Modulation of Bacteria with a Combination of Natural Products in KIMCHI Fermentation

Jong Deog Kim[†], Jin Hyuk Shin, Hyo Jin Seo, Dong Jung Lim¹, Soon Kang Hong², and Tai Sun Shin³

Department of Biotechnology, Chonnam National University, Research center on Anti-Obesity and Health Care, San96-1,

Dunduk-Dong, Yosu, Chonnam, 550-749, Korea

¹Carbohydrate Bioproduct Research Center, Sejong University, 98 Gunja-Dong, Gwangjin-Gu, Seoul 143-747, Korea ²Department of Fire Service Admin, Chodang University, Muanro 1644, Muaneup, Muangun, Chonnam, 534-701, Korea ³Department of Nutrition and Food Science, San96-1, Dunduk-Dong, Yosu, Chonnam, 550-749, Korea

(Received: 2008. 3. 28., Accepted: 2008. 4. 24.)

Three kinds of bacteria that influence Kimchi fermentation, *Lactobacillus plantarium* for acidity, *Leuconostoc mesenteroides* for ripening Kimchi, and *Pichia membranifaciens* for decreasing Kimchi quality, were regulated by natural products including *Theae folium, Taraxacum coreanum, Brassica juncea, Astragali radix, Gynostemma pentaphyllum, Camellia japonica, Agaricus blazei, and <i>Cordyceps militaris*. The common prescription combined *T. folium, T. coreanum* and *C. militaris* and simultaneously regulated these 3 bacteria as follows: the growth of *L. plantarium* and *P. membranifaciens* were inhibited and *L. mesenteroides* was promoted. The most effective mixing ratio was *T. folium: T. coreanum: C. militaris* = 3:2:1. With this new prescription, deep flavor, extended preservation, and a special taste are expected in the Kimchi due to these natural products.

Key Words: Kimchi fermentative bacteria, natural products combination, common prescription, T. folium, T. coreanum, C. militaris

INTRODUCTION

The taste of Kimchi is developed by fermentation of saccharides to lactic acid and other organic acids by lactic acid bacteria, which results in the special flavor and taste. Kimchi requires a maturation temperature and preservation period at 2-7°C for 2-3 weeksto reach pH 4.3 for more delicious flavor and for vitamin production [1].

But as time passes, the taste can worsen due to acidification, softness of texture, and bad odors. To preserve the freshness of the Kimchi, additives [2-4], preservatives [5], pH buffering agents [6, 7], bacteriocin [8] and chitosan [9, 10], which is not safe for consumption, canbe used. Currently, the cold chain method is applied to Kimchi preservation [11]. Thirty bacteria and yeast have been applied to Kimchi fermentation, such as the G(+) bacteria [8], L. brevis, L. plantarium, L. latis, L. mesenteroides, P. cerevisiae [12], S. faeculis, and E. faecalis, the G(-) bacteria, P. nigrifaciens and B. macerans, and the yeast

decrease the septic white membrane.

Tel: +82-61-659-3305, Fax: +82-61-659-3305

E-mail: pasteur@chonnam.ac.kr

MATERIALS AND METHODS

Zygosaccharomyces, which produces organic acids

[13]. The time course of G(+) and G(-) behavior is an

important index of Kimchi fermentation. Our purpose is to

preserve the Kimchi by controlling these bacteria with natural

products as sources of antibacterial, antifungal, antioxidative

[14] and anti-angiogenesis effects [15, 16] such as T. folium,

T. coreanum, G. pentaphyllum, B. juncea, A. radix, C.

japonica flower, C. japonica leaf, C. japonica pericarp, C.

militaris, and A. blazei. To this end, Lactobacillus sp. [13]

should be controlled to protect against an acidic taste,

Leuconostoc sp. [7] should be promoted for delicious and

abundant taste, and Pichia sp. [7] should be controlled to

Natural Products

Green tea was supplied by Okrojeda company. A. radix was purchased from an herbal market. G. pentaphyllum, T. coreanum, C. japonica flower, C. japonica leaf, C. japonica

Some common prescriptions were constructed to simultaneously control these 3 kinds of Kimchi fermentative bacteria using natural products to preserve the delicious, ripening taste of Kimchi.

[†] Corresponding Author: Department of Biotechnology, Chonnam National University, Research Center on Anti-Obesity and Health Care, San96-1, Dunduk-Dong, Yosu, Chonnam, 550-749, Korea

pericarp, and B. juncea were obtained from Dolsan, and C. militaris and A. blazei were supplied by a mushroom farm at Dolsan, Yosu-city.

Extraction of Natural Products

Natural products (300 g) were extracted with 1500 mL of D.W. in a reflux flask for 1hr, and concentrated to 500 mL. Each natural product was used alone, orin combination at a suitable mixing ratio, for experimental purposes.

Strains and Culture

L. plantarium KCTC 3108, L. mesenteroides KCTC 3722, and P. membranifaciens KCTC7628 were purchased from Gene Bank (Seoul, Korea), and cultured with MRS + 0.5% Lactose (pH 7.6), Tomato juice media, and Malt extract [17] at 25% and 150 rpm.

