Notes

Synthesis and Herbicidal Activities of Enantiopure Methiozolins

Jun Ho Nam,^{†,#} Ki-Hwan Hwang,[‡] Suk-Jin Koo,[‡] Chong-Hyeak Kim,[§] Chang-Woo Cho,[#] and Young Kwan Ko^{†,*}

[†]Biomaterials Research Center, Korea Research Institute of Chemical Technology, Daejeon 305-600, Korea ^{*}E-mail: ykko@krict.re.kr

^{*}Moghu Reserch Center, Daejeon 305-333, Korea

[§]Center for Chemical Analysis, Korea Research Institute of Chemical Technology, Daejeon 305-600, Korea [#]Department of Chemistry, Kyungpook National University, Daegu 702-701, Korea

Received October 25, 2011, Accepted November 4, 2011

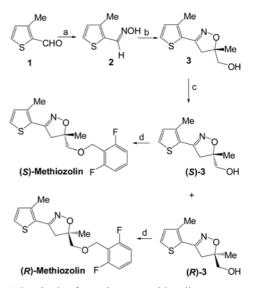
Key Words : Methiozolin, Absolute configuration determination, Optical isomer, Herbicidal activity, Grass weeds

It is well known that the worldwide demands for crops and biomass exceed their supply due to the rapid growth of world population, limitation of cultivation areas, global warming, climate change and subsequent lack of water supply. Herbicides for weeds control have played an important role in promoting productivity of crops yields and biomass.¹ Even though a number of herbicides have been developed for weeds control, more eco-friendly new herbicides having new modes of action are needed because of the environmental safety issues and occurrence of resistant weeds.^{2,3} Recently, a new isoxazoline herbicide methiozolin possessing thiophene ring for the control of grass weeds in turf grass has been commercialized.^{4,5} Methiozolin shows not only excellent herbicidal efficacy toward several grass weeds in turf grass but also favorable toxicological and environmental profiles. Methiozolin has one chiral center in isoxazoline ring. Up to now, methiozolin has been developed as a racemate.

However, it was reported that, in many cases, big differences in biological activities between enantiomers of chiral agrochemicals have been observed.⁶⁻⁸ In addition, success stories for the commercialization of chiral agrochemicals such as (S)-metolachlor⁹ and mefenoxam¹⁰ as a single isomer by Syngenta prompted us to investigate herbicidal activities of each enantiomer of methiozolin. In our previous paper,¹¹ the absolute stereochemistry of one enantiopure intermediate with posititive value of specific optical rotation, which is the key intermediate for synthesis of optically active methiozolin, was determined by single crystal X-ray diffraction analysis after prep- HPLC separation with chiral stationary phase [(R,R)-WHELK-O1 column] using [5methyl-3-(3-methylthiophen-2-yl)-4,5-dihydroisoxazol-5yl]methanol as the racemic mixture. Herein, we want to describe synthesis of enantiopure methiozolins and compare herbicidal activities between optical isomers on some grass weeds under a greenhouse condition.

Key intermediates (*S*)-**3** and (*R*)-**3** for enantiopure methiozolins could be obtained by chiral separation of a racemic [5-methyl-3-(3-methylthiophen-2-yl)-4,5-dihydroisoxazol-5yl]methanol **3** employing chiral prep-HPLC separaton. Specific optical rotation of the first eluted intermediate was +59.96 (c = 1.0, CH₂Cl₂) and that of the second eluted intermediate was -59.95 (c = 1.0, CH₂Cl₂). Absolute configuration of one enantiopure intermediate with positive specific optical rotation was determined to be *S* by single crystal Xray diffraction analysis. And the other enantiopure intermediate with negative but the same absolute value of specific optical rotation could be assigned to have the *R* configuration. Enantiopure methiozolins could be synthesized by a coupling reaction of each chiral intermediate (*S*)-**3** or (*R*)-**3** with 2,6-difluorobenzyl chloride under basic condition as shown in Scheme 1.

