel electrophoresis using 1.0% agarose gel and by lab-on-a-chip system using 1.2% poly(ethylene oxide) (PEO) gel.<sup>21</sup>

**PDMS microchip electrophoresis.** Polydimethyl siloxane (PDMS) oligomer and curing agent (Oprontech Inc., Changwon, Korea) were mixed at the ratio of 10:1 and the resulting bubbles were eliminated by vacuum in desiccator. Then, PDMS was poured on the master and dried at 75 °C for 2 hrs. For the formation of the microchannel, another PDMS layer was oxidized by for 10 min by Tesla coil (BD-10A, Electrotechnique Production, Inc., Chicago, IL, USA) and bonded to the PDMS layer with engraved channel pattern on it.

DBMA-100 (Digital Bio Technology, Korea) equipped with DPSS laser (532 nm, 10 mW) and DBHV-100 high voltage supplier was used. A double-T shape PDMS microchip with long channel length of 3.5 cm, channel width of 80 μm, and channel depth of 50 μm was positioned in place inside DBMA-100 for DNA detection.<sup>22-24</sup> A 6 μL of PCR product was injected by the pinched injection mode and

detected at the long channel with 250 V/cm. PDMS microchip was cleaned with DI water, 0.01 M HCl, 0.10 M NaOH, 1XTBE for 5 min, respectively, prior to the loading of 1.2% PEO containing of 3 μg/mL of EB to the long channel.

The fluorescence signal from PCR product was collected with a 10X microscope objective (Nikon, Japan) into the photomultiplier module (H5784-02, Hamamatsu, Shizuoka-Ken, Japan) and transferred directly through a low-pass filter to an A/D interface board (National Instrument Co., TX, USA). The control of the high-voltage power supply and data collection at 10 Hz was performed by in-house Lab-View program with an IBM compatible computer.

**Acknowledgment.** This work was supported by Kyungnam University Foundation Grant, 2006.

## References

- Kim, D. E.; Jeong, Y. J. *Bull. Korean Chem. Soc.* **2006**, *27*, 1618.
- Kang, S. H.; Jang, S.; Yi, H. K. *Bull. Korean Chem. Soc.* **2005**, *49*, 537.
- Kim, J. M.; Lee, M. Y.; Kim, Y. Z. *Bull. Korean Chem. Soc.* **2005**, *26*, 285.
- Jang, S.; Cho, K.; Chae, J. S.; Kang, S. H. *Bull. Korean Chem. Soc.* **2004**, *25*, 757.
- Wolfé, K. A.; Breadmore, M. C.; Ferrance, J. P.; Power, M. E.; Conroy, J. F.; Norris, P. M.; Landers, J. P. *Electrophoresis* **2002**, *23*, 727.
- Lin, Y. W.; Huang, M. F.; Chang, H. T. *Electrophoresis* **2005**, *26*, 320.
- Wu, Q.; Bienvenue, J. M.; Hassan, B. J.; Kwok, Y. C.; Giordano, B. C.; Norris, P. M.; Landers, J. P.; Ferrance, J. P. *Anal. Chem.* **2006**, *78*, 5704.
- Roussela, Y.; Wilksa, M.; Harrisb, A.; Meinc, C.; Tabaqchali, S. *Journal of Microbiological Methods* **2005**, *62*, 71.
- Shimelis, O.; Zhou, X.; Li, G.; Giese, R. W. *J. Chromatogr. A* **2004**, *1053*, 143.
- Trochimeluka, T.; Fotheringham, J.; Toppb, E.; Schrafta, H.; Leunga, K. T. *Journal of Microbiological Methods* **2003**, *54*, 165.
- Cler, L.; Bu, D.; Lewis, C.; Euhus, D. *Molecular and Cellular Probes* **2006**, *20*, 191.
- Castella, V.; Simonin, N. D.; Casadevall, C. B.; Mangin, P. *Forensic Science International* **2006**, *156*, 70.
- Yoza, B.; Arakaki, A.; Matsunaga, T. *Journal of Biotechnology* **2003**, *101*, 219.
- Yoza, B.; Arakaki, A.; Maruyama, K.; Takeyama, H.; Matsunaga, T. *Journal of Bioscience and Bioengineering* **2003**, *95*, 21.
- Yoza, B.; Matsumoto, M.; Matsunaga, T. *J. Biotechnol.* **2002**, *94*, 217.
- Nakagawaa, T.; Tanakaa, T.; Niwab, D.; Osakab, T.; Takeyamaa, H.; Matsunagaa, T. *Journal of Biotechnology* **2005**, *116*, 105.
- Tian, H.; Hühmer, A. F. R.; Landers, J. P. *Analytical Biochemistry* **2000**, *283*, 175.
- Breadmore, M. C.; Wolfé, K. A.; Arcibal, I. G.; Leung, W. K.; Dickson, D.; Giordano, B. C.; Power, M. E.; Ferrance, J. P.; Feldman, S. H.; Norris, P. M.; Landers, J. P. *Anal. Chem.* **2003**, *75*, 1880.
- Cotton, S. C.; Sharp, L.; Little, J.; Brockton, N. *Am. J. Epidemiol.* **2000**, *151*, 7.
- Cha, I. H.; Kwon, J. J.; Park, K. K. *J. Lor. Oral Maxillofac. Sug.* **2002**, *28*, 364.
- Fung, E. N.; Yeung, E. S. *Anal. Chem.* **1995**, *67*, 1913.
- Kang, S. H.; Jang, S.; Park, S. K. *Bull. Korean Chem. Soc.* **2006**, *27*(9), 1346.
- Kang, C. M.; Back, S. K.; Choi, B. O.; Lee, S.; Chung, K.; Kang, S. H.; Kim, Y. S. *Bull. Korean Chem. Soc.* **2006**, *27*, 1239.
- Kang, C. M.; Back, S. K.; Song, I. G.; Choi, B. O.; Chang, J.; Cho, K.; Kim, Y. S. *Bull. Korean Chem. Soc.* **2006**, *27*(4), 519.