

Combined Effects of Container Volume and Fertilizer Level on Plant Growth, Physiological Characteristics, and Nutrient Uptake of Vinca Plant (*Catharanthus roseus*)

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ABSTRACT The aim of our study was to investigate the interactive effects of container size and nutrient supply on plant growth, chlorophyll synthesis, transpiration, CO₂ assimilation, water use efficiency (WUE), and nutrient uptake of vinca plant (*Catharanthus roseus*). A complete experiment utilizing four concentrations of fertilizer and three volumes of containers was conducted. As the container size was increased, the plant height, leaf area, and dry weight of vinca significantly increased regardless of fertilizer level. The leaf area and dry weight of vinca were highly sensitive to the container size. However, the chlorophyll contents of vinca 20 days after the transplant significantly increased with decreasing container sizes and increasing fertilizer concentrations. Significant differences in transpiration and CO₂ assimilation occurred with the use of different fertilizer solutions, but the highest values for transpiration and CO₂ assimilation were in plants grown in the 15 cm-diameter containers. The highest water use efficiency was observed in the plants grown in 10 cm-containers with 4 dS/m of fertilizer, and there were no significant differences in WUE values among container sizes with fertilizer concentrations of 0, 1, or 2 dS/m. No significant difference in nutrient uptake was observed among the fertilizer levels or among the container sizes. However, at a fertilizer concentration of 4 dS/m, the uptake of several nutrients, including N, P, K, Ca, Mg, B and Fe, was higher in small containers than in larger ones.

Keywords : container volume, fertilizer level, plant growth, Vinca

Catharanthus roseus (L.) G. Don. is one of the most important medicinal plants, and an ornamental bedding plant belonging to the family Apocynaceae. Two varieties of *C. roseus* can be distinguished based on the flower color,

namely, the pink-flowered rosea and the white-flowered alba [Jaleel and Panneerselvam, 2007]. *C. roseus* plant is commercially important due to the presence of medicinal alkaloids and its ornamental value [Jaleel *et al.*, 2006]. It is a perennial tropical plant that produces more than 100 monoterpenoid indole alkaloids (MIAs), including two commercially important cytotoxic dimeric alkaloids used in cancer chemotherapy [Jaleel *et al.*, 2007c].

C. roseus derives its economic importance from the highly valued leaf anticancer alkaloids, vincristine and vinblastine, and the antihypertensive alkaloid ajmalicine found in its root [Srivastava and Srivastava, 2007; Karthikeyan *et al.*, 2007; Jaleel *et al.*, 2006]. The entire plant is rich in alkaloids, but the maximum concentrations of alkaloids are found in the root bark particularly during flowering. An infusion of leaves is used to treat menorrhagia, and the juice of the leaves is applied externally to relieve wasp stings. All parts of the plant have with hypoglycemic properties that are used to treat diabetes [Jaleel *et al.*, 2007a; Jaleel *et al.*, 2007d]. Several studies have already been carried out regarding medicinal importance [Jaleel *et al.*, 2006; Jaleel *et al.*, 2007e] and the growth regulatory effects of this plant [Jaleel *et al.*, 2007f; Jaleel *et al.*, 2007b].

Restrictions on the nutrient supply and container volumes affect plant vigor and size. A small root volume is commonly observed in intensive agriculture with drip irrigation. Pot culture without soil in intensive horticulture severely limits the growth and development of the plant root system (Bar Tal *et al.*, 1994a). A suitable restriction of root growth was suggested as a practical method for inducing early flowering to shorten the growing period of some field and vegetable crops and for their potential economic success of high density planting of crops, such as cotton (Carmi, 1986; Ben Porath

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and Baker, 1990), tomato (Ruff *et al.*, 1987), and sweet pepper (NeSmith *et al.*, 1992).

Reducing the soil volume often leads to dwarf plants (Carmi and Heuer, 1981; Flocker *et al.*, 1959; Richards and Rowe, 1977; Tschaplinski and Blake, 1985.), but there is also altered distribution of relative dry weight in inhibited plants, to indicate a reduction in total plant growth (Carmi, 1986; Krizek *et al.*, 1985; Richards and Rowe, 1977), however, root growth is more impaired in a restricted volume as compared to aerial part, creating a relative decrease in root dry weight (Carmi and Shalhevet, 1983). Plants grown in small containers characteristically develop a shorter, more densely branched root system than those grown in large containers; this, in turn, may affect the total plant growth because the roots are an important source of growth substances for the plant (Richards and Rowe, 1977).