Specific optical rotation of one enantiopure methiozolin synthesized from (+)-[5-methyl-3-(3-methylthiophen-2-yl)-4,5-dihydroisoxazol-5-yl] methanol (*S*)-**3** was +55.0 (c = 1.0, CH₂Cl₂) and that of the other enantiopure methiozolin synthesized from (-)-[5-methyl-3-(3-methylthiophen-2-yl)-4,5-dihydroisoxazol-5-yl]methanol (*R*)-**3** was -54.5 (c = 1.0, CH₂Cl₂). Finally, absolute configuration of one enantiopure methiozolin with positive specific optical rotation was deter-



Scheme 1. Synthesis of enantiopure methiozolins. Reagents and conditions: (a) $H_2NOH \cdot HCl$, MeOH; (b) NCS, CH_2Cl_2 , 2-methylpropen-1-ol, NaHCO₃. (c) chiral prep-HPLC separation. (d) 2,6-difluorobenzyl chloride, NaOH, THF

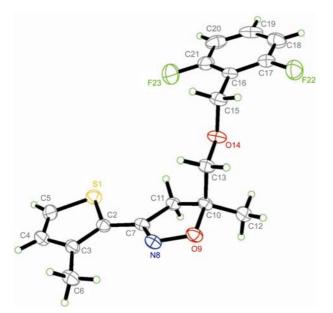


Figure 1. ORTEP drawing of (S)-methiozolin.

mined to be the S configuration by single X-ray diffraction analysis (Figure 1).¹² In addition, the other enantiopure methiozolin with negative but the same absolute value of specific optical rotation could be assigned to be the R configuration.

Biological Evaluation Pre-emergence herbicidal activities of racemic methiozolin and its enantiopure isomers toward troublesome grass weeds such as *Digitaria sanguinalis* and *Poa annua* in turf grass were tested under a greenhouse condition. As shown in Figure 2, herbicidal activities of racemic methiozolin and its optical isomers towards *Digitaria sanguinalis* and *Poa annua* were in order of (*S*)-isomer, racemic methiozolin and (*R*)-isomer. Surprisingly, (*R*)-isomer exhibited no herbicidal activities toward grass weeds tested.

In conclusion, each enantiomer of methiozolin was synthesized from the key intermediates (*S*)-**3** and (*R*)-**3**, respectively, which were prepared by prep-HPLC separation with chiral stationary phase. The absolute configuration of (*S*)methiozolin was determined by single crystal X-ray diffraction analysis. Interestingly, only (*S*)-isomer of methiozolin showed the herbicidal activities toward grass weeds.

Experimental

¹H NMR (500 MHz) and ¹³C NMR (75 MHz) spectra were recorded in CDCl₃. The signal positions are reported in parts per million relative to TMS (δ scale) used as an internal standard. IR spectra are reported in cm⁻¹. Mass spectra, elemental analysis and single crystal X-ray diffraction analysis were carried out at Korea Research Institute of Chemical Technology, Daejeon, Korea. All reagents were purchased from commercial sources and used without further treatment. Flash chromatography was carried out using silica gel 60 (230-400 mesh ASTM) using various solvent mixtures.

Chiral separation was performed on a YOUNGLIN SDV30plus series HPLC system (YOUNGLIN, Seoul, South Korea). The injection volume was 20 μ L. The following HPLC columns purchased from (*R*,*R*)WHELK-O1 of REGIS Chemical Industries (USA) were used: The Whelk-O 1 Chiral Stationary Phase is based on 1-(3,5-Dinitro benzamido)-1,2,3,4-tetra-hydrophenanthrene.

3-Methyl-2-thiophenecarboxaldoxime (2). Freshly distilled 3-methyl- 2-thiophenecarboxaldehyde (126.0 g, 1.0 mol) was dissolved in methanol/water solution (200 mL/150 mL), and hydroxylamine hydrochloride (76.5 g, 1.1 mol) was added during 15 min at 10-15 °C with good stirring. A solution of NaOH (44.0 g, 1.1 mol) in water (50 mL) was added dropwise to the reaction mixture, and it was stirred for an additional 15 min at room temperature. The reaction mixture was concentrated by rotary evaporation to precipitate white crystalline solids. The precipitates were filtered and dried under vacuum to afford 3-methyl-2-thiophenecarboxaldoxime (139.0 g, 98.5%) mp 111.0-112.0 °C; ¹H NMR (500 MHz, CDCl₃) δ 2.44 (s, 3H), 6.95 (d, *J* = 5.1 Hz, 1H), 7.49 (d, *J* = 5.1 Hz, 1H), 7.81 (s, 1H); C NMR (75

