# 紫蘇葉 추출물의 抗凝血 활성에 관한 研究

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# A Study of Anticoagulation Activity from Perillae Folium Extract

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# ABSTRACT

**Objectives :** This research was investigated to find out the effect of the anticoagulant Perillae folium extract.

**Methods :** To examine an active effect of anticoagulation in Perillae folium extract, the study measured Prothrombin time(PT) and activated partial thromboplastin time(APTT) of human plasma in vitro and measured bleeding time and arterio-venous shunt model in rats *in vivo*.

**Results :** Bleeding time of Perillae folium extract *in vivo* had a significant increase 1.6 times and thrombus weight of Perillae folium extract had a significant reduction of thrombus weight as 68%. Perillae folium extract had an effect of anticoagulation by operating on extrinsic pathway factor II, V, VII, X and intrinsic pathway factor VIII, IX, X, VI, VII in the coagulation system.

**Conclusions :** Considering the above mentioned results, it is judged that a Perillae folium extract has a control effect of thrombus creation.

Key words: Perillae folium, Anticoagulant, Thrombosis, Prothrombin time, Bleeding time

# Introduction

Perillae folium is an annual herbaceous belonging to Lamiaceae family with its origin from China that has Perilla frutescens Britton var. acuta kudo leaves with unique fragrance and also grows wild in various regions of our country.

Leaves sprout face to face with long leafstalk, often purplish color and serrations in the edges,

and ones with strong fragrance and without branches and leafstalk are considered better.

The leaves of Perilla frutescens, a traditional Asian medicinal herb, have been used in the Orient for centuries to treat various conditions including depression<sup>1)</sup>, infection<sup>2)</sup>, inflammation<sup>3,4)</sup> and allergies<sup>5,6)</sup>. Several studies have also suggested that extract of Perillae folium take in antitumor growth in a murine skin model in vivo<sup>7,8)</sup>. In vitro

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<sup>·</sup> Acceptance : 2008. 11. 30. · Adjustment : 2008. 12. 17. · Adoption : 2008. 12. 22.

studies have reported that Perillae folium components, rosmarinic acid and luteolin, can induce apoptosisin a variety of cancer cell lines<sup>9-12</sup>. However, the mechanism of the antitumorigenesis effects of whole Perillae folium extract remains unclear.

Formation of blood clots in blood vessels can be divided largely into 3 steps. Step 1 is the process in which thromboplastin is separated from the damaged tissue or cut thrombocyte when bleeding, step 2 is the process in which prothrombin is converted to thrombin under presence of  $Ca^{2+}$ , and step 3 is the process in which various coagulation factors are continuously activated to form blood clot by changing fibrinogen in blood plasma into insoluble fibrin using thrombin. Clot is formed by extrinsic pathway that begins from damage in blood vessel walls or surrounding tissues and intrinsic pathway that begins from coagulation factors within blood vessels<sup>13–15)</sup>.

The purpose of this study is to conduct an experiment on anti-coagulation effect of Perillae folium in the screening process for development of anticoagulation medical supplies and to report its results.

## Materials and methods

#### 1. Preparation of Perillae folium

Perillae folium (*Perilla frutescens* var. acuta Kudo) had been purchased from Hebei region of China, examined by the Department of Oriental Medicine Resources at Joongbu University for careful selection and used as samples while being refrigerated at 4°C.

460 mL of extract obtained by 3 hours of heating of 30 g Perillae folium(PLF) in 1,000 mL distilled water was filtered once using a filter paper (Whatman No. 1) and decompressed and concentrated using a rotary vacuum evaporator. It was then settled in a  $-80^{\circ}$ C deep freezer for 4 hours and lyophilized by freeze dryer for 12 hours to obtain 6.2 g of powder. The powder was refrigerated (4°C) until used for this experiment and had been diluted to 10 mg/mL (PLF 1), 30 mg/mL (PLF 2) or 50 mg/mL (PLF 3) according to necessity. Preparation of Human plasma

Fresh frozen plasma for research use was donated by Daejeon, Chungnam Chapter of National Red Cross for human plasma, and separated human plasma was refrigerated with volumes of 10 mL in 15 mL conical tubes to be defrosted to 37°C before use.