No studies on the influence of the nutritional supply status on flowering and fruit set of crops have been reported. Moreover, the plant response to pot size is closely related to the nutritional supply. The restriction imposed by small container resulted in more short lateral roots and change in the root metabolism that affects the uptake rate of the nutrients and water both per unit of root area and for the entire plant (Peterson *et al.*, 1991a; Peterson *et al.*, 1991b). Plants with small root volumes need frequent irrigation. The aeration and hydraulic coefficients of the soil or growth medium decreased as the root density increases. Technically, if the root volume is smaller than the lowest theoretical size for sufficient nutrient absorption, it becomes impossible for the roots to supply enough nutrients for the plant (Bar Tal *et al.*, 1994a; Bar Tal *et al.*, 1994b).

Similarly, the influence of the nutritional supply status on the flower and fruit set of sweet pepper have also been reported. Sweet pepper growing in minimal media experienced a delay in flowering (50%) of 4-6 days and a delay in the initial fruit set by seven days, compared with sweet pepper grown in the field (Shrivastava, 1996). Nitrogen fertilization did not influence the fruiting time, mean fruit weight, or mid-harvest time of sweet pepper (Vos and Frinking, 1997).

The objective of this study was to evaluate the interactive effects of container volume and nutritional supply levels that are commonly adopted in greenhouse production on the growth and physiological characteristics of vinca plant.

MATERIALS AND METHODS

Plant material

Plug seedlings of Vinca 'Pacifica Punch' were obtained from Park Wholesale Inc. (Greenwood, SC) and transplanted into plastic pots on April 13, 2007. The pots were filled with peat-based growing medium (Fafard 2P, Fafard, Inc., Anderson, SC). The medium was composed of peat and perlite. Initial pH and electrical conductivity (EC) of nutrition solution were pH 5.98 and 0.61 dS/m, respectively. The pH was measured using a Model IQ150 Handheld pH/mV/Temperature meter (IQ Scientific Instruments, Inc., San Diego CA). The EC was measured using a Sigma Probe (Delta-T Devices Ltd, Burwell, Cambridge, UK). After transplantation, all the plants were placed on ebb and flow benches in a greenhouse. The temperatures of the greenhouse during day and night time were set to 24°C and 20°C respectively.

Cultural conditions

To determine the effects of pot size on plant growth, plants were transplanted into round pots with a 5, 10 or 15 cm diameter, having same height (20 cm). The plants were sub-irrigated daily with a fertilizer solution with an EC of 0.5, 1, 2 or 4 dS/m. The fertilizer solutions were made using a water-soluble N-P-K fertilizer (15-5-15, Peat-Lite Special, The Scotts Co.), and the pH of each fertilizer solution was not controlled. Water was pumped onto the benches using submersed water pumps (Nokorode-2; Little Giant, Oklahoma City, Okla.). About 3 min were required for the pumps to fill the trays with approximately 3 cm of fertilizer solution, and 6 min for the water to drain back into the holding tanks. The fertilizer solution was stored in 210 L plastic barrels and was replenished as needed. The EC of the fertilizer was adjusted every refilling time.

Measurement for physiological parameters

The EC and water content in the substrate was measured at regular intervals through out the growing period using a Sigma Probe EC meter (Delta-T Devices Ltd., Burwell, Cambridge, UK) and a Theta Probe soil moisture sensor (Delta-T Devices Ltd., Burwell, Cambridge, UK). Plant height, leaf area, dry weight and flower width of the vinca plant were measured at the end of experiment.

The chlorophyll content of the leaves was measured at regular intervals through out the growing period using a SPAD-502 chlorophyll meter (Minolta, Ramsey, NJ), which measures chlorophyll content in arbitrary units (here referred to as SPAD units).

