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# Dietary Supplementation of Magnesium Sulfate during Late Gestation and Lactation Affects the Milk Composition and Immunoglobulin Levels in Sows

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ABSTRACT: This experiment was conducted to investigate the effects of dietary supplementation of magnesium sulfate (MgSO<sub>4</sub>) during late gestation and lactation on sow and litter performance, fecal moisture, blood biochemistry parameters, immunoglobulin levels and milk composition in sows. Forty-eight sows (Yorkshire×Landrace, 4th to 5th parity) were randomly allocated to 1 of 4 dietary treatments supplemented with 0, 200, 400, or 600 mg/kg MgSO<sub>4</sub>(n = 12). The experiment started on day 90 of gestation and continued through day 21 of lactation. Blood samples were collected on day 107 of gestation, day 0 (farrowing) and 21 (weaning) of lactation for the analyses of the blood biochemistry parameters and immunoglobulin levels. The colostrum and milk samples were obtained on day 0 and 14 of lactation, respectively. Fecal samples were collected from the sows on day 107 of gestation as well as day 7 and 20 of lactation to determine fecal moisture content. The results showed that the survival percentage of piglets and the litter weight at weaning were decreased linearly (p<0.05) and other parameters of the sow or litter performance were not influenced (p>0.05) by MgSO<sub>4</sub> supplementation. The fecal moisture content of the sows were increased (p<0.05) linearly as dietary MgSO<sub>4</sub> increased on day 7 and 20 of lactation. Supplementation with MgSO<sub>4</sub> increased the plasma magnesium (Mg) level linearly (p<0.05) and had a trend to increase total protein level (p>0.05 and p<0.10). However, an increase in the dietary MgSO<sub>4</sub> level resulted in a linear decrease in the colostrum fat content (p<0.05). Dietary MgSO<sub>4</sub> supplementation enhanced the immunoglobulin G (IgG) level (linear, p<0.05) in plasma on day of farrowing and immunoglobulin A (IgA) level in colostrum (quadratic, p<0.05) and milk (linear, p<0.05) of the sows. These results indicated that supplementation with MgSO<sub>4</sub> during late gestation and lactation may have the potential to prevent sow constipation, but may also result in some negative effects. (Key Words: Magnesium Sulfate, Sow, Performance, Fecal Moisture, Immunoglobulin, Milk Composition)

### INTRODUCTION

Magnesium (Mg) is a macro-mineral that participates in the metabolism of carbohydrates, nucleic acids, proteins and lipids and contributes to the action of many enzymes as well as cellular transport (Trawńska et al., 2013). Magnesium ion is required for the synthesis of ATP and proper functioning of immune system and it mitigates states of supernormal excitability and enhances resistance to stress (Maguire and Cowan, 2002; Wolf and Cittadini, 2003;

soybean diets fed to monogastric animals contain adequate amounts of Mg, and deficiencies of Mg are rare (Lee and Britton, 1980). However, studies have demonstrated that supplementing Mg to swine diets alleviated the effects of stress and improved meat quality (D'souza et al., 1998; Apple et al., 2005). It has been also confirmed that MgSO<sub>4</sub> had the capability of reducing heat stress-induced oxidative damage in broilers (Yang et al., 2012). Moreover, MgSO<sub>4</sub> can be used as a laxative in a clinical practice (Ikarashi et al., 2012). It has been demonstrated that dietary MgSO<sub>4</sub> supplementation could reduce the number of constipated fininshing pigs and furthermore, the addition of 400 mg/kg MgSO<sub>4</sub> in the finishing pig diet could effectively prevent

and control the occurrence of constipation (Tang, 2000).

Guerrera et al., 2009; Siener et al., 2011). Common corn-

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Most previous studies have focused on the effects of dietary  $MgSO_4$  intake on broilers, cows and growing-finishing pigs, however, fewer experimental studies are available on the effect of  $MgSO_4$  supplementation on sows.

With the increase of the intensiveness of sow breeding reproductive barriers occurred, such constipation (Liu and Luo, 2010). As sows approach farrowing, mild constipation is a common state because the intestine is less active (Tabeling et al., 2003). Feed intake and intestinal activity undergo major changes around farrowing when sows are often constipated (Oliviero et al., 2009). Constipation is associated with udder infections in sows during early lactation (Hermansson et al., 1978; Smith, 1985) and may cause pain or discomfort, as well as affect the duration of farrowing (Cowart, 2007). In commercial production, sow perinatal feed can be top-dressed with MgSO<sub>4</sub> at 1.0% for several days as a treatment for constipation (Shi, 2010). Furthermore, MgSO<sub>4</sub> can be included in the fininshing pig diets at 400 mg/kg as a preventative of constipation (Tang, 2000). It has not yet been investigated whether MgSO<sub>4</sub> supplementation of sows influences performance, milk composition and other parameters. Therefore, the aims of this study were to determine whether supplementation of MgSO<sub>4</sub> to the sow diets at 200, 400 and 600 mg/kg during late gestation and lactation could prevent the sow constipation and to investigate the effects of MgSO<sub>4</sub> on sow and litter performance, blood biochemistry parameters, immunoglobulin levels and milk composition in sows.