#### 2. Animals

Male Sprague-Dawley rats were purchased from Core Tech Co., Ltd., 12-week-old male Sprague Dawley (SD) rats with body weight of 350±10 g had been received from Core Tech Co., Ltd. and were used for this experiment after adaptation period of at least one week. Until the day of experiment, they were provided with enough compressed feed (crude protein 22.1% or above, crude fat 8.0% or below, crude fiber 5.0% or below, crude ash 8.0% or below, calcium 0.6% or above, phosphorus 0.4% or above, Samyang, Korea) and water. Breeding condition was fixed at 22±2℃ room temperature, 50±10% relative humidity, 12 hours of lighting (07:00~19:00) and 150~300Lux illuminance.

### 3. Prothrombin time (PT) measurement

For measurement of prothrombin time, Quick's one stage method<sup>16)</sup> was used. 45 uL of blood plasma and 3 mL of human plasma were put inside a dish and on top of ice. Then, 5 uL of Perillae folium extract (10 mg/mL) was put in the well of strip well plate and 5 uL of distilled water was put in the other well as control group.

After heating for 5 minutes at 37°C, 100 uL of PT reagent was added and time for coagulation was measured under extinction of 405nm.

# 4. Activated Partial Thromboplastin Time (APTT) measurement

3 mL of human plasma was put inside a dish and on top of ice. 5 uL of Perillae folium extract (10 mg/mL) was put in the well of strip well plate, and 5 uL of distilled water was put in the other well as control group. For each group, 45 uL of plasma was added and heated for 2 minutes in a microplate reader.

After adding 50 uL of APTT reagent and heating for 5 minutes, 100 uL of 25mM calcium chloride (CaCl<sub>2</sub>) was added to measure the time required for coagulation under extinction of 405nm.

#### 5. Bleeding time (BT) measurement

400 mg/kg Perillae folium extract was oral administered to SD rats once each day for 5 days and medicine was given on the 6th day. After using the method<sup>17)</sup> of Han *et al* that used 1 mL abdominal injection of 1.25 g/kg ether after 60 minutes for anesthesia, 0.3 cm was cut on the end of the tail. 5 cm of remaining tail was immediately put inside  $37.5^{\circ}$ C saline solution and time for hemostasis was measured.

#### 6. Thrombus weight measurement

400 mg/kg Perillae folium extract was oral administered to SD rats once each day for 5 days and medicine was given on the 6th day. After 1 mL abdominal injection of 1.25 g/kg ether was made after 60 minutes for anesthesia, canulation was done on left carotid artery and right jugular vein using polyethylene catheters. Then, 5cm silicon tube filled with a cotton thread was filled up using saline and blood was made to flow through the tube connected to artery and vein to induce shear-stress and thrombus for 15 minutes. Created thrombus was dried and weighed for evaluation of antiplatelet efficacy.

#### 7. Statistical analysis

Experimental results had been expressed as mean value±standard deviation for 3 repeated trials. For testing of significance of each group, T-test of Student was used with the statistics program SPSS (ver. 12.0, Korea). After comparing between the values of each experimental group, values with 95% confidence or above (p<0.05) were determined to have significance.

## Results and discussion

#### 1. Effects on PT

As a result of studying the effects on PT by adding Perillae folium extract to human plasma, control group showed 12.15±1.73 second and 50 mg/mL group showed 20.65±2.42 second. In comparison to control group, 50 mg/mL group showed twice as much increase in speed of blood flow in PT, showing that PT became longer as concentration of Perillae folium extract increased (Table 1).

Table 1. Effect of Perillae Folium Extract on Prothrombin Time

Group	Concentration(mg/mL)	PT(second)
Control	0	12.15±1.73
PLF1	10	19.53±1.62***
PLF2	30	20.90±2.82***
PLF3	50	20.65±2.42***

Control : Untreated, PLF : aqueous extract of Perillae folium.

\* : Statistically significant value compared to control by Student's t-test. (\*\*\*, p  $\leq$  0.001).

