# Anti-oxidative Activities of Phenolic Compounds from barks of *Pinus densiflora* Siebold et Zuccarini

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**Abstract** – Phytochemical examination of the barks of *Pinus densiflora* Siebold et Zuccarini has led to the isolation of one phenylpropanoid, one lignan, one flavonoid, one flavan 3-ol and two procyanidins :  $4 \cdot O \cdot \beta \cdot D$ -glucopyranosyl- *p*-coumaric acid (1), 2,3-dihydro-2-(4-methoxy)-7-hydroxy-3-hydroxymethyl-5-(3-hydroxy propyl)-benzofuran  $3 \cdot O \cdot \alpha \cdot D$ -glucopyranoside (2), taxifolin  $3' \cdot O \cdot \beta \cdot D$ -glucopyranoside (3), (+)-catechin (4), procyanidin B1 (5) and epicatechin-(4 $\beta$ -8)-catechin-(4 $\alpha$ -8)-catechin (6). Among them, Compound 4, 5 and 6 showed potent anti-oxidative activities and these anti-oxidative activities were significantly different compared with ascorbic acid as positive control.

Keywords - Pinus densiflora, antioxidative activity, lignan, flavonoid, procyanidin, DPPH

#### Introduction

The barks of *Pinus densiflora* Siebold et Zuccarini have been used for Korean traditional medicine as the remedies for bleeding, rheumatitis, bruise and scald, etc. From the barks of *Pinus densiflora* several phenoic compounds, icariside E4 (Miyase *et al.*, 1989), (+)isolariciresinol 9'-O- $\beta$ -D-xylopyranoside, and (+) isolariciresinol (Hans *et al.*, 1992), quercetin 3-O- $\beta$ -D-glucopyranoside (Lee and Bae, 2001), 5,7,8,4'-tetrahydroxy-3methoxy-6-methylflavone 8-O- $\beta$ -D-glucopyranoside (Jung *et al.*, 2001), and shikimic acid (Ishimaru *et al.*, 2002), dihydroquercetin-3'-O- $\beta$ -galactoside (Kim *et al.*, 1991), (+)-catechin, catechin-(4 $\alpha$ -6)-catechin, epicatechin-(4 $\beta$ -6)-catechin and catechin-(4 $\alpha$ -6)-catechin, (4 $\beta$ -6)-catechin (Song and Oh, 1996) were isolated.

Pycnogenol which is polyphenols containing extract from the barks of *Pinus maritima* was known to have anti-inflammatory activities including inhibitory effect of NO production (Fabio *et al.*, 1998), inhibition of inflammatory mediator from macrophage (Erben *et al*, 2000), inhibition of interlukin-1, proinflammatory cytokine (Cho *et al.*, 2000) and anti-oxidative effect (Packer *et al.*, 1999). As parts of our continuing search for new antioxidative and anti-inflammatory agents from natural sources, we isolated one phenylpropanoid, one lignan, one flavonoid, one flavan 3-ol and two procyanidins from the

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barks of *Pinus densiflora* and evaluated their antioxidative activities.

## **Experimental**

General Experimental Procedures – The NMR spectra were recorded on Varian Gemini 2000, 300 MHz (USA) and Bruker AMX-500, 500 MHz (Germany) spectrometer, using internal standard pulse program, with chemical shifts reported in ppm downfield from TMS. Column chromatography was carried out on Sephadex LH-20 (25 - 100  $\mu$ m, Pharmacia, Sweden), MCI-gel CHP20P (75~150  $\mu$ m, Mitsubishi, Japan) and YMC-gel ODS-A (230/70, 400/230, 500/400 mesh, YMC, Japan). TLC was performed on aluminum plates pre-coated with Kieselgel 60F254 (Merck). All other chemicals and solvents were of analytical grade.

**Plant Material** – The barks were prepared from the *Pinus densiflora* collected from Huk-seuk Dong, Seoul, South Korea, on September, 2008). A voucher specimen (PD2008) was deposited at the herbarium of the College of Pharmacy, Chung-Ang University.