The transpiration and CO₂ assimilation of individual pairs of vinca leaves were measured on the 15th day post transplantation. Leaf gas exchange parameters including the specific transpiration (*E*: evaporation) and CO₂ assimilation (*PN*) were measured with a portable photosynthesis system (CIRAS-1, PP Systems, Haverhill, Mass.). The water use efficiency (WUE) was calculated by the following equation: $WUE = PN [\mu\text{mol CO}_2 \text{ m}^{-2}\text{s}^{-1}] / E [\text{mmol H}_2\text{O m}^{-2}\text{s}^{-1}]$.

The remaining shoots were dried in a forced-air drying oven at 80°C for a minimum of 3 days before the dry mass was measured. The nutrient content of the shoot samples was analyzed. The tissue N was determined with a CNS 2000 analyzer (LECO Corp., St. Joseph, Mich.) (Mills and Jones, 1996), while P, K, Ca, Mg, S, Al, B, Cu, Fe, Mn, Na, and Zn were determined by dry ashing and by inductively coupled plasma (ICP) spectrometry (Jones and Case, 1990).

Statistics

The experimental design was a randomized complete block with three replicates. The data were analyzed by regression analysis and least significant differences (LSD) test using the Statistical Analysis Software (SAS Institute, Cary, NC).

RESULTS AND DISCUSSION

Plant growth

The restriction imposed by the container volume combined with restrictions in fertilization significantly affected all aspects of the vegetative growth including plant height and, especially leaf area (Fig. 1). With increasing container size, plant height and leaf area of vinca significantly increased at all fertilizer concentrations. Plant height (19.1 to 21.0 cm) and leaf area (196.2 to 341.9 cm²) in particular showed the highest values at a fertilizer concentration (as an EC value of) of 1.0 dS/m regardless of the container size and showed a stimulatory effect at lower concentration. However, at the highest concentration of 4.0 dS/m, all growth characteristics analyzed were lower in value compared with plants grown in

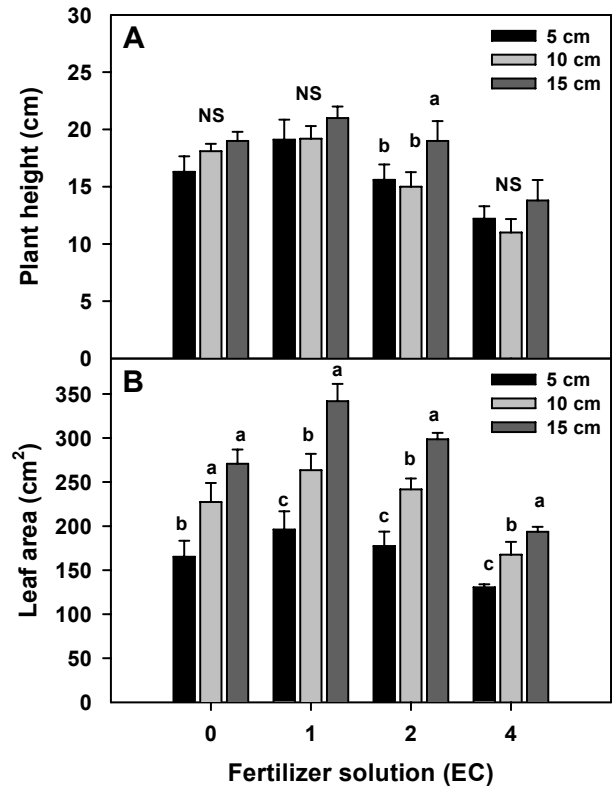


Fig. 1. Combined effects of container size and fertilizer solution on plant height (A) and leaf area (B) of vinca plant. Within each fertilizer solution, means followed by the same letter are not significantly different at $p < 0.05$. Each bar represents standard error of the mean.

the other lower fertilizer concentrations and compared with the untreated control. Small container volumes often produce smaller plants than do larger containers. In addition, plants in smaller containers tend to produce lower yields than comparable plantings established from larger container volumes (Marsh and Paul, 1988; Weston and Zandstra, 1986).

