

Evaluation of Immunotoxicity of Shizukaol B Isolated from *Chloranthus japonicus*

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Abstract

Dimeric sesquiterpenoid shizukaol B (SKB) was isolated from *Chloranthus japonicus* Sieb. Except that SKB inhibited adhesion molecule expression in monocytes and endothelial cells, no more biological and pharmacological activity of SKB had been reported until now. In this study, we examined immunosuppressive activity of SKB. SKB strongly inhibited lipopolysaccharide (LPS)-induced B cell proliferation with IC₅₀ of 137 ng/ml, but slightly or not concanavalin A-induced T cell proliferation, LPS-induced macrophage NO production, and LPS-induced dendritic cell maturation. As a mechanism, SKB strongly induced apoptotic death of B cells, but not other cell types. These results suggested that SKB induced toxicity-mediated immunosuppression against B cells.

Key Words: Shizukaol B, *Chloranthus japonicus*, Immunosuppression, Cytotoxicity, B cells

INTRODUCTION

The genus *Chloranthus* has been taxonomically placed in the Chloranthaceae and has long been used as folk medicine for their antitumor, antifungal, and anti-inflammatory activities (Kuang *et al.*, 2008). The components of genus *Chloranthus* are volatile oil, simple coumarins, amide alkaloids and sesquiterpene lactones having a lindenane skeleton named shizukanolides and chloranthalactones (Kuang *et al.*, 2009). *Chloranthus japonicus* is a perennial herb that grows in the southern part of Korea, Japan, and China (Kwon *et al.*, 2006). As a biologically active substance from this plant, dimeric sesquiterpenoid shizukaol B (SKB) inhibited expression of several adhesion molecules, such as intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1) and E-selectin, in HL-60 and human umbilical vein endothelial (HUVEC) cells, suggesting that shizukaol B might inhibit monocytes adhesion to endothelial cells, which was a critical step in the pathogenesis of atherosclerosis (Kwon *et al.*, 2006). Except this data, there were no reports on the pharmacological or biological activity of SKB until now. In this study, we investigated immunosuppressive activity of SKB and demonstrated that SKB might induce apoptosis-mediated immunosuppression. Of note was that SKB preferentially in-

duced apoptosis in B cells, but not T cells, macrophages, and dendritic cells.

MATERIALS AND METHODS

Materials

Female C57BL/6 mice (6-8 weeks old) were obtained from Korea Research Institute of Bioscience and Biotechnology (Chungbuk, Korea). Mice were housed in specific pathogen-free conditions at 21-24°C and 40-60% relative humidity under a 12-hr light/dark cycle. All animals were acclimatized for at least 1 week prior to the experiments. The experimental procedures used in this study were approved by the Chungbuk National University Animal Experimentation Ethics Committee. The dried roots of *Chloranthus japonicus* Sieb. (Chloranthaceae) were extracted three times with MeOH at room temperature for 3 days (Kwon *et al.*, 2006). The MeOH extract was separated into two fractions using CHCl₃ as the non-aqueous phase. The CHCl₃ fraction was loaded on a silica gel column (Merck, 230-400 mesh) and eluted with a step-gradient of CHCl₃-MeOH (50:1). The fraction was separated using reverse phase column chromatography (YMC-Gel ODS-A, 70-230 mesh), and eluted successively with 70% MeOH. The

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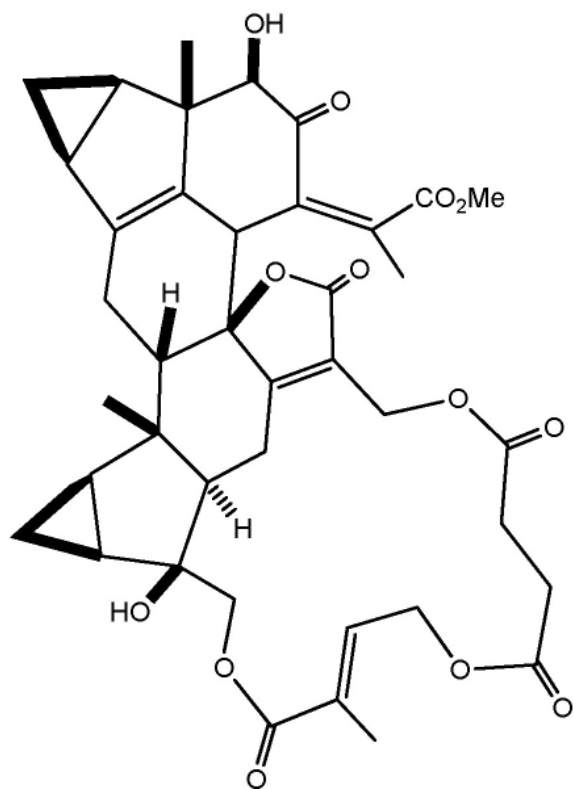


Fig. 1. Structure of SKB.

fraction was subjected to semi-preparative HPLC and gave shizukaol B (SKB)(Fig. 1).