**Extraction and Isolation** – The fresh of barks of *Pinus densiflora* (1.5 kg) were extracted three times with 80% aqueous acetone at room temperature. After removal of Me<sub>2</sub>CO in vacuo, the aqueous solution was filtered. The filtrate was concentrated and then applied to a column of Sephadex LH-20. Elution with H<sub>2</sub>O containing increasing proportion of MeOH afforded 5 fractions. Repeated

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column chromatography of fraction 2, MCI gel CHP 20P with a gradient solvent system of  $H_2O$ : MeOH (100 :  $0 \rightarrow 0$  : 100) and low pressure liquid column chromatography yielded 2 (200 mg). Repeated column chromatography of fraction 3, low pressure liquid column chromatography with a gradient solvent system of  $H_2O$ : MeOH (100 : 0  $\rightarrow$  0 : 100) and MCI gel with a gradient solvent system of  $H_2O$  : MeOH (100 :  $0 \rightarrow 0$  : 100) yielded 1 (40 mg). Repeated column chromatography fraction 4, followed by MCI with a gradient solvent system of H<sub>2</sub>O : MeOH (100 :  $0 \rightarrow 0$  : 100) yielded 3 (1.5 g). Repeated column chromatography fraction 5, followed by MCI-gel with a gradient solvent system of  $H_2O$ : MeOH (100 : 0  $\rightarrow$  0 : 100), and followed by low pressure liquid column chromatography with a gradient solvent system of  $H_2O$ : MeOH (100 : 0  $\rightarrow$  0 : 100),  $(90:10 \rightarrow 50:50)$  yielded 4 (300 mg), 5 (150 mg), and 6 (100 mg).

**4-O-β-D-Glucopyranosyl-** *p*-coumaric aicd (1) – White amorphous powder;  $[\alpha]_D{}^{20}$ : -59.1° (*c* = 0.01, MeOH), IR v<sup>KBr</sup><sub>Max</sub> cm<sup>-1</sup>: 3400 (OH), 1452, 1550 (aromatic C = C), <sup>1</sup>H-NMR (500 MHz, MeOH-*d*<sub>4</sub> + D<sub>2</sub>O): δ 3.31-3.91 (5H in total, m, glc-3,5,2,4,6), 4.96 (1H, d, *J* = 7.5Hz, g-1), 6.40 (1H, d, *J* = 15.9Hz, H-1'), 7.08 (2H, d, *J* = 8.7Hz, H-3, 5), 7.39 (1H, d, *J* = 15.9Hz, H-2'), 7.49 (2H, d, *J* = 8.7Hz, H-2, 6). <sup>13</sup>C-NMR (125MHz, MeOH-*d*<sub>4</sub> + D<sub>2</sub>O): δ 176.0 (COOH), δ 159.7 (C-4), δ 141.2 (C-1'), δ 131.5 (C-1), δ 130.1 (C-2, 6), δ 124.5 (C-2'), δ 118.0 (C-3, 5), δ 102.0 (C-1''), δ 78.1 (C-5''), δ 77.8 (C-3''), δ 74.8 (C-2''), δ 71.3 (C-4''), δ 62.4 (C-6'').