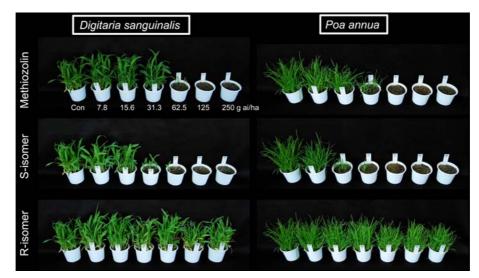


Figure 2. Herbicidal activities of racemic methiozolin and enantiopure isomers toward Digitaria sanguinalis and Poa annua.

Notes

MHz, CDCl₃) δ 14.5 124.9 129.2 130.4 139.6 141.1.

[5-Methyl-3-(3-methyl-thiophen-2-yl)-4,5-dihydroisoxazol-5-yl]methanol (3). To a solution of 3-methyl-2thiophenecarboxaldoxime (2) (139.0 g, 0.98 mol) in CH₂Cl₂ (1000 mL) was added N-chloro succinimde (152.2 g, 1.14 mol) by potions for 45 min at 10-40 °C. After the reaction mixture was stirred for an additional 3 h, it was poured into methylene chloride (1000 mL). The solution was washed with water (200 mL \times 2) and brine (300 mL \times 2), dried over MgSO₄, and filtered. The filtrate was placed in a 2-L round bottomed flask, and 2-methylpropen-1-ol (86.6 g, 1.2 mol) and sodium bicarbonate (108.0 g, 1.2 mol) were slowly added to the mixture for 30 min at 5 °C. The reaction mixture was stirred for 12 hr at room temperature, washed with brine (200 mL \times 2). The organic layer was dried over MgSO₄, and concentrated by rotary evaporation to give a pale yellow solid. A total of 50 mL of ethyl acetate and 250 mL of hexane were added to the solid with good shaking, and then, the suspension was cooled in an ice bath. The solid precipitates were filtered and washed with hexane to give [5-Methyl-3-(3-methyl-thiophen-2-yl-4,5-dihydroisoxazol-5yl]methanol (3) as a racemic mixture (186.0 g, 90%). mp 94.9-97.7 °C; ¹H NMR (500 MHz, CDCl₃) δ 1.44 (s, 3H), 2.24 (s, 3H), 3.06 (d, J = 16.35 Hz, 1H), 3.53 (d, J = 16.4 Hz, 1H), 3.59 (d, J = 11.9 Hz, 1H), 3.76 (d, J = 12.0 Hz, 1H), 6.93 (d, *J* = 5.0 Hz, 1H), 7.29 (d, *J* = 4.2 Hz, 1H); C NMR (75 MHz, CDCl₃) δ 16.5 22.6 44.6 67.2 86.9 126.1 126.3 132.0 139.0 153.1.

Chiral Separation of [5-Methyl-3-(3-methyl-thiophen-2-yl)4,5-dihydroisoxazol-5-yl]methanol (3). Racemic [5-Methyl-3-(3-methyl-thiophen-2-yl)-4,5-dihydroisoxazol-5yl]methanol was separated by preparative chiral HPLC under the condition of (*R*,*R*)WHELK-O1 column, hexane/ isopropylalcohol 7.5/2.5 v/v mobile phase, flow-rate 4.0 mL/min, $k \neq 254$ nm, 100 µL injection volume to give the first eluted isomer (*S*)-3 and the second eluted isomer (*R*)-3. $[\alpha]_{D}^{20}$ of the first eluted isomer (*S*)-3 = +59.96 (*c* = 1.0, CH₂Cl₂); $[\alpha]_{D}^{20}$ of the second eluted isomer (*R*)-3 = -59.95 (*c* = 1.0, CH₂Cl₂). The first eluted isomer was recrystallized in ethyl acetate to provide the single crystal for X-ray crystallograph.