Modulation of Kimchi Fermentative Strains

Regulation of Kimchi fermentative strains was investigated by the difference in O.D. at 660 nm using a spectrophotometer (UV-2101PC, Shimadzu, Japan) every 6 hr. Strains were cultured in 250-mL flasks that contained sample (5 mL) and medium (95 mL) at 25°C.

Preparation of Kimchi Juice

Kimchi is made from Chinese cabbage and radish mixed with various resources such as hot pepper, garlic, ginger, jotgal, onions and salt as a control kimchi, and a common prescribed kimchi added in control kimchi with natural productscombination, fermented at different temperature, 4°C, 10°C and 20°C. Every 7days, 300g of each kimchi is crushed with mixer and filtered through a Whatman No. 2 paper, and centrifuged at 5,000 rpm for 10 min, the supernatant was used as a kimchi juice.

Measurement of pH, Salinity and Acidity

pH and salinity were measured with Consort C803 (Turnhout, Belgium), and acidity was followed Vanderzant's methods[18] using Kimchi juice.

Counting strains

For selecting *Leuconostoc* and *Lactobacillus* strains from fermented vegetables, *Lactobacilli*MRS (Difco Laboratories; Detroit, MI, USA) agar medium containing 0.1% bromphenol bilu(BPB)[19], for selection *Entrococcus* and *Pediococcus*, MRS agar containing 0.5% bile salt, azide dextrose agar (Difco)[20], for *Lactococcus*, KF *Streptococcus* agar(Difco) and M17 containing 0.002% bromophenol purple(BCP)[21], for yeast, potato dextrose agar(Difco) controlled as a pH 3.5 with 10% tartaric acid[21], and total number of bacteria was followed by Jang's method[21]using Gram staining.

RESULTS AND DISCUSSION

Selection of Natural Products for Growth Inhibitory Effect of *L. plantarium*

Fig. 1 shows the growth inhibitory effects of natural products on L. plantarium [7, 22-25] including T. folium, B. juncea, A. radix, T. coreanum, G. pentaphyllum, C. japonica flower, C. japonica leaf, C. japonica pericarp, C. militaris, and A. blazei. Of these natural products, T. folium, T. coreanum, C. japonica pericarp and C. militaris exhibited higher controlling effects on L. plantarium, which would decrease the sour taste of Kimchi. Another paper suggested that G. ularensis, S. chinensis, and A. sessliflorum would serve the same purpose [24], but these natural products are not suitable for food, except G. ularensis.

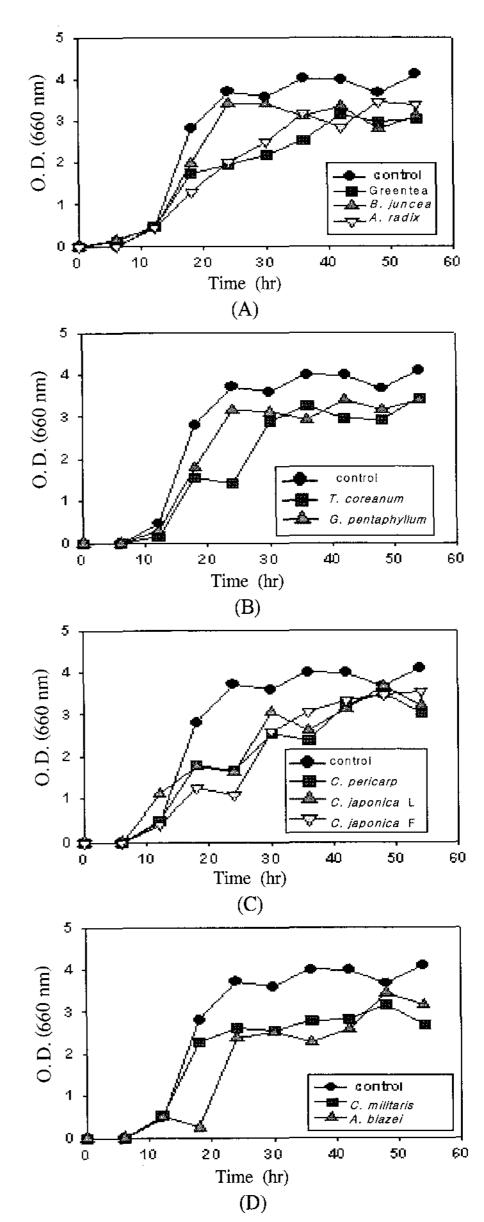


Figure 1. The growth inhibitory effects of natural products against L. plantarium. T. folium, T. coreanum, C. japonica pericarp and C. militaris exhibited higher controlling effects on L. plantarium.

Selection of Natural Products for Growth-Promoting Effects on *L. mesenteroides*

The growth of *L. mesenteroides*, which give flavor to Kimchi, was promoted by natural products such as *T. folium*, *T. coreanum*, and *C. militaris*(Fig. 2). Lee [24] showed a controlling effect for *L. mesenteroides* and not a promoting effect with *A. tuberosum* extract this bacterium provides a deep flavor by producing mannitol [26-27]. Park [17] also indicated that *S. baicalensis* and *P. amurense* extracts could control *L. mesenteroides*.