## 2,3-Dihydro-2-(4-methoxy)-7-hydroxy-3-

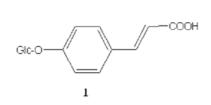
hydroxymethyl-5-(3-hydroxypropyl)benzofuran  $3-O-\beta$ -L-glucopyranoside (2) – White amorphous powder;  $[\alpha]$  $_{D}^{20}$ : -24.3° (c = 0.01, MeOH), IR v<sup>KBr</sup><sub>Max</sub> cm<sup>-1</sup>: 3400 (OH), 2840, 2800 (C-C), 1500, 1450 (aromatic C = C), Negative FAB MS : m/z 508 [M – H]<sup>-</sup>; <sup>1</sup>H-NMR (500 MHz, Me<sub>2</sub>CO- $d_6$  + D<sub>2</sub>O) :  $\delta$  1.73 (2H, m, H-8'), 2..51 (2H, t, J = 7.5, 8 Hz, H-7'), 3.43-3.85 (8H in total, m, glu-3,5,2,4,6, H-8, 9a, 9b), 3.81 (3H, s, OCH<sub>3</sub>), 4.91 (1H, d, J = 7.3 Hz, glu-1), 5.55 (1H, d, J = 5.7 Hz, H-7), 6.60 (2H, d, J = 12.2, H-2', 6'), 6.93 (1H, dd, J = 8.4, 1.2 Hz, H-6), 7.08 (1H, d, J=1.2Hz H-2), 7.12 (1H, d, J=8.4 Hz H-5). <sup>13</sup>C-NMR (125MHz, Me<sub>2</sub>CO- $d_6$  + D<sub>2</sub>O) :  $\delta$  150.0 (C-4), δ 147.0 (C-3), δ 145.9 (C-3'), δ 141.7 (C-2'), δ 137.9 (C-1), δ 136.4 (C-10), δ 129.4 (C-4'), δ 119.0 (C-6), δ 116.9 (C-5), δ 116.8 (C-1'), δ 116.3 (C-5'), δ 110.1 (C-2), δ 102.1 (OCH<sub>3</sub>), δ 87.5 (C-7), δ 77.6 (C-4"), δ 77.2 (C-2"), δ 74.3 (C-1"), δ 70.8 (C-3"), δ 64.7 (C-9), δ 62.2 (C-5"), δ 61.7 (C-8'), δ 56.5 (C-9'), δ 55.3 (C-8), δ 35.5 (C-7'), δ 32.3 (C-6').

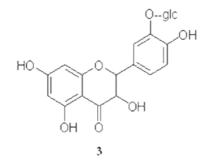
**Taxifolin 3'-***O*- $\beta$ **-D**-glucopyranoside (3) – Brown amorphous powder;  $\left[\alpha\right]_{D}^{20}$ : +22.1° (c = 0.01, MeOH), IR v<sup>KBr</sup><sub>Max</sub> cm<sup>-1</sup> : 3400 (OH), 2840, 2800 (C-C), 1550, 1450 (aromatic C = C), Negative FAB MS : m/z 467 [M – H]<sup>-</sup>; <sup>1</sup>H-NMR (300 MHz, Me<sub>2</sub>CO- $d_6$  + D<sub>2</sub>O) :  $\delta$  3.31-3.92 (5H in total, m, glc-3,5,2,4,6), 4.67 (1H, d, J = 11.4Hz, H-3), 4.88 (1H, d, J = 7.5Hz, glc-1), 5.07 (1H, d, J = 11.4Hz, H-2), 5.96 (1H, d, J=2.1Hz, H-6), 5.99 (1H, d, J=2.1Hz, H-8), 6.93 (1H, d, J = 8.1Hz, H-5'), 7.13 (1H, dd, J = 8.1, 2.1Hz, H-6'), 7.45 (1H, d, J = 2.1Hz, H-2'). <sup>13</sup>C-NMR  $(75 \text{ MHz}, \text{ Me}_2\text{CO-}d_6 + D_2\text{O}) : \delta 198.4 \text{ (C-4)}, \delta 168.4 \text{ (C-}$ 7), δ 165.0 (C-5), δ 164.2 (C-9), δ 149.2 (C-4'), δ 146.1 (C-3'), δ 129.7 (C-1'), δ 124.8 (C-6'), δ 118.9 (C-2'), δ 116.8 (C-5'), δ 104.0 (C-1"), δ 101.5 (C-10), δ 97.2 (C-6), δ 96.2 (C-8), δ 84.3 (C-2), δ 78.0 (C-5"), δ 77.4 (C-3"), δ 74.6 (C-2"), δ 72.9 (C-3), δ 71.1 (C-4"), δ 62.4 (C-6").