Isolation of spleen cells

Spleen cells were obtained from specific pathogen free C57BL/6 mice (female, 6-7 weeks) and were freed of red blood cells by lysis buffer treatment (Han *et al.*, 2001). Splenic B cells were isolated by negative depletion by using biotinylated antibodies to CD4, CD8, GR-1, and CD11c (BD Pharmingen) and Dynabeads M-280 Streptavidin (Invitrogen, Carlsbad, CA, USA). Splenic T cells were isolated by negative depletion using biotinylated antibodies to B220, GR-1, and CD11c (Han *et al.*, 2005b). Purity was typically >90%. Cells were cultured in RPMI 1640 medium (Invitrogen, Carlsbad, CA, USA) supplemented with 10% fetal calf serum (Hyclone, Logan, UT, USA), 2 mM L-glutamine and 50 μM 2-mercaptoethanol (Sigma, St. Louis, MO, USA).

Generation of primary macrophages and dendritic cells

Bone marrow (BM) cells were obtained from femurs and tibias of C57BL/6 mice. After red blood cells were lysed, whole BM cells (2×10⁵ cells/ml) were cultured in 100-mm² culture dishes in 10 ml/dish of complete medium containing 10 ng/ml M-CSF to generate primary macrophages or 2 ng/ml GM-CSF (R&D Systems, Minneapolis, MN, USA) to make dendritic cells (DC). On day 3, another 10 ml of fresh complete medium containing 10 ng/ml M-CSF (or 2 ng/ml GM-CSF) was added, and half of the medium was changed on day 6. On day 8, non-adherent and loosely adherent cells were harvested by vigorous pipetting and used as primary macrophages or DCs (Kim *et al.*, 2007). Primary macrophages recovered from these cultures were generally >85% F4/80⁺, but not CD3⁺ and B220⁺ (Shi *et al.*, 2004). Cells were treated with LPS and SKB for 24