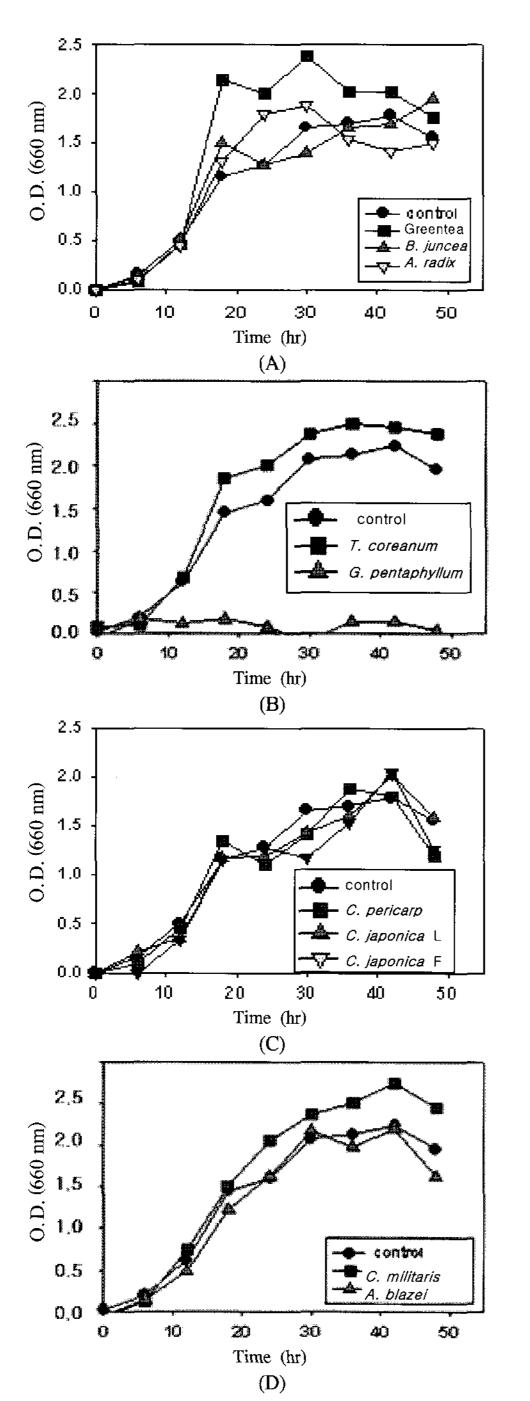


Figure 2. The growth-promoting effects of natural products against L. mesenteroides. The growth of L. mesenteroides was promoted by T. folium, T. coreanum, and C. militaris.

Selection of Natural Products for Growth Inhibitory Effect of P. membranifaciens

Fig. 3 exhibites the growth of *P. membranifaciens* was inhibited by natural products such as *T. folium, T. coreanum, G. pentaphyllum, C. japonica* flower, *C. japonica* leaf, *C. japonica* pericarp, and *C. militaris*, which would decrease the septic white membrane during Kimchi fermentation. The septic effect of this white membrane is not good for appearance, produces a bad odor and decreases Kimchi flavor and preservation duration.

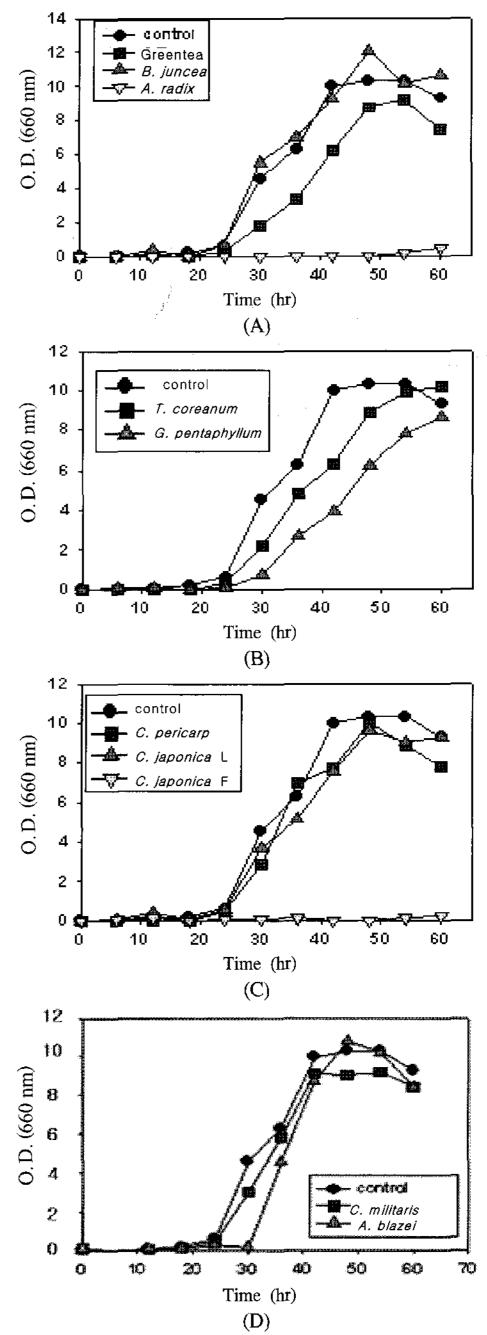


Figure 3. The growth-inhibitory effects of natural products against *P. membranifaciens*. The growth of *P. membranifaciens* was inhibited by *T. folium*, *T. coreanum*, *G. pentaphyllum*, *C. japonica* flower, *C. japonica* leaf, *C. japonica* pericarp, and *C. militaris*.