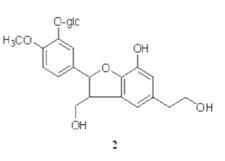
(+)-Catechin (4) – Brown amorphous powder;  $[\alpha]_D^{20}$ : +14.7° (*c* = 0.01, MeOH), IR  $v^{\text{KBr}}_{\text{Max}}$  cm<sup>-1</sup>: 3329 (OH), 1628 (C = C), 1550, 1452 (aromatic C = C), Negative FAB MS : *m*/*z* 289 [M – H]<sup>-</sup>; <sup>1</sup>H-NMR (300 MHz, Me<sub>2</sub>CO-*d*<sub>6</sub> + D<sub>2</sub>O) :  $\delta$  2.51 (1H, dd, *J* = 16.2, 8.7 Hz, H-4ax), 2.92 (1H, dd, *J* = 16.2, 5.4 Hz, H-4eq), 4.00 (1H, m, H-3), 4.56 (1H, m, *J* = 8.1Hz, H-2), 5.88 (1H, d, *J* = 2.4 Hz, H-6), 6.04 (1H, d, *J* = 2.4 Hz, H-8), 6.75 (1H, dd, *J* = 8.1, 1.8 Hz, H-6'), 6.79 (1H, d, *J* = 8.1, H-5'), 6.92 (1H, d, *J* = 1.8 Hz, H-2'). <sup>13</sup>C-NMR (75MHz, Me<sub>2</sub>CO*d*<sub>6</sub> + D<sub>2</sub>O) :  $\delta$  157.9 (C-7),  $\delta$  157.4 (C-9),  $\delta$  157.0 (C-5),  $\delta$ 145.9 (C-3'),  $\delta$  145.8 (C-4'),  $\delta$  132.1 (C-1'),  $\delta$  120.1 (C-6'),  $\delta$  115.8 (C-2'),  $\delta$  115.4 (C-5'),  $\delta$  100.6 (C-10),  $\delta$  96.2 (C-6),  $\delta$  95.3 (C-8),  $\delta$  82.7 (C-2),  $\delta$  68.3 (C-3),  $\delta$  28.8 (C-4).

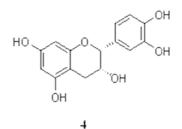
**Procyanidin B-1 (5)** – Brown amorphous powder; [α]  $_{D}^{20}$ : -12.4° (*c* = 0.01, MeOH), IR v<sup>KBr</sup> Max cm<sup>-1</sup>: 3467 (OH), 1620, 1550 (aromatic C = C), Negative FAB MS : *m/z* 579 [M – H]<sup>-</sup>; <sup>1</sup>H-NMR (300 MHz, Me<sub>2</sub>CO-*d*<sub>6</sub> + D<sub>2</sub>O) : δ 2.60 (1H, dd, H-4t ax), 2.80 (1H, dd, H-4t eq), 3.98 (1H, m, H-3t), 4.12 (1H, s, H-3u), 4.66 (1H, s, H-4u), 4.80 (1H, s, H-2t), 5.10 (1H, s, H-2u), 5.99-6.04 (3H, in total, H-8u, 6u, 6t), 6.71-7.01 (6H, in total, H-2', 5', 6' ut). <sup>13</sup>C-NMR (75 MHz, Me<sub>2</sub>CO-*d*<sub>6</sub> + D<sub>2</sub>O) : 27.5 (C-4t), 36.7 (C-4u), 67.7 (C-3t), 72.5 (C-3u),6.9 (C-2u), 81.9 (C-2t), 95.5 (C-8u), 96.1 (C-6u), 96.9 (C-6t), 101.0 (C-10u,t), 107.7 (C-8t), 114.8-115.9 (C-2',5'u,t), 119.3-119.4 (C-6'u,t), 132.1-132.4 (C-1'u,t), 145.2-145.6 (C-3',4'u,t), 155.4-157.7 (C-9,7,5 u,t)

**Epicatechin** (4β-8)-catechin (4α-8)-catechin (6) – Brown amorphous powder;  $[\alpha]_D{}^{20}:-97.4^\circ$  (c = 0.01, MeOH), IR v<sup>KBr</sup><sub>Max</sub> cm<sup>-1</sup>:3467 (OH), 1620, 1550 (aromatic C = C), Negative FAB MS : m/z 867 [M – H]<sup>-</sup>; <sup>1</sup>H-NMR (300 MHz, Me<sub>2</sub>CO- $d_6$  + D<sub>2</sub>O) : δ 2.60 (1H, dd, H-4t ax), 2.90 (1H, dd, H-4t eq), 4.04-4.06 (3H, in total,