The dry weight was one of the most sensitive parameters affected by the container size. At all concentrations of fertilizer, the dry weight in small containers was significantly reduced. In particular, the dry weight was the highest (5.2 to 15.6 g plant⁻¹) at a fertilizer concentration of 1.0 dS/m, regardless of container size, and the lowest (2.3 to 8.3 g plant⁻¹) at 4.0 dS/m, which demonstrates the stimulatory effect of lower concentrations (Fig. 2A). On the other hand, the flower width of vinca grown in smaller containers was significantly reduced with increase in fertilizer concentration. Furthermore, flower width of vinca grown in larger containers

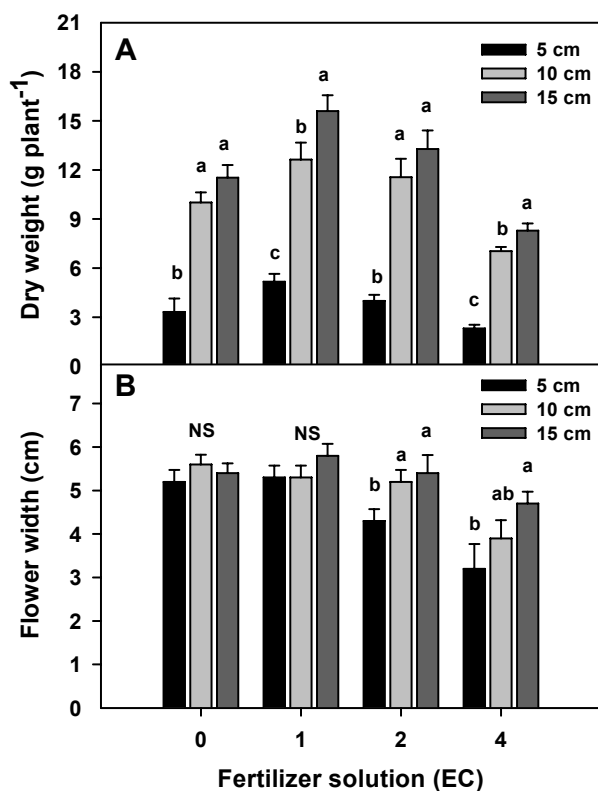


Fig. 2. Combined effects of container size and fertilizer solution on dry weight (A) and flower width (B) of vinca plant. Within each fertilizer solution, means followed by the same letter are not significantly different at $p < 0.05$. Each bar represents standard error of the mean.

showed the less reduction with increasing fertilizer concentration (Fig. 2B).

In general, as the container size increased, the plant leaf area, shoot biomass, and root biomass increased (Cantliffe, 1993). Root and shoot growth, biomass accumulation and partitioning, photosynthesis, leaf chlorophyll content, plant water relations, nutrient uptake, respiration, flowering, and yield are all affected by the root restriction due to container size.

Chlorophyll synthesis

The chlorophyll contents of vinca 20 days post transplantation significantly increased with decreasing container size and increasing fertilizer concentration (Fig. 3). With the highest fertilizer concentration of 4 dS/m, the chlorophyll content of vinca was higher with a 5 cm-diameter container (50.56 mgg^{-1} F.W.) than with 10 cm-diameter (43.56 mgg^{-1}

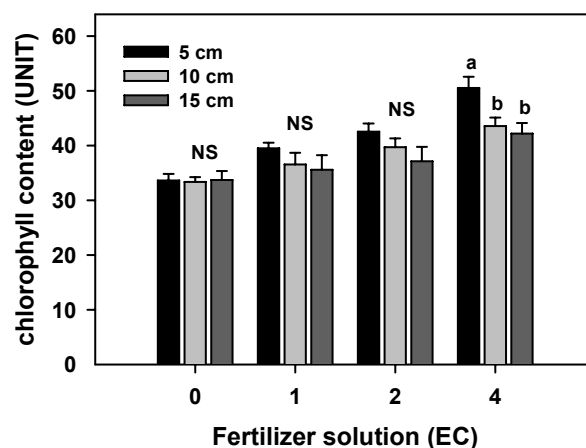


Fig. 3. Combined effects of container size and fertilizer solution on chlorophyll content of vinca plant. Within each fertilizer solution, means followed by the same letter are not significantly different at $p < 0.05$. Each bar represents standard error of the mean.