Construction of A Common Prescription to Modulate Kimchi Fermentative Bacteria.

The following combinations of natural products were prepared: a.) control; b.) T. folium, A. radix and C. militaris c.) T. Folium, T. coreanum and C. militaris d.) T. folium, T. coreanum and A. radix e.) T. folium, T. coreanum, A. radix and C. militaris f.) T. folium and C. militaris g.) T. folium and T. coreanum. Combination C is the best prescription for the combined inhibitory/promoting effects on the Kimchi fermentative bacteria, L. plantarium, L. mesenteroides, and P. membranifaciens(Fig. 4).

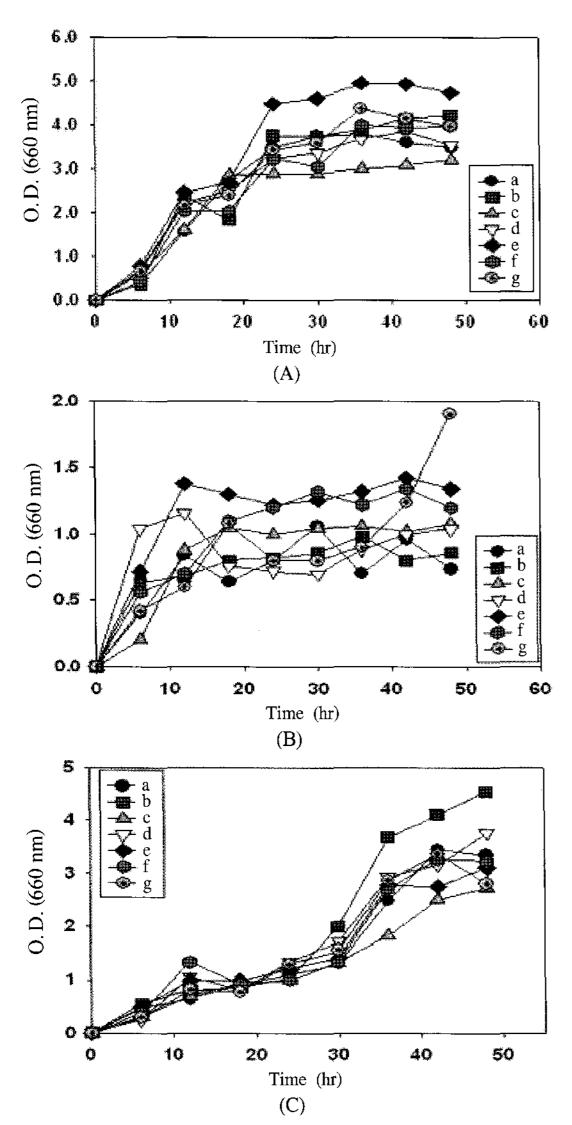


Figure 4. Common prescription to control 3 kinds of bacteria that influence Kimchi fermentation: (a) growth inhibition of L. plantarium (b) growth promotion of L. mesenteroides (c) growth inhibition of P. membranifaciens.

Optimum Combination Ratio for The Common Prescription

When the growth rates of the three bacteria were compared under conditions with natural products mixed of *T. folium*, *T. coreanum*, and *C. militaris* with mixing ratio as 1:1:1, 2:1:1,

1:2:1, 1:1:2, 3:2:1, 2:3:1, 1:3:2, and 1:1:2. The ratios of 1:1:1 and 3:2:1 exhibited higher growth promoting effects for *L. mesenteroides* (Fig. 5). For growth inhibition of *L. plantarium*, the ratios 1:2:1 and 3:2:1 showed growth inhibitory effects (Fig. 6), and for *P. membranifaciens*, the ratio of 3:2:1 showed growth inhibitory effect (Fig. 7). From these results, the best ratio is 3:2:1, which simultaneously modulated all 3 strains.