Lengthening of PT means that thrombin creation is delayed by suppressing activation of extrinsic pathway factors that participate in conversion of prothrombin to thrombin. From such result, Perillae folium extract was found to have an impact on extrinsic pathway in coagulation system.

#### 2. Effects on APTT

As a result of studying the effects on APTT by adding Perillae folium extract to human plasma, control group showed 39.98±1.36 sec, 30 mg/mL group showed 60.29±2.62 sec and 50 mg/mL group showed 81.93±4.85 second. In comparison to control group, experimental groups showed twice as much increase in APTT, showing that APTT is dependent on concentration of Perillae folium extract because APTT increased with increasing concentration (Table 2).

Increasing APTT means that creation of thrombus due to creation of fibrin is suppressed by activation intrinsic pathway factors. Perillae folium extract was found to have an impact on intrinsic pathway in coagulation system.

Table 2. Effect of Perillae Folium Extract on Activated Partial Thromboplastin Time

Group	Concentration(mg/ml)	APTT(second)
Control	0	39.98±1.36
PLF1	10	47.41±5.13
PLF2	30	60.29±2.62**
PLF3	50	81.93±4.85***

#### 3. Effects on bleeding time

As a result of comparison between effects on bleeding time in rats with Perillae folium extract and rats with Ginkgo biloba extract (GBE), bleeding time was 498.3±75.3 sec for control group and 505.7±62.9 sec for GBE group, showing no significant difference.

Perillae folium extract group showed bleeding time of 811.7±9.60 sec, meaning that bleeding time had a significant increase as about 1.6 times in comparison to control group (Fig. 1).

Measurement of bleeding time in vivo allows estimation of coagulation response in rats and can

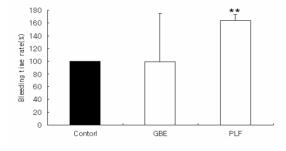


Fig. 1. Effect of Perillae folium extract on the bleeding time

Rats was treated with oral administration of 400  $\ensuremath{\operatorname{mg/kg}}$  of GBE and PLF, representatively.

be used as the basis for determining the response according to absorption of Perillae folium extract. The results suggest that Perillae folium extract affected coagulation system through absorption in the body.

#### 4. Effects on thrombus weight

Effects on thrombus weight in rats with Perillae folium extract and rats with GBE were compared.

Thrombus weight was  $286.67\pm32.15$  mg for control group,  $253.30\pm15.28$  mg for GBE group and  $195.08\pm23.45$  mg for Perillae folium extract group, showing a significant different of about 68% reduction (Fig. 2).

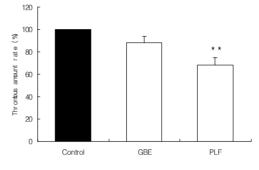


Fig. 2. Inhibition effect of Perillae folium extract on thrombus weight

Rats was treated with oral administration of 400 mg/kg of GBE and PLF, representatively.

Control : Untreated, GBE : Ginkgo biloba extract,  $\mathrm{PLF}$  : Perillae folium extract.

### Conclusions

In order to investigate anti-coagulation activity of Perillae folium, PT and APTT were measured in vitro in human plasma and effects of medication of Perillae folium extract in rats on bleeding time and thrombus weight were found in vivo to obtain the following conclusion.

1. As a result of measuring PT and APTT in human plasma, PT showed a significant increase in speed of blood flow in 50 mg/mL Perillae folium

Control : Untreated, GBE : Ginkgo biloba extract,  $\mathrm{PLF}$  : Perillae folium extract.

extract and APTT was significant by more than a factor of 2 in 50 mg/mL.

2. BT from Perillae folium extract showed a significant increase of about 61% in vivo, and formation of thrombus weight significantly decreased by about 68%.

3. Perillae folium extract was found to have influence on extrinsic pathway factors II, V, VII, X and intrinsic pathway factors VIII, IX, X, XI, XII in coagulation system.

Meaning of such results is that Perillae folium extract impedes coagulation of blood and shows anti-coagulation effects. A continued study on detailed control passway of blood coagulation factor will be conducted.

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