F.W.) and 15 cm-diameter containers (42.22 mgg^{-1} F.W.). However, there is no difference in chlorophyll content among container sizes with lower fertilizer concentration (Fig. 3). The restriction of container volume caused an increase in chlorophyll content in vinca leaves.

NeSmith *et al.* (1992) reported that the decline in the leaf photosynthetic rate of bell peppers in response to a decreased root volume was coupled with a reduction in leaf chlorophyll content.

Transpiration, CO₂ assimilation and WUE

There were significant differences in transpiration and CO₂ assimilation among the various fertilizer concentrations. Similarly, there were differences among container sizes but only the highest fertilizer concentration. The highest values occurred with 15 cm-diameter containers. Similarly, there was a significant difference in transpiration among fertilizer solutions. Especially, the highest transpiration value was found at a fertilizer concentration of 1 dS/m (2.15 to 3.05 $\text{mmol H}_2\text{O m}^{-2}\text{s}^{-1}$) and the lowest at 4 dS/m (0.21 to 1.12 $\text{mmol H}_2\text{O m}^{-2}\text{s}^{-1}$). The change in transpiration in plants grown in different container sizes showed significant differences with a fertilizer level of 4 dS/m only, and the highest values were obtained when plants were grown in a 15 cm-container (Fig. 4A). The CO₂ assimilation (PN) measured on the 15th day post transplantation was the highest at a

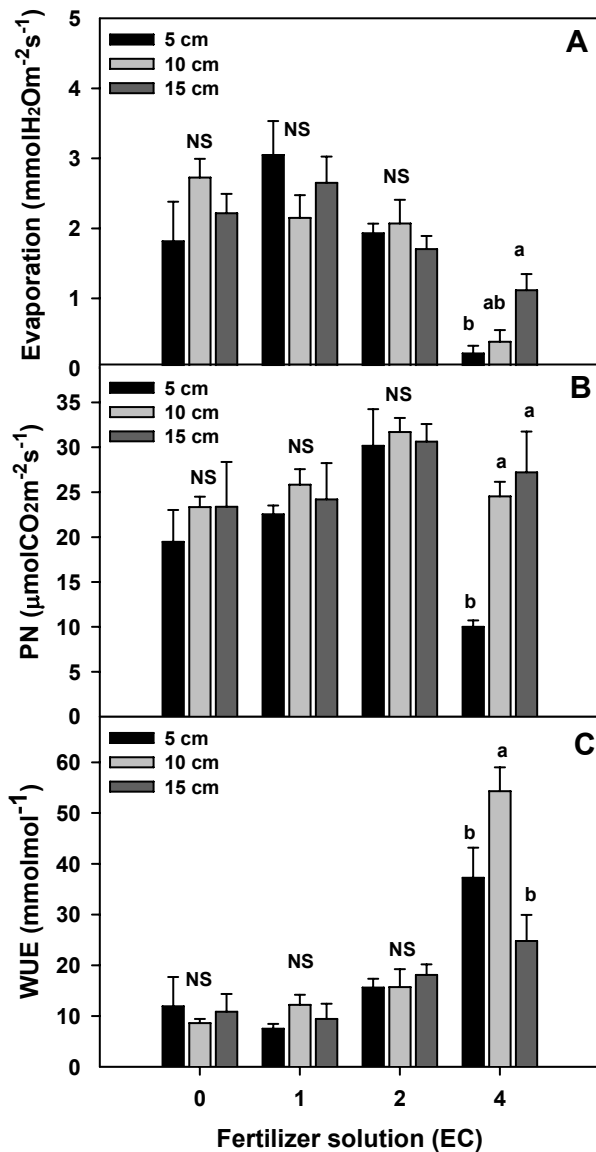


Fig. 4. Combined effects of container size and fertilizer solution on specific transpiration (Evaporation) (A), CO₂ assimilation (PN) (B) and water use efficiency (WUE) (C) of vinca plant. Within each fertilizer solution, means followed by the same letter are not significantly different at $p < 0.05$. Each bar represents standard error of the mean.