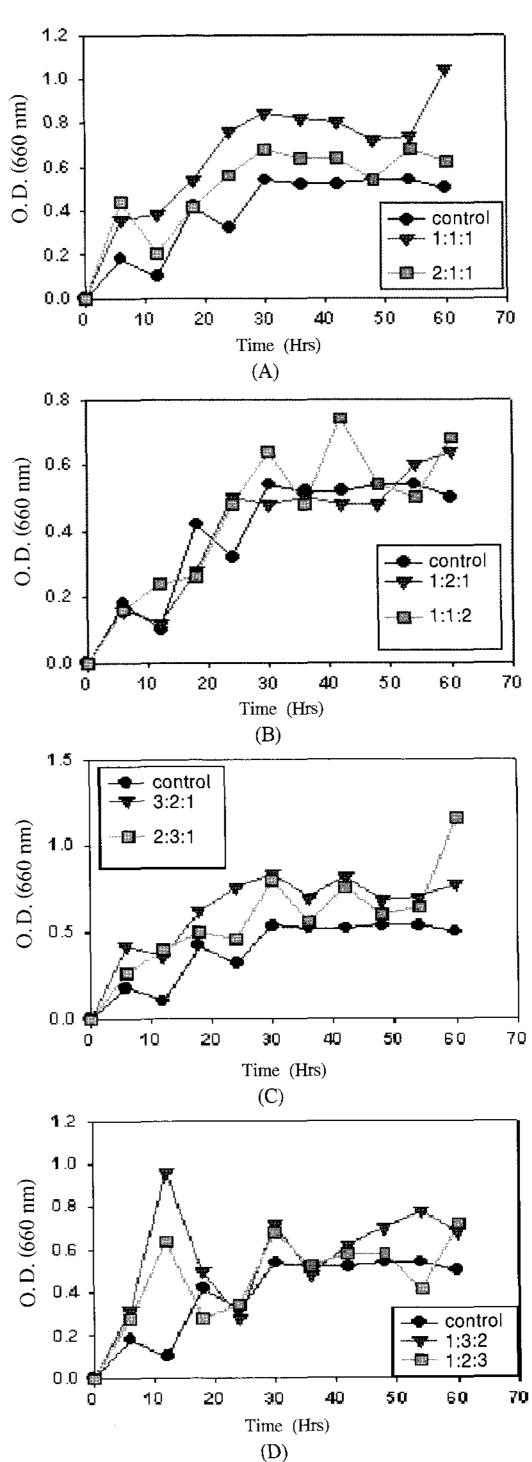


Figure 5. The most effective mixing ratio of natural products for promoting *L. mesenteroides* growth. *T. folium: T. coreanum: C. militaris*, the ratios of 1:1:1 and 3:2:1 exhibited higher growth promoting effects for *L. mesenteroides*.

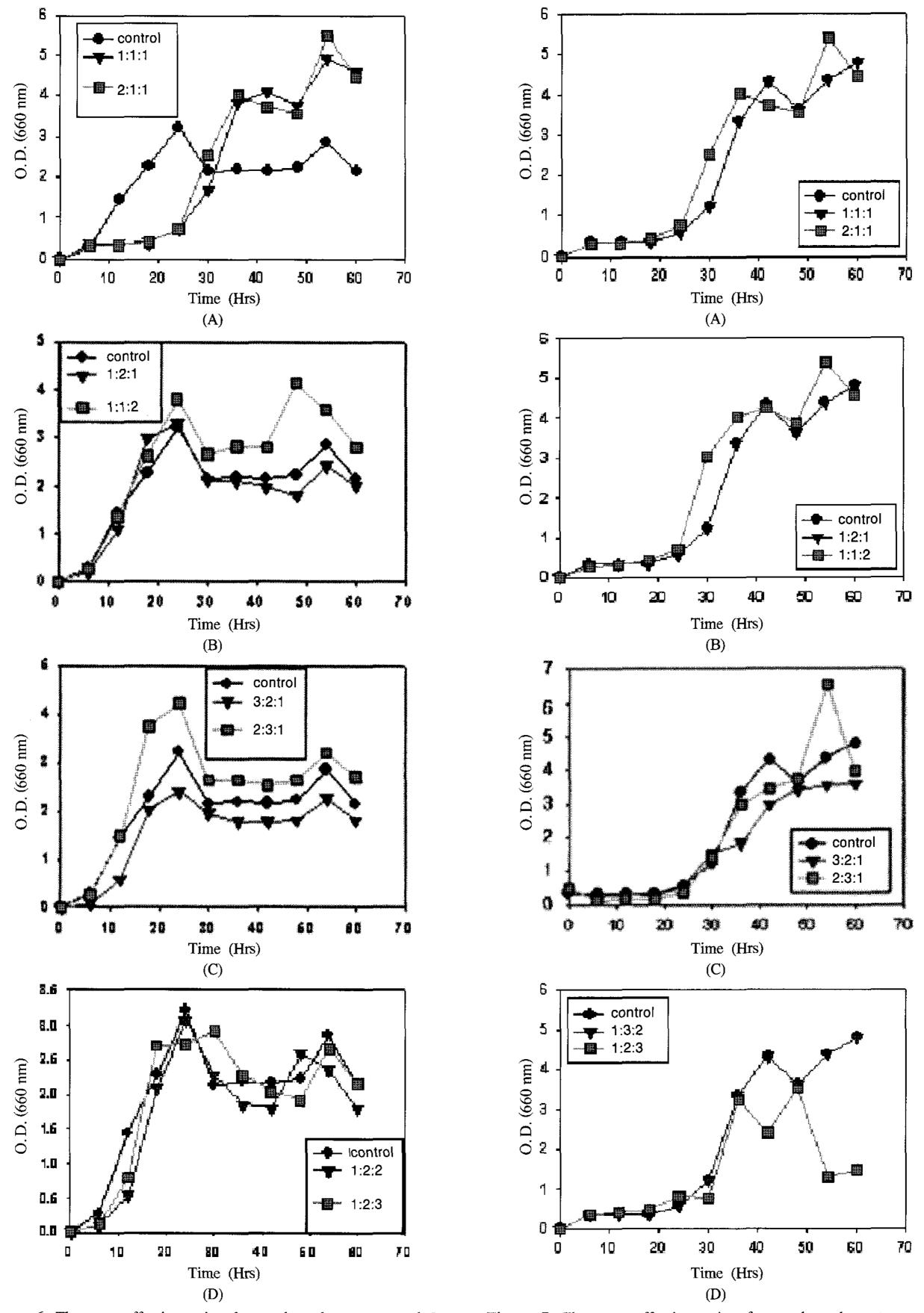


Figure 6. The most effective ratio of natural products to control L. plantarium. For growth inhibition of L. plantarium, the ratios 1:2:1 and 3:2:1 showed growth inhibitory effects.