(S)-5-(2,6-Difluorobenzyloxymethyl)-5-methyl-3-(3-methylthiophen-2-yl)-4,5-dihydroisoxazole. To a solution of (S)-[5-methyl-3-(3-methyl-thiophen-2-yl)-4,5-dihydroisoxazol-5-yl]methanol (S)-3 (105.5 g, 0.5 mol) in THF (400 mL) was added NaOH (66.6 g, 1.1 mol) during 15 min at 40 °C, and the mixture was stirred for 15 min at that temperature. A solution of 2,6-difluorobenzyl chloride (97.6 g, 0.6 mol) in THF (60 mL) was added to the reaction mixture during 15 min, and the reaction temperature increased to 60-70 °C. The reaction mixture was stirred for an additional 1 h at that temperature and poured into ice water (500 mL). The solution was extracted with ethyl acetate (500 mL \times 2), and the organic layer was washed with 1N HCl solution (200 mL \times 2) and brine (200 mL \times 2). The organic layer was dried over MgSO₄ and concentrated to give crude product, which was purified by silica gel column chromatography (300 g,

Bull. Korean Chem. Soc. 2012, Vol. 33, No. 1 299

1:20 ethyl acetate/hexane) to provide pure product as oil. The oily product was recrystallized in hexane to give (*S*)-5-(2,6-difluorobenzyl)oxymethyl-5-methyl-3-(3-methylthiophen-2-yl)-1,2-isoxazoline as a white crystalline solid (100.0 g, 60%). mp 45-46 °C. ¹H NMR (500 MHz, CDCl₃) δ 1.47 (s, 3H), 2.46 (s, 3H), 3.02 (d, *J* = 16.45 Hz, 1H), 3.45 (d, *J* = 16.50 Hz, 1H), 3.54 (d, *J* = 9.9 Hz, 1H), 3.60 (d, *J* = 9.85 Hz, 1H), 4.72 (s, 2H), 6.93-6.89 (m, 3H), 7.31-7.26 (m, 2H); MS *m/z* (relative intensity): 338 (17.8), 337 (42.5), 180 (87.3), 137 (100), 127 (53.6); IR (KBr, cm⁻¹): 3104.0, 2973.2, 2928.2, 2882.7, 1626.8, 1471.0, 1234.7, 1097.5, 1056.6, 786.0; Anal. Calcd for C₁₇H₁₇F₂NO₂S : C, 60.52; H, 5.08; N, 4.15; S, 9.50. Found: C, 60.53; H, 5.20; N, 4.14; S, 9.43; UV λ_{max} : 279.5 cm⁻¹; $[\alpha]_D^{20} = +55.50$ (*c* = 1.0, CH₂Cl₂).

(R)-5-(2,6-Difluorobenzyloxymethyl)-5-methyl-3-(3-methylthiophen-2-yl)-4,5-dihydroisoxazole. To a solution of (R)-[5-methyl-3-(3-methyl-thiophen-2-yl)-4,5-dihydroisoxazol-5-yl]methanol (R)-3 (105.5 g, 0.5 mol) in THF (400 mL) was added NaOH (66.6 g, 1.1 mol) during 15 min at 40 °C, and the mixture was stirred for 15 min at that temperature. A solution of 2,6-difluorobenzyl chloride (97.6 g, 0.6 mol) in THF (60 mL) was added to the reaction mixture during 15 min, and the reaction temperature increased to 60-70 °C. The reaction mixture was stirred for an additional 1 h at that temperature and poured into ice water (500 mL). The solution was extracted with ethyl acetate (500 mL \times 2), and the organic layer was washed with 1 N HCl solution (200 mL \times 2) and brine (200 mL \times 2). The organic layer was dried over MgSO₄ and concentrated to give crude product, which was purified by silica gel column chromatography (300 g, 1:20 ethyl acetate/hexane) to provide pure product as oil. The oily product was recrystallized in hexane to give (R)-5-(2,6-difluorobenzyl)oxymethyl-5-methyl-3-(3-methylthiophen-2-yl)-1,2-isoxazoline as a white crystalline solid (100.0 g, 60%). mp 45-46 °C. ¹H NMR (500 MHz, CDCl₃) δ 1.47 (s, 3H), 2.46 (s, 3H), 3.02 (d, J = 16.45 Hz, 1H), 3.45 (d, J = 16.50 Hz, 1H), 3.54 (d, J = 9.9 Hz, 1H), 3.60 (d, J =9.85 Hz, 1H), 4.72 (s, 2H), 6.93-6.89 (m, 3H), 7.31-7.26 (m, 2H); MS m/z (relative intensity): 338 (17.8), 337 (42.5), 180 (87.3), 137 (100), 127 (53.6); IR (KBr, cm⁻¹): 3104.0, 2973.2, 2928.2, 2882.7, 1626.8, 1471.0, 1234.7, 1097.5, 1056.6, 786.0; Anal. Calcd for C₁₇H₁₇F₂NO₂S : C, 60.52; H, 5.08; N, 4.15; S, 9.50. Found: C, 60.53; H, 5.20; N, 4.14; S, 9.43; UV λ_{max} : 279.5 cm⁻¹; $[\alpha]_D^{20} = -54.50$ (*c* = 1.0, CH₂Cl₂).