fertilizer concentration of 2 dS/m (30.63 to 31.70 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$) and the lowest was found at a concentration of 4 dS/m (10.00 to 27.20 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$). However, no significant difference in CO₂ assimilation was observed at 0, 1, and 2 dS/m (Fig. 4B). The WUE, on the other hand, showed the highest values in plants grown in 10 cm diameter-containers (54.32 mmolmol^{-1}) at a fertilizer concentration of 4 dS/m and

the lowest value of WUE found at this fertilizer concentration was found in plants grown in 15 cm diameter-containers. However, there was no significant difference in WUE among container sizes at 0, 1, and 2 dS/m of fertilizer (Fig. 4C).

The reduced plant biomass under root restricting conditions could possibly be due to a lower photosynthetic rate; however, few container size or root restriction experiments have measured photosynthetic rate. Our study indicates that the whole-plant photosynthetic rate decreases with increased root restriction in bell pepper. A similar trend was observed for the leaf photosynthetic rate, but to a lesser degree (NeSmith *et al.*, 1992).

Nutrition uptake

Below a fertilizer concentration of 2 dS/m, no significant difference in nutrient uptake by vinca plant was observed among fertilizer solutions and among container sizes. However, only at a fertilizer concentration of 4 dS/m, the uptake of several nutrients, including N, P, K, Ca, Mg, B, and Fe, was higher in plant grown in small containers vs. grown in large containers (Table 1).

The plant response to pot size is closely related to the nutritional supply. The restriction imposed by a small container produced high densities of short lateral roots and a change in the root metabolism that affects the nutrient and water uptake rate both per unit of root area for the whole plant (Peterson *et al.*, 1991a; Peterson *et al.*, 1991b). Technically, the supply of sufficient nutrients becomes impossible when the small root volume is smaller than the theoretical lower limit for nutrient element absorption (Bar Tal *et al.*, 1994a; Bar Tal *et al.*, 1994b).

CONCLUSION

The restriction of rhizosphere, particularly in combination with different levels of nutrient supply induces a reduction in plant growth, including decreased plant height, leaf area and dry weight, as well as a decrease in the flower width of vinca plant. The increased fertilizer concentration combined with the larger container volume significantly increased the leaf area and the dry weight. However, the chlorophyll content significantly increased with higher nutrient supplies and with the restriction of root volume. Specifically, the transpiration and CO₂ assimilation of vinca plant were significantly inhibited at

Table 1. Effects of nutrient and container size on macro- and micro-nutrient uptake of vinca plant.

Fertilizer solution (dS/m)	Container diameter (cm)	Macronutrient (%)						Micronutrient (%)				
		N	P	K	Ca	Mg	S	B	Cu	Fe	Mn	Zn
0	5	2.7	0.3	2.6	0.6	0.5	0.2	64.3	5.1	62.8	91.7	28.0
	10	2.5	0.3	2.3	0.6	0.5	0.2	28.0	4.4	34.8	92.1	31.2
	15	2.8	0.3	2.6	0.5	0.5	0.2	26.6	3.1	33.4	76.0	26.0
1	5	3.5	0.3	3.2	0.5	0.4	0.2	31.4	3.8	39.0	79.7	23.5
	10	3.8	0.3	3.2	0.5	0.4	0.2	30.6	4.6	23.3	81.7	30.6
	15	4.2	0.4	3.4	0.5	0.5	0.2	32.2	4.0	55.2	60.1	32.9
2	5	4.2	0.4	3.6	0.6	0.5	0.2	37.6	9.2	71.8	123.1	45.7
	10	4.4	0.4	3.5	0.5	0.4	0.2	32.2	9.2	52.9	122.8	50.8
	15	4.2	0.4	3.6	0.6	0.4	0.2	29.3	8.2	68.0	111.0	55.6
4	5	5.8	0.5	4.4	1.1	0.8	0.2	54.3	5.5	95.4	114.1	34.4
	10	5.0	0.4	3.4	0.7	0.4	0.2	45.1	7.0	72.7	132.9	45.1
	15	4.9	0.4	3.2	0.5	0.4	0.2	38.3	6.7	61.2	100.8	52.4