### **MATERIALS AND METHODS**

The Institutional Animal Care and Use Committee of Northeast Agricultural University (Harbin, China) approved all of the experimental protocols.

# Animal, treatment and management

Forty-eight sows (Yorkshire×Landrance, 4th to 5th parity) were artificially inseminated with semen from a pool of Duroc boars. The sows were randomly allocated to 1 of 4 dietary treatments and fed with the basal diet or the basal diet supplemented with 200, 400 or 600 mg/kg MgSO<sub>4</sub>. The experiment started on day 90 of gestation and continued through day 21 of lactation. The ingredients and chemical composition of the basal diets for gestating and lactating sows are shown in Table 1. All of the diets were formulated to meet or exceed the sow's nutrient requirements, as recommended by the National Research Council (1998). The magnesium sulfate monohydrate (MgSO<sub>4</sub>·H<sub>2</sub>O, ≥99%; MgSO<sub>4</sub>,  $\geq$ 86%; Mg,  $\geq$ 17.21%) used for the experiments was purchased from Harbin Veterinary Medicine Wholesale Market (Harbin, China). During late gestation, twelve sows of each treatment were randomly housed in 3 pens of 4.0

**Table 1.** Composition of experimental diets (as-fed basis)

	Gestation	Lactation
Ingredients (%)		
Maize	65.07	56.25
Wheat bran	18.00	10.00
Full-fat soybean, 35.5% CP	-	5.00
Soybean meal, 43% CP	13.30	21.20
Fish meal	-	2.00
Soy oil	-	2.00
Dicalcium phosphate	1.05	1.05
Limestone	1.08	1.00
Salt	0.40	0.40
Choline chloride (50%)	0.10	0.10
Vitamin and mineral premix <sup>1</sup>	1.00	1.00
Chemical composition		
Metabolizable energy <sup>2</sup> (MJ/kg)	11.92	12.74
Crude protein <sup>3</sup> (%)	13.97	18.88
Crude fibre <sup>3</sup> (%)	3.52	3.30
Calcium <sup>3</sup> (%)	0.71	0.79
Total phosphorus <sup>3</sup> (%)	0.56	0.62
Magnesium <sup>3</sup> (%)	0.20	0.19

CP, crude protein.

m×3.0 m and all gestating sows were in the same room. The sows were fed approximately 3.0 kg/d of the gestation diet from day 90 of gestation until the day of farrowing. On day 107 of gestation, the sows were moved to the farrowing room and housed in individual farrowing stalls (1.5 m×2.1 m). On day 0 of lactation, the sows were not fed the lactation diet. The feed allowance of the sows was raised to 2.0 kg on day 1. Thereafter, this amount was increased daily by 1.0 kg until ad libitum feeding. The lactation diet was provided three times daily. The sows and piglets had free access to water from nipple waterer throughout the experiment. The farrowing stalls had a piglet creep area provided with a heat lamp. The average daily feed intake (ADFI) of the sows during lactation was recorded. The sow weight was recorded at entry into the farrowing room and at the time of weaning. The backfat thickness was measured at P2 (6 cm from the midline at the head of the last rib) with an ultrasonic device (Renco Lean Meater Type 7, Minneapolis, MN, USA) within 24 h after farrowing and on day 21 of lactation. The weaning-to-estrus interval (WEI) of each sow was recorded from weaning to 7 days postweaning. At day 3 postpartum, the piglets received an iron injection, and the males were castrated at day 5 postpartum.

Supplied per kg of diet: 20 mg of Cu as  $CuSO_4 \cdot SH_2O$ ; 120 mg of Zn as  $ZnSO_4$ ; 0.3 mg of Se as  $Na_2 SeO_3 \cdot H_2O$ ; 40.07 mg of Mn as  $MnSO_4 \cdot H_2O$ ; 100.50 mg of Fe as  $FeSO_4 \cdot H_2O$ ; 0.3 mg of I as  $Ca(IO_3)_2$ ; vitamin A, 13,500 IU; vitamin  $D_3$ , 3,000 IU; vitamin E, 15 IU; vitamin  $K_3$ , 3 mg; thiamine, 1.8 mg; riboflavin, 6 mg; pyridoxine, 0.3 mg; vitamin  $B_{12}$ , 0.024 mg; niacin, 24 mg; pantothenic acid,15 mg; folate, 0.9 mg; biotin, 0.03 mg.

<sup>&</sup>lt;sup>2</sup> Calculated values.