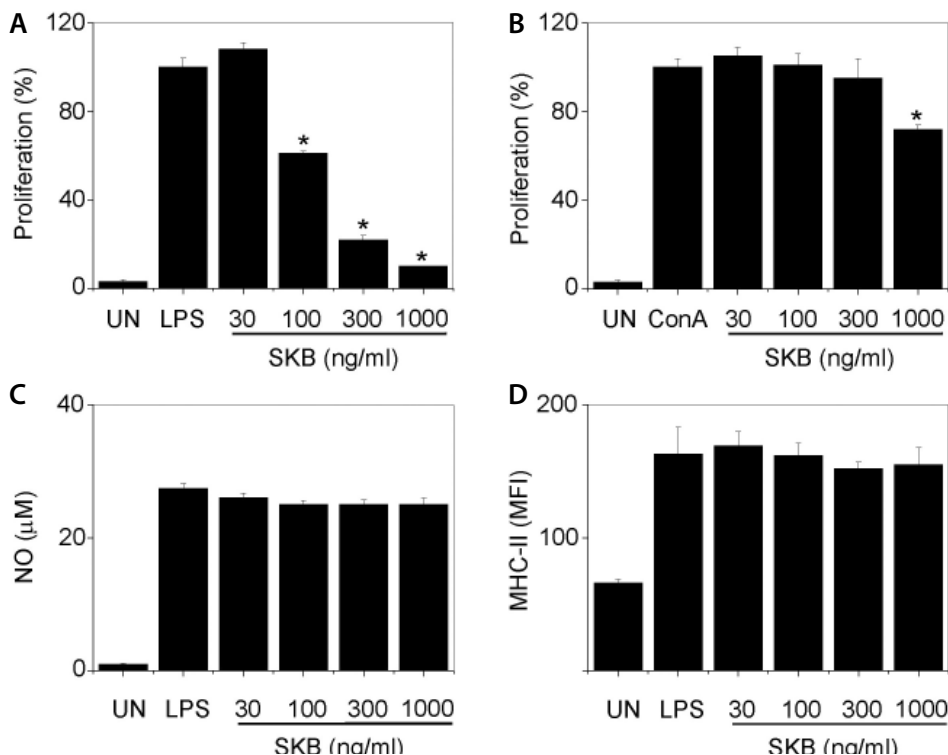


Fig. 2. Effect of SKB on the functions of immune cells. B cells were treated with 1 μg/ml of LPS and/or SKB for 72 h (A). T cells were treated with 1 μg/ml of ConA and/or SKB for 72 h (B). Cells were labeled with [³H]-thymidine for the last 18 h and harvested using an automated cell harvester. Primary macrophages were treated with 1 μg/ml of LPS and/or SKB for 24 h and NO production was measured with Griess reagent (C). Dendritic cells were treated with 1 μg/ml of LPS and/or SKB for 24 h and MHC-II expression level was measured with flow cytometer (D). Significances were determined using the ANOVA test versus LPS- or ConA-treated groups (*p<0.05).

hr. Isolated supernatants were mixed with an equal volume of Griess reagent and then incubated at room temperature for 10 min. Nitrite production was determined by measuring absorbance at 540 nm versus a NaNO_2 -derived standard curve (Han *et al.*, 2005a). Immature DCs (iDC) recovered from these cultures on day 8 were generally >85% CD11c⁺, but not CD3⁺ and B220⁺. iDCs were treated with LPS for 24 hr to induce maturation and SKB was treated at concentrations ranging from 30 to 1,000 ng/ml. Cell staining was performed using a combination of FITC-conjugated anti-MHC-II plus PE-conjugated CD11c antibodies. Cells were analyzed using a FACSCanto flow cytometer (BD Biosciences, San Jose, CA, USA), and data were analyzed using WinMdi software (Scripps, La Jolla, CA, USA). Forward and side scatter parameters were used to gate live cells.

Lymphoproliferation assay

T cells were activated with concanavalin A (Con A, 1 $\mu\text{g/ml}$) and B cells were activated with lipopolysaccharide (LPS, 1 $\mu\text{g/ml}$) (Han *et al.*, 1998). SKB was treated at concentrations ranging from 30 to 300 ng/ml. Cells were pulsed with ³H-thymidine (113 Ci/nmol, NEN, Boston, MA, USA) at a concentration of 1 $\mu\text{Ci/well}$ for the last 18 hr and harvested on day 3 using an automated cell harvester (Inotech, Dottikon, Switzerland). The amount of ³H-thymidine incorporated into cells was measured using a Wallac Microbeta scintillation counter (Wallac, Turku, Finland).

Cytotoxicity assay

Cell viability was examined by the propidium iodide (PI) nuclear staining method. Cells were stained with 1 $\mu\text{g/ml}$ of PI and analyzed with a FACSCalibur flow cytometer (BD Biosciences, San Jose, CA, USA). Cells stained with PI were con-

sidered dead cells (Kim *et al.*, 2009).

Annexin V and propidium iodide (PI) staining assay

Cells were treated with different concentration of SKB (30, 100, and 300 ng/ml) for 24 h. Apoptosis of T and B cells was determined by Annexin V/PI apoptosis detection kit (Invitrogen, Carlsbad, CA, USA). Briefly, the cell pellet was re-suspended in PBS followed by incubation with Annexin V-conjugated with FITC and PI. The fluorescence of the cells was then analyzed with a FACSCalibur flow cytometer (BD Biosciences, San Jose, CA, USA). It could discriminate intact cells (Annexin V⁻/PI⁻), early apoptotic cells (Annexin V⁺/PI⁻), and late apoptotic/necrotic cells (Annexin V⁺/PI⁺) (Nagahara *et al.*, 2002).

Cell cycle analysis

Cells were re-suspended in 75% ethanol. After centrifugation for 5 min at 1,000 rpm at 4°C, the pellet was treated with 2 mg/ml RNase A at 37°C for 20 min and stained with 50 $\mu\text{g/ml}$ propidium iodide (PI) containing 0.1% Triton X-100 and 0.02 mg/ml EDTA. Cells were then analyzed with a FACSCalibur flow cytometer (BD Biosciences, San Jose, CA, USA) (Pallardy *et al.*, 1999).

Statistics

Data represent the mean \pm STD of more than three samples and all experiments were performed more than three times. Standard deviations (STD) were calculated using the Student's *t* test and *p*-values were calculated using ANOVA software (GraphPad Software, San Diego, CA, USA).