the highest fertilizer concentration (4 dS/m), while the transpiration and CO₂ assimilation were enhanced at the lower fertilizer concentrations regardless of container volume. On the other hand, the WUE of vinca plant showed the highest value at the highest fertilizer concentration, with significant differences depending on container volume. Finally, significant differences in N, P, K, Ca, Mg, B, and Fe uptake were observed only at a fertilizer concentration of 4 dS/m, and higher uptake was observed when plants were grown in small containers compared with plants grown in large containers. In conclusion, our findings indicate that root restriction combined with increase in fertilizer concentration reduced the vegetative and reproductive growth, changed some physiological characteristics, enhanced the chlorophyll synthesis and the nutrient uptake of the vinca plant.

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REFERENCES

- Bar Tal, A., A. Feigin, I. Rylski, and E. Pressman. 1994a. Root pruning and N-NO₃ solution concentration effects on tomato plant growth and fruit yield. *Sci. Hort.* 58 : 91-103.
- Bar Tal, A., A. Feigin, I. Rylski, and E. Pressman. 1994b. Root pruning and N-NO₃ solution concentration effects on nutrient uptake and transpiration by tomato. *Sci. Hort.* 58 : 77-90.
- Ben Porath, A. and D.N. Baker. 1990. Taproot restriction effects on growth, earliness, and dry weight partitioning of cotton. *Crop Sci.* 30: 809-814.
- Cantliffe, D. J. 1993. Pre- and postharvest practices for improved vegetable transplant quality. *HortTechnology* 3 : 415-417.
- Carmi, A. 1986. Effects of root zone volume and plant density on the vegetative and reproductive development of cotton. *Field Crop Res.* 13 : 25-32.
- Carmi, A. and B. Heuer. 1981. The role of roots in control of bean shoot growth. *Ann. Bot.* 48 : 519-527.
- Carmi, A. and J. Shalhevet. 1983. Root effects on cotton growth and yield. *Crop Sci.* 23 : 875-878.
- Flocker, W. J., J. A. Vomocill, and F. D. Howard. 1959. Some growth responses of tomato to soil compaction. *Soil Sci. Soc. Amer. Proc.* 23 : 188-191.
- Jaleel, C. A., and R. Panneerselvam. 2007. Variations in the antioxidative and indole alkaloid status in different parts of two varieties of *Catharanthus roseus*, an important folk herb. *Chin. J. Pharmacol. Toxicol.* 21 : 487-494.
- Jaleel, C. A., P. Manivannan, A. Kishorekumar, B. Sankar, and R. Panneerselvam. 2007a. Calcium chloride effects on salinity induced oxidative stress, proline metabolism and indole alkaloid accumulation in *Catharanthus roseus*. *C. R. Biol.* 330 : 674-683.
- Jaleel, C. A., P. Manivannan, B. Sankar, A. Kishorekumar, R. Gopi, R. Somasundaram, and R. Panneerselvam. 2007b. *Pseudomonas fluorescens* enhances biomass yield and