Acknowledgments. This work was supported by the R&D Program of MKE/KEIT [10035240, Development of new herbicides for resistant weeds with mutated gene].

References

- Pimentel, D.; Pimentel, M. Food, Energy and Society, Arnold, E., London, CRC press: 1979.
- Boger, P.; Wakabayashi, K.; Hirai, K. Herbicide Classes in Development (mode of acton, targets, genetic engineering, chemistry), Berlin, Heidelberg: Springer-Verlag: 2002.
- 3. Bryant, R.; Bite, M. *Ag Chem New Compound Review*, United Kingdom, Agranov: 2011.

- 300 Bull. Korean Chem. Soc. 2012, Vol. 33, No. 1
- Hwang, I. Y.; Kim, H. R.; Jeon, D. J.; Hong, K. S.; Song, J. H.: Cho, K. Y. J. Agric. Food Chem. 2005, 53, 8639.
- Koo, S.-J.; Hwang, K.-H.; Jeon, M.-S.; Kim, S.-H.; Lim, J.-S.; Lee, D.-K.; Chung, K.-H.; Ko, Y. K.; Ryu, J. W.; Koo, D. W.; Woo, J. C. Kor. J. Weed Science 2010, 30, 323.
- 6. Williams, A. Pestic. Sci. 1996, 46, 3.
- Kurihara, N.; Miyamoto, J.; Paulson, G. D.; Zeeh, B.; Skidmore, M. W.; Hollingworth, R. M.; Kuiper, H. A. *Pure and Appl. Chem.* 1997, *69*, 2007.
- 8. Williams, A. Phytoparasitica 2000, 28, 293.
- 9. Moser, H.; Rihs, G.; Sauter, H. P.; Bohner, B. *Pesticide Chemistry: Human Welfare and the Environment*, Vol. 1, Pergamon, New York: 1982.
- 10. Sukul, P.; Spiteller, M. Reviews of Environmental Contamination and Toxicology 2000, 164, 1.
- 11. Ko, Y. K.; Ryu, J. W.; Koo, D. W.; Woo, J. C.; Kim, C.-H. Acta Cryst. 2011, E67, o1040.
- 12. Crystal data of (*S*)-methiozolin compound: solvent of crystal growth (hexane-diethyl ether); empirical formular C₁₇H₁₇NO₂F₂S, Fw = 337.38, crystal dimensions 0.40 mm × 0.19 mm × 0.08 mm, monoclinic, space group *P*2₁, *a* = 10.449(1) Å, *b* = 5.1485(7) Å, *c* = 30.622(5) Å, $\alpha = 90^{\circ}$, $\beta = 96.936(9)^{\circ}$, $\gamma = 90^{\circ}$, *V* = 1635.3(4) Å³, Z = 4, $D_{calcd} = 1.370$ g/cm³, $F_{000} = 704$, MoK α ($\lambda = 0.71073$ Å), $R_1 = 0.0495$, $wR_2 = 0.1047$ ($I > 2\sigma(I)$). The X-ray data has been deposited in CCDC with number 848038.