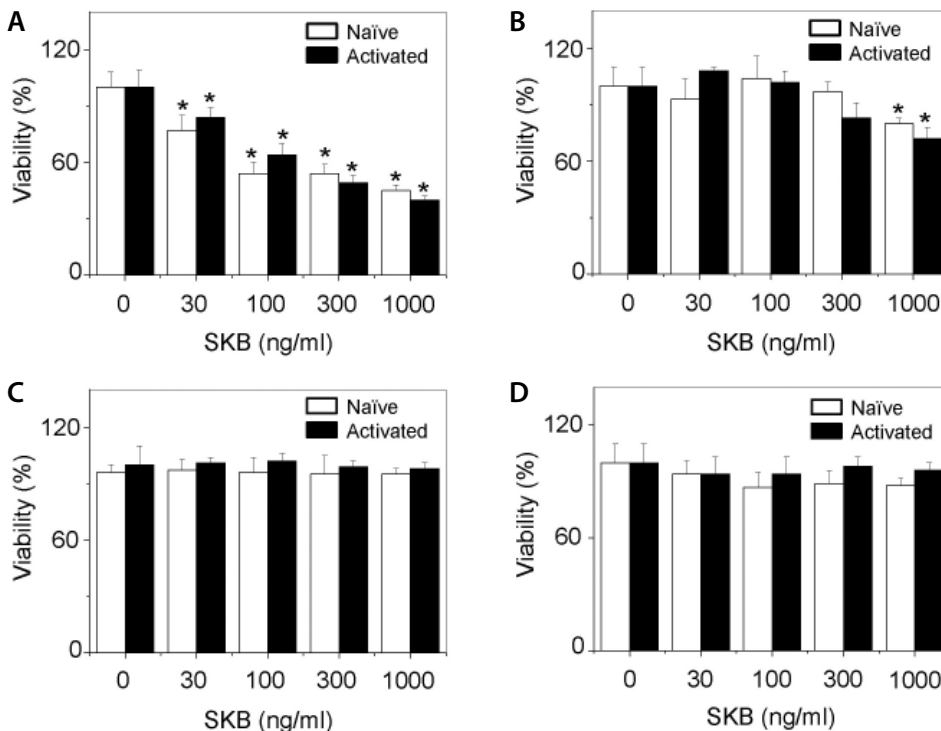


Fig. 3. Effect of SKB on the viability of immune cells. B cells (A), T cells (B), macrophages (C), and dendritic cells (D) were co-treated with LPS and SKB for 24 h to examine the effect of SKB on activated cells. Or these cells were incubated with SKB alone to examine the effect of SKB on naïve cells. Cells were stained with PI and analyzed by flow cytometric assay. PI-negative cells were considered viable cells. Significances were determined using the ANOVA test versus LPS- or ConA-treated groups (* $p < 0.05$).

RESULTS

SKB inhibits immune functions of B cells, but not T cells, macrophages, and DCs

First, we investigated the immunosuppressive activities of SKB on several immune cells, such as B cells, T cells, macrophages, and DCs, to determine cell-type selectivity of SKB. SKB inhibited LPS-induced B cell proliferation with IC_{50} of 137 ng/ml (Fig. 2A). However, SKB slightly affected ConA-induced T cell proliferation at 1,000 ng/ml (Fig. 2B). In addition, SKB did not inhibit LPS-induced macrophage NO production (Fig. 2C) and LPS-induced MHC-II expression in DCs (Fig. 2D). These results suggested that SKB strongly affected immune functions of B cells, but slightly or not those of other immune cell types.

SKB induces cytotoxicity to B cells, but not other cell types

Next, we examined whether immunosuppressive activity of SKB was due to the cytotoxicity. Purified B cells were incubated with LPS to make activated B cells or without LPS to make naïve B cells. SKB was directly added to medium from the beginning of culture. SKB induced cytotoxicity to both naïve and activated B cells with IC_{50} of 307 and 265 ng/ml, respectively (Fig. 3A). However, SKB did slightly or not affect viability of T cells (Fig. 3B), macrophages (Fig. 3C), and DCs (Fig. 3D). These data suggested that B cell-selective immuno-

suppressive activity of SKB might be due to B cell-susceptible cytotoxicity of SKB.

SKB induces apoptosis in B cells, but not T cells

In order to determine whether B cell death was due to the apoptosis-inducing activity of SKB, B cells were incubated with SKB for 24 h and stained with FITC-labeled Annexin V and propidium iodide (PI). Annexin V binds with phosphatidylserine, which becomes exposed to the outer membrane from the inner membrane during apoptosis. SKB increased annexin V binding to B cells (Fig. 4A, C), whereas annexin V binding to T cells were not changed by SKB (Fig. 4B, C).