Figure 7. The most effective ratio of natural products to control *Pichia* sp. For *P. membranifaciens*, the ratio of 3:2:1 showed growth inhibitory effect.

Characteristics of Kimchi Prepared with Common Prescription

Variation of pH, Acidity and Salinity

Kimchi fermentation factors[28-29] prepared with common prescription. were changed a various direction. For that, pH [30-31], acidity and salinity were investigated at various preserving temperatures, 4° C, 10° C and 20° C. For pH, at each temperature, Fig. 8(a) shows pH of common prescribed kimchi is higher compared with each control, which was expected to prevent souring of the Kimchi taste [32].

As with pH, for acidity, Fig. 8(b) expresses acidity of common prescribed kimchi is lower than that of controls, indicating that acids were not fully produced with the common prescription and that it can extend the duration of preservation.

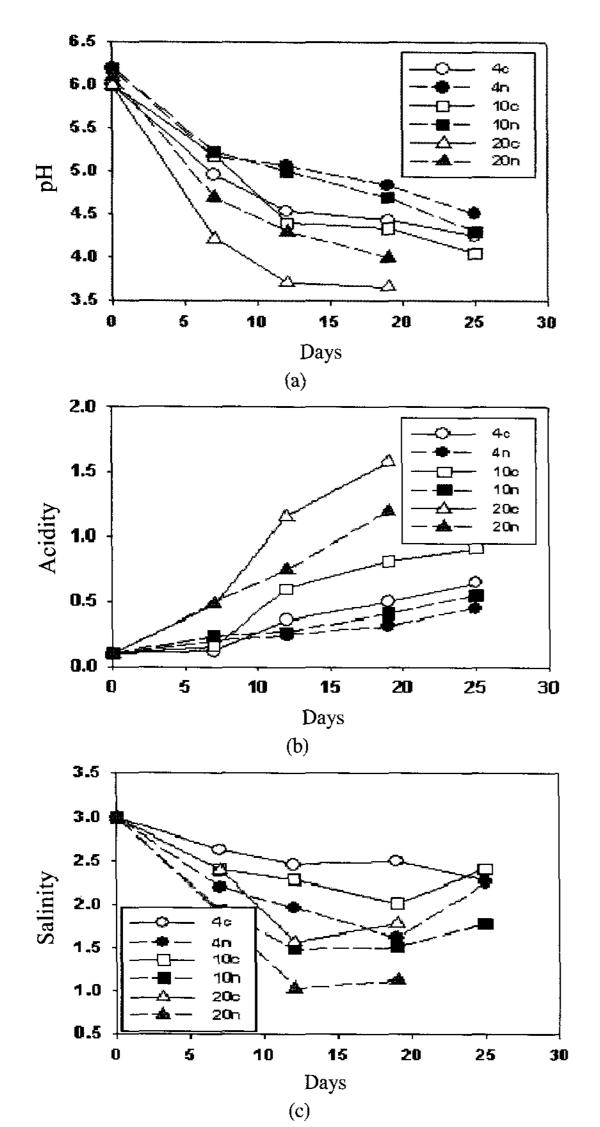


Figure 8. pH (a), acidity (b), and salinity (c) changes in Kimchi prepared with natural products according to the preservation temperatures and the number of days. 4c, 10c, and 20c: control, 4n, 10n, and 20n: natural product of each preserving temperature. pH, acidity and salinity were modulated excellently resulted in preventing sour taste, extending the duration of preservation, and preparing lower salt Kimchi.

Fig. 8(c) shows that salinity of this new type of Kimchi is tended to be lower with the natural products than that of control according to preserving temperature therefore, this common prescription would be helpful in preparing lower salt Kimchi[33].

Changes of Kimchi Fermentative Bacteria.

L. plantarium did not appear in Kimchi preserved at 4° C [Fig. 9(a)], and these bacteria were found in lower numbers $(3.8 \times 10^5 \text{ and } 4.8 \times 10^5)$ than in control $(7 \times 10^5 \text{ and } 10 \times 10^5)$ after 7 days in Kimchi preserved at 10° C, and 20° C, respectively. Therefore, this common prescription successfully controlled L. plantarium compared to control, which is important for reducing the sour taste and increasing long-term preservation.