- ajmalicine production in *Catharanthus roseus* under water deficit stress. *Colloids Surf. B: Biointerf.* 60 : 7-11.
- Jaleel, C. A., R. Gopi, B. Sankar, P. Manivannan, A. Kishorekumar, R. Sridharan, and R. Panneerselvam. 2007c. Alterations in germination, seedling vigour, lipid peroxidation, and proline metabolism in *Catharanthus roseus* seedlings under salt stress. *South Afr. J. Bot.* 73 : 190-195.
- Jaleel, C. A., R. Gopi, G. M. Alagu Lakshmanan, and R. Panneerselvam. 2006. Triadimefon induced changes in the antioxidant metabolism and ajmalicine production in *Catharanthus roseus* (L.) G. Don. *Plant Sci.* 171 : 271-276.
- Jaleel, C. A., R. Gopi, P. Manivannan, and R. Panneerselvam. 2007d. Responses of antioxidant defense system of *Catharanthus roseus* (L.) G. Don. to paclobutrazol treatment under salinity. *Acta Physiol. Plant.* 29 : 205-209.
- Jaleel, C. A., R. Gopi, P. Manivannan, and R. Panneerselvam. 2007e. Antioxidative potentials as a protective mechanism in *Catharanthus roseus* (L.) G. Don. plants under salinity stress. *Turkish J. Bot.* 31 : 245-251.
- Jaleel, C. A., R. Gopi, P. Manivannan, B. Sankar, A. Kishorekumar, and R. Panneerselvam. 2007f. Antioxidant potentials and ajmalicine accumulation in *Catharanthus roseus* after treatment with gibberellic acid. *Colloids Surf. B: Biointerf.* 60 : 195-200.
- Jones, J. B. and V. W. Case. 1990. Sampling, handling, analyzing plant tissue samples. p. 389-427. *In* R. L. Westerman (ed.) *Soil testing and plant analysis*. 3rd ed. SSSA, Madison, WI.
- Karthikeyan, B., C. A. Jaleel, R. Gopi, and M. Deiveekasundaram. 2007. Alterations in seedling vigour and antioxidant enzyme activities in *C. roseus* under seed priming with native diazotrophs. *J. Zhejiang Univ. Sci.* B8 : 453-457.
- Krizek, D. T., A. Carmi, R. M. Mirecki, F. W. Snyder, and J. A. Bunce. 1985. Comparative effects of soil moisture stress and restricted root zone volume on morphogenetic and physiological response of soybean (*Glycine max.*(L.) Merr.). *J. Expt. Bot.* 36 : 25-38.
- Marsh, D. B. and K. B. Paul. 1988. Influence of container type and cell size on cabbage transplant development and field performance. *HortScience* 23 : 310-311.
- Mills, H. A. and J. B. Jr. Jones. 1996. *Plant Analysis Handbook II. A practical sampling, preparation, analysis, and interpretation guide*. Athens, GA : Micromacro Publishing. 422 pp.
- NeSmith, D. S., D. C. Bridges, and J. C. Barbour. 1992. Bell pepper responses to root restriction. *J. Plant Nutr.* 15 : 2763-2776.
- Peterson, T. A., M. D. Reinsel, and D. T. Krizek. 1991a. Tomato (*Lycopersicon esculentum* Mill., cv. 'Better Bush') plant response to root restriction. I. Alteration of plant morphology. *J. Exp. Bot.* 42 : 1233-1240.
- Peterson, T. A., M. D. Reinsel, and D. T. Krizek. 1991b. Tomato (*Lycopersicon esculentum* Mill., cv. 'Better Bush') plant response to root restriction. II. Root respiration and Ethylene Generation. *J. Exp. Bot.* 42 : 1241-1249.
- Richards, D. and R. N. Rowe. 1977. Effects of root restriction, root pruning and 6-benzylaminopurine on the growth of peach seedling. *Ann. Bot.* 41 : 729-740.
- Ruff, M. S., D. T. Krizek, R. M. Mirecki, and D. W. Inouye. 1987. Restricted Root Zone Volume : Influence on Growth and Development of Tomato. *J. Amer. Soc. Hort. Sci.* 112 : 763-769.
- Shrivastava, A. K. 1996. Effect of fertilizer levels and spacings on flowering, fruit set and yield of sweet pepper (*Capsicum annuum* var. *grossum* L.) cv. Hybrid Bharat. *Adv. Plant Sci.* 9 : 171-175.
- Srivastava, N. K. and A.K. Srivastava. 2007. Influence of gibberellic acid on ¹⁴CO₂ metabolism, growth, and production of alkaloids in *Catharanthus roseus*. *Photosynthetica.* 45 : 156-160.
- Tschaplinski, T. J. and T. J. Blake. 1985. Effects of root restriction on growth correlations, water relations and senescence of alder seedling. *Physiol. Plant.* 64 : 167-176.
- Vos, J. G. M. and H. D. Frinking. 1997. Nitrogen fertilization as a component of integrated crop management of hot pepper (*Capsicum* spp.) under tropical lowland conditions. *Intern. J. Pest. Manag.* 43 : 1-10.
- Weston, L. A. and B. H. Zandstra. 1986. Effect of root container size and location of production on growth and yield of tomato transplants. *J. Amer. Soc. Hort. Sci.* 11 : 498-501.