Phosphatidylserine exposure is believed to occur in the earlier phase of apoptotic cell death. In order to discriminate early and late apoptotic cells, we stained cells with annexin V and PI from 6 to 24 h. SKB increased early apoptotic B cells (annexin⁺PI⁻) from 14% to 26% at 6 h after SKB treatment (Fig. 5A). Twenty-four hours after SKB treatment, late apoptotic B cells (annexin⁺PI⁺) were strongly increased from 22% to 47%. However, SKB did not affect annexin V binding to T cells from 6 to 24 h (Fig. 5B). Overall, these results suggested that SKB induced apoptosis in B cells, but not T cells.

SKB induces B cell cycle arrest at sub-G₁

Apoptotic cells show a reduced DNA staining by PI, lower than that of G₀/G₁ cells (sub-G₁ peak). This was due to the DNA fragmentation, which allowed small fragmented DNA to diffuse

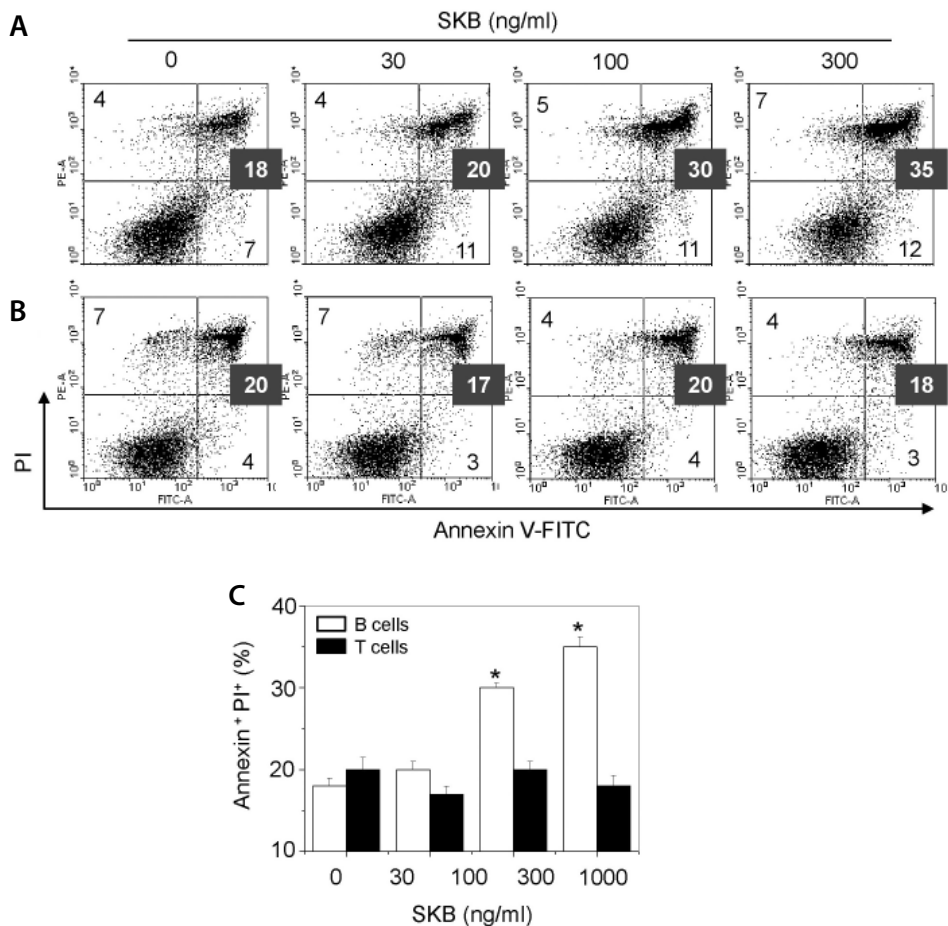


Fig. 4. Induction of B cell apoptosis by SKB. B cells (A) and T cells (B) were treated with SKB for 24 h and stained with annexin V-FITC and PI. Annexin V- and PI-positive cells were considered apoptotic cells. Representative dot plots (A and B) and mean \pm SD (C) of three-independent experiments are shown. Significances were determined using the ANOVA test versus SKB-untreated groups (* p <0.05).