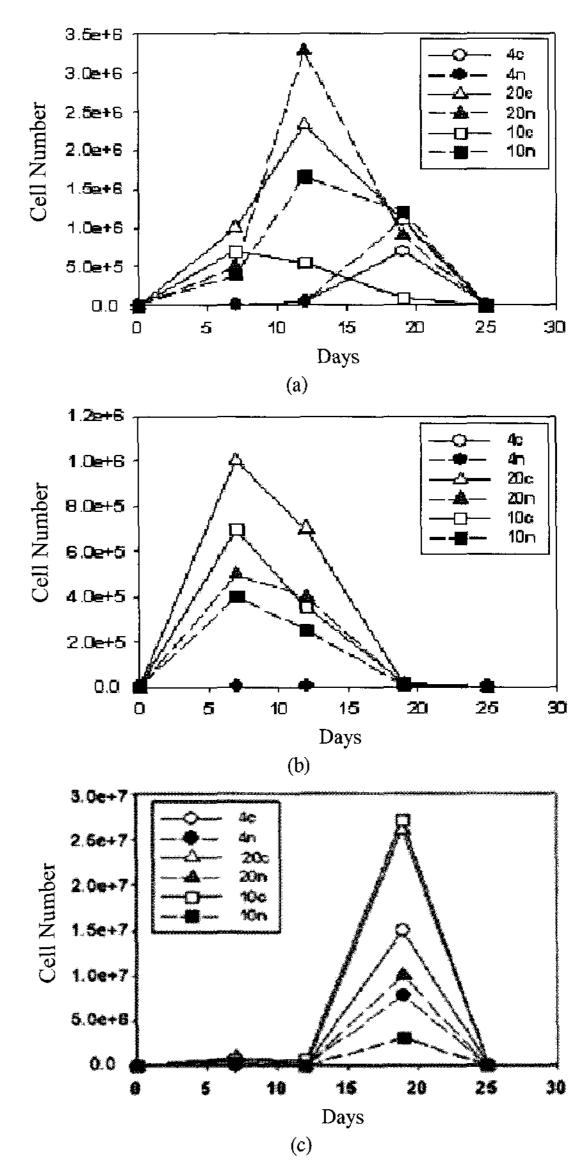


Figure 9. Controlling the number of Lactobacillus sp. (a), Leuconostoc sp. (b), Pichia sp. (c) in Kimchi according to preservation temperature and days. 4c, 10c, and 20c: control, 4n, 10n, and 20n: natural productof each preserving temperature. Controlled L. plantarium and P. membranifaciens reducd the sour taste and increased long-term preservation, and promoted L. mesenteroides improved the flavor of the new type of Kimchi.

L. mesenteroides was found in higher numbers (1.5×10^6) and 3.3×10^6) than in control (0.5×10^6) and 2.3×10^6) in Kimchi preserved at 10° C and 20° C after 12 days [Fig. 9(b)], respectively. Also, in Kimchi preserved at 20° C, L. mesenteroides was 6.6 times (3.3×10^6) higher than control at 4° C (5×10^5) and 10° C (5×10^5) .

Therefore, this common prescription promoted growth of *L. mesenteroides*, which should improve the flavor of the new type of Kimchi.

In the last part of the fermentation process, P. membranifaciens was higher, with more than 15×10^6 in control at all 3 preserving temperatures [Fig. 9(c)]. These bacteria produce a white membrane over the Kimchi, caused by its decomposition. But in Kimchi prepared with the common prescription, the number of P. membranifaciens decreased remarkably [Fig. 9(c)] at each preserving temperature. This is expected to increase the duration of preservation and keep the flavor of the Kimchi.

REFERENCES

- 1. Woo, S. J. and H. J. Lee (1987), Study on the Resazurin test as a rapid method for the estimation of maturity of Kimchi. *Korean J. Food Sci. Technol.* 19(3). 19-25.
- Choi, M. Y., E. J. Choi, E. Lee, B. C. Cha, H. J. Park, and T. J. Rhim (1996), Effect of pine needle (Pinus densiflora Seib. et Zucc) upon Kimchi fermentation. J. Korean Soc. Food Sci. Nutr. 25(6), 899-906.
- 3. Jang, K. S., M. J. Kim, Y. A. Oh, M. S. Kang, and S. D. Kim (1991), Changes in carotene content of Chinese cabbage Kimchi containing various submaterials and lactic acid bacteria during fermentation. J. Korean Soc. Food Nutr. 20(1), 5-12.
- 4. Son, Y. M., K. O. Kim, and D. W. Jeon (1996), The effect of low molecular weight chitosan with and without other preservatives on the characteristics of Kimchi during fermentation. *Korean J. food Sci. Technol.* **28**(5), 888-896.
- 5. Jang, K. S. (1988), Studies on the natural pH adjusters for Kimchi. J. Korean Soc. Food Sci. Nutr. 18(3), 321-327.
- 6. KIM, S. D. (1985), Effect of pH adjuster on the fermentation of Kimchi. J. Korean Soc. Food Sci. Nutr. 14(3), 259-264.
- 7. Kim, S. J., and K. H. Park (1995), Retardation of Kimchi fermentation by the extracts of *Allium tuberosum* and growth inhibition of related microorganisms *J. Korean Soc. Food Sci. Nutr.* 27(5), 813-818.
- 8. Choi, Y. B., H. J. Choi, H. S. Baik, and H. K. Jun (1997), Evaluation of optimum condition for the electrofusion between *Lactobacillus* sp. JC-7 isolated from Kimchi and *Lactobacillus* acidophilus88. Kor. J. Appl. Microbiol. Biotechnol. 25, 121-128.
- 9. Kim, K. O., H. A. Moon, and D. W. Jeon (1995), The effect of low molecular weight chitosans on the characteristics of Kimchi during fermentation. *Korean J. Food Sci. Technol.* 27(3), 420-427.
- 10. Min, T. I. (1988), Kimchi fermentation and microorganism. The Culinary Society of Korean Academy. 4(1), 96-105.
- 11. Park, K. J., and S. J. Woo (1988), Effect of Na-acetate, Na-malate and K-Sorbate on the pH, acidity and sourness during Kimchi fermentation. *Korean J. Food Sci. Technol.* **20**(1), 40-44.
- 12. Park, Y. H, and H. J. Song (1991), Antimicrobial activity of Lactobacillus plantarum Lp2 isolated from Kimchi. Kor J. Appl. Microbial. Biotechnol. 19(6), 637-643.
- 13. Lim, J. K., H. J. Seo, E. O. Kim, M. Meydani, and J. D. Kim (2006), Identification of anti-angiogenic and anti-cell adhesion