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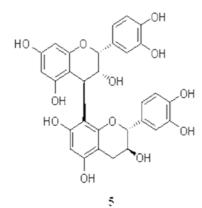
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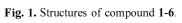
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H-3u, 3m, 3t), 4.48-4.72 (5H, in total, H-4u, 4m, 2u, 2m, 2t), 5.95-6.20 (4H, in total, H-8u, 6u, 6m, 6t), 6.68-7.08 (9H, in total, H-2', 5', 6' u,m,t). <sup>13</sup>C-NMR (75 MHz, Me<sub>2</sub>CO- $d_6$  + D<sub>2</sub>O) : 29.1 (C-4t), 36.6 (C-4u), 38.1 (C-4m), 68.2 (C-3t), 72.9 (C-3 u,t), 76.9 (C-2u), 82.9 (C-2t), 84.1 (C-2m), 95.8 (C-6 u,m,t, 8u), 100.6 (C-10 u,m,t), 107.7-108.3 (C-8 m,t), 115.1-115.9 (C-2',5'u,m,t), 119.1-120.6 (C-6'u,m,t), 131.7-132.2 (C-1'u,m,t), 145.7-145.9 (C-3',4'u,m,t), 155.2-157.4 (C-9,7,5 u,m,t)

### **Biological Assay**

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**DPPH radical scavenging activity** – The antioxidant activities of the six compounds were determined on the basis of the scavenging activity of the stable 1,1-diphenyl-2-picrylhydrzyl (DPPH) free radical by a previously described method with a slight modification (Hatano *et al.*, 1989). 20  $\mu$ l of the compound in EtOH (99.5% ethanol) was added to 180  $\mu$ l of a DPPH solution (7.887 mg DPPH in 200 mL EtOH). After mixing gently and standing at for 30 min, the optical density was measured

Table 1. Anti-oxidative Activities of Compound 1 - 6

Compounds	IC <sub>50</sub> (µg/ml)
1	None
2	35.79
3	22.97
4	3.91
5	2.98
6	3.44
Ascorbic acid	5.28

at 515 nm using an ELISA reader (BIO-RAD, Japan). The free radical scavenging activity was expressed as follow:

DPPH scavenging activity (%) =  $\{(Ac - As)/(Ac - Ab)\} \times 100$ 

When Ac was the absorbance of the control, As was the absorbance of the sample and Ab was the absorbance of the blank. The  $IC_{50}$  values were defined as the concentration that could scavenge 50% DPPH free radical. L-ascorbic acid was used as positive control.

## **Results and Discussion**

The fresh barks of *Pinus densiflora* were extracted with 80% Me<sub>2</sub>CO and the extract was subjected to a combination of sephadex LH-20 for the fractionations. And repeated column chromatography on each fraction to afford  $4-\beta$ -D-glucopyranosyloxyl-*p*-coumaric acid (1, Park et al., 1996), 2,3-dihydro-2-(4-methoxy)-7-hydroxy-3-hydroxymethyl-5-(3-hydroxypropyl)-benzofuran  $3-O-\beta$ -D-glucopyranoside (2, Lundgren et al., 1981), taxifolin 3'- $O-\beta$ -D-glucopyranoside (3, Oleszek et al., 2002), (+)catechin (4, Choi et al., 2001), procyanidin B-1 (5, Lee et al., 1992) and epicatechin (4 $\beta$ -8)-catechin (4 $\alpha$ -8)-catechin (6, Hsu et al., 1985) and their structures were identified by the comparisons with the reported spectral and physical data in the literatures. The anti-oxidative activities were tested by DPPH radical scavenging method. Especially, compound 4 (IC<sub>50</sub> =  $3.91 \mu g/ml$ ) which is flavan-30l and 5 (IC<sub>50</sub> = 2.98  $\mu$ g/ml) and 6 (IC<sub>50</sub> = 3.44 µg/ml) which are procyanidin exhibited potent of free radical scavenging activity compared with positive control, ascorbic acid (IC<sub>50</sub> =  $5.28 \mu g/ml$ ). But compound 1 which is phenylpropanoid did not exhibit DPPH radical scavenging activity. Compound 2 ( $IC_{50} = 35.79 \ \mu g/ml$ ) and 3 (IC<sub>50</sub> = 22.97  $\mu$ g/ml) which are lignan and flavonoid showed mild DPPH radical scavenging activities compared with 4, 5 and 6 (Table 1.). These results showed that flavan 3-ol (4) and procyanidins (5 and 6) have

potent free radical scavenging activities and the barks of *Pinus densiflora* is good source as anti-oxidative agent.

## Acknowledgements

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