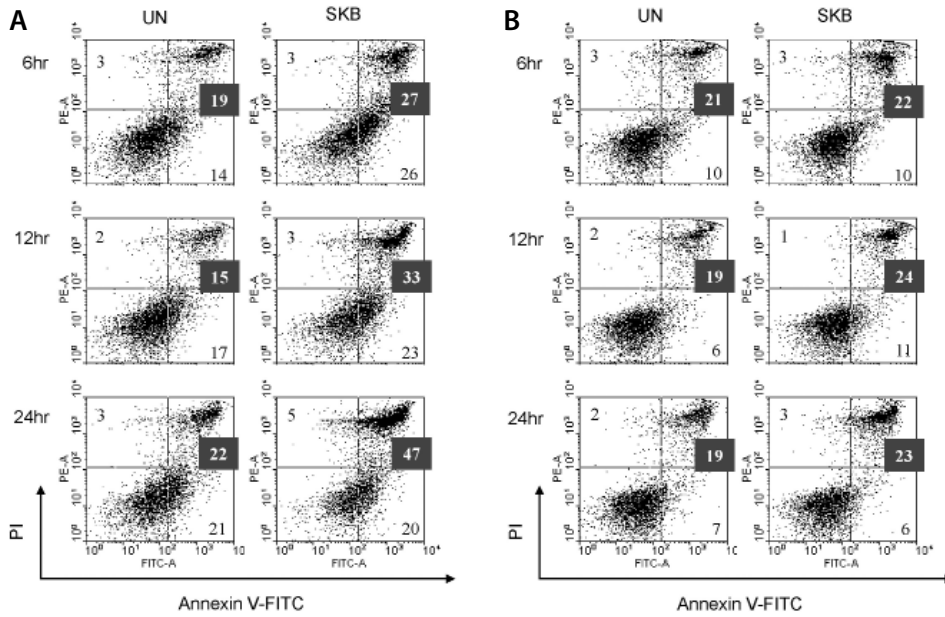


Fig. 5. Early and late apoptosis of B cells induced by SKB. B cells (A) and T cells (B) were treated with SKB from 6 h to 24 h to examine separately early and late apoptosis. After incubation for 6, 12, and 24 h, cells were stained with annexin V-FITC and PI. Annexin V-positive and PI-negative cells were considered early apoptotic cells. Annexin V-positive and PI-positive cells were considered late apoptotic cells. Representative dot plots of three-independent experiments are shown.

SKB (ng/ml)