- materials from Enterobacteria of the Trachurus japonicus. J. Microbiol. Biotech. 16(10), 1544-1553.
- 14. Lee, S. H. and W. J. Choi (1998), Effect of medicinal herbs' extracts on the growth of lactic acid bacteria isolated from Kimchi and fermentation of Kimchi. *Korean J. Food Sci. Technol.* **30**(3), 624-629.
- 15. Kim, J. D., L. Liu, W. Guo, and M. Meydani (2006), Chemical structure of flavonolsids in relation to modulation of angiogenesis and immune-endothelial cell adhesion. *J. Nutrition. Biochem.* 17, 165-176.
- 16. Lee, Y. H. and I. W. Yang (1970), Studies on the packaging and preservation of Kimchi. J. Korean Agri. Chem. Soc. 13(3), 207-218.
- 17. Park M. K., K. S. Jung, and M. J. In (2004), Effects of Scutellaria baicalensis and Phellodendron amurenseextracts on growth of lactic acid bacteria and Kimchi fermentation. J Korean Soc. Food Nutr. 33, 420-426.
- 18. Vanderzant, C. and D. F. Splittstoesser (1992), Compendium of methods for the microbiological examination of foods, 3ed. American Health Association.
- 19. AlZoreky, N. and W. E. Sandine (1991), Aselective and differential agar medium. J. Food Sci. 56, 1729-1734.
- 20. Lee, M. K., W. S. Park, and K. H. Kang (1996), Selective media for isolation and enumararion of lactic acid bacteria from Kimchi. J. Korean. Soc. Food Sci. Nutr. 25(5), 754-760.
- 21. Han, H. U., C. R. Lim, and H. K. Park (1990), Determination of microbial community as an indicator of Kimchi fermentation, *Korean J. Food Sci. Technol.* 22(1), 26-32.
- 22. Kim, S. J. and K. H. Park (1995), Antimicrobial activities of the extracts of vegetable Kimchi stuff. *Korean J. Food Sci. Technol.* 27(2), 216-220.
- 23. Ko, Y. T. and I. H. Baik (2002), Changes in pH, sensory properties and volatile odor components of Kimchi by heating. *Korean J, Soc. Food Sci Nutr.* **34**(6), 1123-1126.
- 24. Lee, S. H, N. Y. Park, and W. J. Choi (1999), Changes of the lactic acid bacteria and selective inhibitory substances against Homo and Hetero lactic acid bacteria isolated from Kimchi. Kor. J. Appl. Microbiol. Biotechnol. 27(5), 410-414.
- 25. Park, Y. I. and D. H. Jo (1986), Microbial inhibition by an isolate of *Pediococcus* from Kimchi. J. Korean Agri. Chem.Soc. 29(2), 12-18.
- 26. Choi, I. K., S. H. Jung, B. J. Kim, S. Y. Park, J. Kim, and H. U. Han (2003), Novel leuconostoc citrunum starter culture system for fermentation of Kimchi, a fermented cabbage product. Antonie van Leeuwenhoek. 84, 247-253.
- 27. Kim, B., J. Lee, J. Jeon, J. Kim, and H. Han (2003), *Leuconostoc inhaesp.* nov., a lactic acid bacterium isolated from Kimchi. *Int. J. Syst. Evol. Microbiol.* 53, 1123-1126.
- 28. Han, H. U., C. R. Lim, and H. K. Park (1990), Determination of microbial community as an indicator of Kimchi fermentation. *Korean J. Food Sci. Technol.* **22**(1), 26-32.
- 29. Yoon, S. H. (1979), A Study on the antagonistic activity of *Enterobacteria* to lactic acid bacteria occurring kimchi fermentation. *K. J. N.* 12(1), 9-15.
- 30. Choi, S. Y., M. K. Lee, K. S. Choi, Y. J. Koo, and W. S. Park (1998), Changes of fermentation characteristics and sensory evaluation of Kimchi on different storage temperature. *Korean J. Food Sci. Technol.* **30**(3), 644-649.
- 31. No, H. K., I. K. Park, and S. D. Kim (1995), Extension of shelf-life of Kimchi by addition of chitosan during dalting. *J. Korean Soc. Food Nutr.* **24**(61), 932-936.
- 32. Lee, S. Y., E. O. Kim, H. J. Seo, M. Y. Kim, and J. D. Kim (2004) Development of new method for antioxidant capacity with ORP-pH system. *Biotechnol. Bioprocess Eng.* 9, 514-518.
- 33. Kim, S. D., S. H. Lee, M. J. Kim, and Y. A. Oh (1991), Changes in pectic substance of lower salted Chinese cabbage Kimchi with pH adjuster during fermentation. *J. Korean Soc. Food Nutr.* 17(3), 255-261.