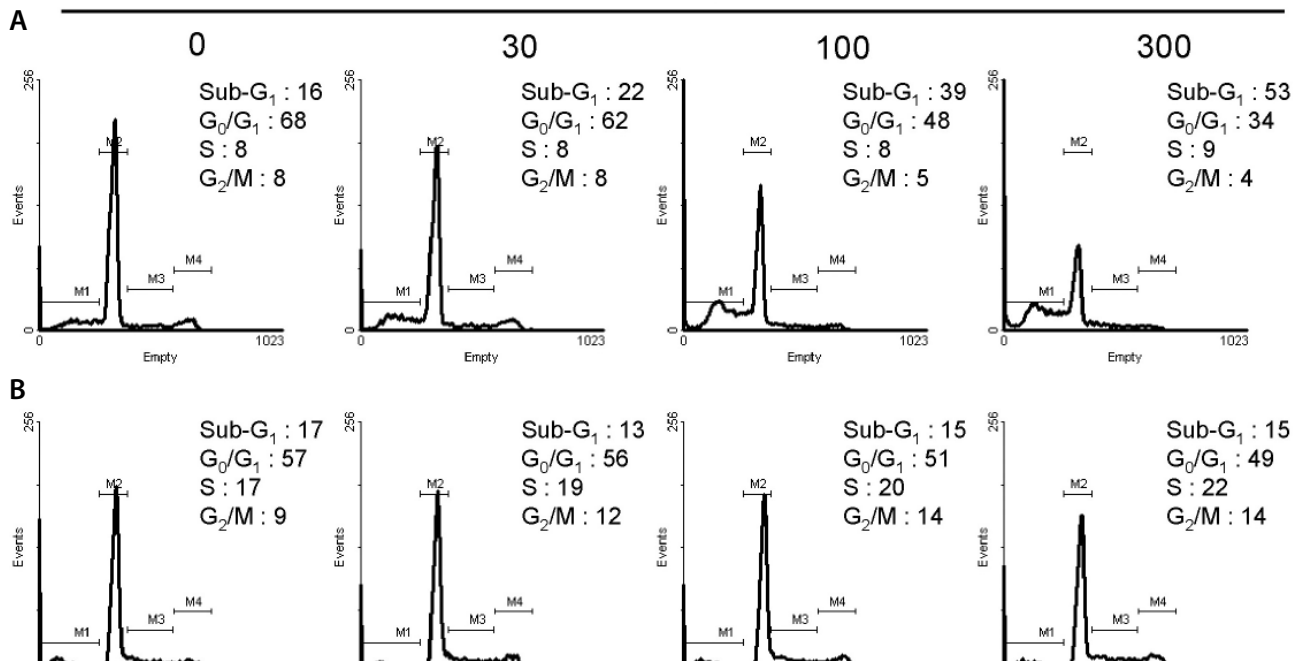


Fig. 6. Effect of SKB on cell cycles. B cells (A) and T cells (B) were treated with SKB from 24 h, re-suspended in 75% ethanol, and stained with 50 μ g/ml PI containing 0.1% Triton X-100 and 0.02 mg/ml EDTA. Cells were then analyzed with a FACSCalibur flow cytometer. Representative histograms of three-independent experiments are shown. M1: sub-G₀, M₂: G₀/G₁, M₃: S, and M₄: G₂/M phase of cell cycle.

out through cell membrane-permeabilized by ethanol treatment during experiment. As shown in Fig. 6A, SKB increased sub-G₁ peak in B cells. However, SKB slightly increase S peak in T cells without affecting sub-G₁ peak (Fig. 6B).

DISCUSSION

The immune system is now recognized as a target organ for the adverse effects of many agents including drugs and chemicals. Immunotoxic molecules may lead to two main types of effects: immunosuppression, which may result in an increased susceptibility to tumors or infectious diseases, and dysregula-

tion of the immune response, leading to hypersensitivity and autoimmunity (Pallardy *et al.*, 1999). One way to investigate the harmful effects of different chemicals is to study apoptosis in immune cells (Nagarkatti *et al.*, 2010). Assessment of apoptosis relied on morphological and biochemical modifications of the dying cells. Apoptosis is characterized by cell shrinkage, nuclear condensation, changes in cell membrane and mitochondria, DNA fragmentation, and protein degradation by caspases (Duffin *et al.*, 2010).

In this study, we investigated B cell apoptosis induced by natural compound SKB. A notable change in plasma membrane of SKB-treated B cells was observed. Preservation of plasma membrane integrity is one of the major characteristics of apoptosis (Schutz *et al.*, 2009). Loss of phospholipid asymmetry, with exposure of phosphatidylserine residues normally maintained exclusively in the inner leaflet, is an early and widespread phenomenon (Nagahara *et al.*, 2002). Annexin V, in the presence of Ca²⁺, preferentially binds to negatively charged phosphatidylserine and this property is used to quantify apoptosis by flow cytometry (Duffin *et al.*, 2010). Uptake of PI appears in late stages of apoptosis triggered *in vitro* and denotes the secondary necrosis of dying cells which are not engulfed as they would be *in vivo*. We showed here that SKB increased annexin V binding to B cells, which suggested that SKB-treated B cells lost phospholipid asymmetry and exposed phosphatidylserine to outer membrane of cells. Apoptotic cells analyzed with classic methods for cell cycle determination show a reduced DNA staining by DNA-intercalating dyes such as PI, lower than that of G0/G1 cells (sub-G1 peak) (Williams, 2004). SKB also reduced DNA staining with PI and accumulated cells at sub-G1 stage. This phenomenon is generally assumed to be due to the generation of DNA fragments that, following ethanol permeabilization of the plasma membrane, can diffuse through this altered membrane and lower total DNA content. It has also been suggested that this effect may be explained by a reduced accessibility of the dye to condensed chromatin. Whatever the exact mechanism, analysis of this sub-G1 peak by flow cytometry has the advantages of being simple and quantitative as the percentage of cells with reduced stainability in a given population can be measured accurately (Pallardy *et al.*, 1999).

In summary, we demonstrated here that SKB might induce apoptosis-mediated immunosuppression by targeting susceptible B cells, but not T cells, macrophages and dendritic cells. These data suggest that SKB might be used in anti-B cell therapy, since B cell depletion with anti-CD20 antibodies has become an accepted second line modality of therapy for patients with autoimmune diseases, such as rheumatoid arthritis, lupus, and diabetes (Basten, 2010). However, we could not clarify in this study why B cells were more susceptible to SKB than other cells, which required the continuous efforts to identify molecular mechanism and target proteins interacting with SKB in B cells.

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REFERENCES

- Basten, A. (2010) The role of B cells in transplantation and immunopathic diseases. *Immune. Netw.* **10**, 81-84.
- Duffin, R., Leitch, A. E., Fox, S., Haslett, C. and Rossi, A. G. (2010) Targeting granulocyte apoptosis: mechanisms, models, and therapies. *Immunol. Rev.* **236**, 28-40.
- Han, S. B., Kim, H. M., Kim, Y. H., Lee, C. W., Jang, E. S., Son, K. H., Kim, S. U. and Kim, Y. K. (1998) T-cell specific immunosuppression by prodigiosin isolated from *Serratia marcescens*. *Int. J. Immunopharmacol.* **20**, 1-13.
- Han, S. B., Lee, C. W., Yoon, Y. D., Lee, J. H., Kang, J. S., Lee, K. H., Yoon, W. K., Lee, K., Park, S. K. and Kim, H. M. (2005a) Prevention of arthritic inflammation using an oriental herbal combination BDx-1 isolated from *Achyranthes bidentata* and *Atractylodes japonica*. *Arch. Pharm. Res.* **28**, 902-908.
- Han, S. B., Moratz, C., Huang, N. N., Kelsall, B., Cho, H., Shi, C. S., Schwartz, O. and Kehrl, J. H. (2005b) Rgs1 and Gna2 regulate the entrance of B lymphocytes into lymph nodes and B cell motility within lymph node follicles. *Immunity* **22**, 343-354.
- Han, S. B., Park, S. H., Jeon, Y. J., Kim, Y. K., Kim, H. M. and Yang, K. H. (2001) Prodigiosin blocks T cell activation by inhibiting interleukin-2/Ralpha expression and delays progression of autoimmune diabetes and collagen-induced arthritis. *J. Pharmacol. Exp. Ther.* **299**, 415-425.
- Kim, J. Y., Kang, J. S., Kim, H. M., Kim, Y. K., Lee, H. K., Song, S., Hong, J. T., Kim, Y. and Han, S. B. (2009) Inhibition of phenotypic and functional maturation of dendritic cells by manassantin a. *J. Pharmacol. Sci.* **109**, 583-592.
- Kim, J. Y., Yoon, Y. D., Ahn, J. M., Kang, J. S., Park, S. K., Lee, K., Song, K. B., Kim, H. M. and Han, S. B. (2007) Angelan isolated from *Angelica gigas* Nakai induces dendritic cell maturation through toll-like receptor 4. *Int. Immunopharmacol.* **7**, 78-87.
- Kuang, H., Xia, Y., Yang, B., Wang, Q. and Lu, S. (2008) Sesquiterpene glucosides from *Chloranthus japonicus* sieb. *Chem. Biodivers* **5**, 1736-1742.
- Kuang, H. X., Xia, Y. G., Yang, B. Y., Wang, Q. H. and Lu, S. W. (2009) Lignan constituents from *Chloranthus japonicus* sieb. *Arch. Pharm. Res.* **32**, 329-334.
- Kwon, O. E., Lee, H. S., Lee, S. W., Bae, K., Kim, K., Hayashi, M., Rho, M. C. and Kim, Y. K. (2006) Dimeric sesquiterpenoids isolated from *Chloranthus japonicus* inhibited the expression of cell adhesion molecules. *J. Ethnopharmacol.* **104**, 270-277.
- Nagahara, Y., Ikekita, M. and Shinomiya, T. (2002) T cell selective apoptosis by a novel immunosuppressant, FTY720, is closely regulated with Bcl-2. *Br. J. Pharmacol.* **137**, 953-962.
- Nagarkatti, M., Rieder, S. A., Vakharia, D. and Nagarkatti, P. S. (2010) Evaluation of apoptosis in immunotoxicity testing. *Methods. Mol. Biol.* **598**, 241-257.
- Pallardy, M., Biola, A., Lebecq, H. and Breard, J. (1999) Assessment of apoptosis in xenobiotic-induced immunotoxicity. *Methods* **19**, 36-47.
- Schutz, C., Fischer, K., Volkl, S., Hoves, S., Halbritter, D., Mackensen, A. and Fleck, M. (2009) A new flow cytometric assay for the simultaneous analysis of antigen-specific elimination of T cells in heterogeneous T cell populations. *J. Immunol. Methods* **344**, 98-108.
- Shi, G. X., Harrison, K., Han, S. B., Moratz, C. and Kehrl, J. H. (2004) Toll-like receptor signaling alters the expression of regulator of G protein signaling proteins in dendritic cells: implications for G protein-coupled receptor signaling. *J. Immunol.* **172**, 5175-5184.
- Williams, O. (2004) Flow cytometry-based methods for apoptosis detection in lymphoid cells. *Methods Mol. Biol.* **282**, 31-42.