<sup>&</sup>lt;sup>3</sup> Analyzed values.

Commercial creep diets were offered to the piglets at day 7 postpartum. The piglets were not cross-fostered, and they were counted and weighed at birth and weaning.

# Sample collection

Blood, colostrum, milk, and fecal samples were collected from eight sows per experimental treatment. Blood samples (10 mL) were collected from the ear vein of the sows into heparin tubes on day 107 of gestation as well as day 0 and 21 of lactation for the analyses of blood biochemistry parameters and immunoglobulin levels. The blood samples were centrifuged at 1,000×g for 10 min. Then, all of the plasma samples were stored at –20 °C until needed for analysis.

Approximately 30 mL of colostrum was manually collected 1 h after the birth of the first piglet from all functional glands. Piglets were removed from the sow until completion of farrowing to prevent suckling and the initiation of gut closure. To facilitate milk sampling on day 14 of lactation, piglets were removed from the sow 1 h before and milk ejection was induced after intramuscular administration of 2 mL of oxytocin. The colostrum and milk samples were immediately frozen at -20°C until the analysis.

Fresh fecal samples were collected from the sows on day 107 of gestation as well as day 7 and 20 of lactation. The fecal samples were stored at  $-20\,^{\circ}$ C until analysis. Every morning before the daily cleaning, we observed the faeces of each sow from five days before to five days after farrowing. The sow with dry and pellet-shaped faeces or no faecal production for three or four consecutive days was defined as severe constipation (Oliviero et al., 2009).

# Chemical analyses

Samples of the gestation and lactation diets were analyzed for crude protein (CP), crude fibre (CF), calcium (Ca) and phosphorus (P) (Association of Official Analytical Chemists [AOAC], 2006). Dietary Mg was determined by atomic absorption spectrophotometer (PE AA 800, Waltham Mass, USA). The milk composition was analyzed using a fully automatic milk analyzer (Milko ScanTM FT+ Analyser, Foss, Hillerød, Denmark). The biochemistry parameters including plasma Mg, total protein and triglycerides, were measured by a fully automated biochemical analyzer (Unicel DxC 800 Synchron Clinical System, Beckman Coulter, Fullerton, CA, USA). The immunoglobulin concentrations in the plasma, colostrum and milk were assessed by immune turbidimetry using immunoglobulin-specific kits (Sanwei Biological Engineering Company Limited, Shandong, China). The milk samples were ultracentrifuged at 3,000×g at 4°C for 30 min to remove the free fat, and the supernatant was separated and kept frozen until the analyses of the immunoglobulin levels. The supernatant samples were analyzed in duplicate at dilution levels of 1:2 for colostrum immunoglobulin G (IgG), 1:1 for colostrum immunoglobulin A (IgA) and 1:1 for colostrum immunoglobulin M (IgM), respectively. The determination of the immunoglobulins was performed using an ultraviolet spectrophotometer (UV-2401PC, Shimadzu Co., Kyoto, Japan). The optical density (OD) values of the standards and samples at 700 nm (IgG) or 340 nm (IgA and IgM) were measured. The immunoglobulin concentrations of the samples (g/L) were calculated by comparing the OD value of the samples to the standard curve. The fecal samples (10 g) were thawed at room temperature and weighed on a dry plate and then weighed again after drying for 24 h at 65°C in a constant temperature drying box to a constant weight. The water content of the feces was calculated based on the difference between the wet and dry fecal weights.

#### Statistical analysis

Differences between treatments were examined using a one-way ANOVA (SPSS 17.0, SPSS Inc., Chicago, IL, USA) with individual sows and their litters being the experimental unit. The linear and quadratic effects of MgSO<sub>4</sub> level were assessed using orthogonal polynomials. A p-value of less than 0.05 was considered statistically significant and tendency for all analysis was p>0.05 and p<0.10.

#### **RESULTS**

#### Sow and litter performance

The effects of MgSO<sub>4</sub> on sow and litter performance are shown in Table 2. There were no differences (p>0.05) in total number of piglets born, the number of piglets born alive, the number of stillborn piglets or the number of piglets per litter at weaning among the treatments. The litter birth body weight (BW), the average piglet birth BW, the average birth weight of all live-born piglets, the average piglet weaning BW and average daily gain (ADG) were not affected by dietary treatments (p>0.05). The survival percentage and the litter weight at weaning were declined linearly (p<0.05) with increasing level of MgSO<sub>4</sub>. The ADFI of the sows during lactation, the sow BW loss from day 107 of gestation to weaning and the change in backfat during lactation were also not affected (p>0.05) by adding MgSO<sub>4</sub>. However, the WEI was decreased linearly (p = 0.05).

## Fecal moisture content

As is shown in Table 3, fecal moisture content was increased (linear effect, p<0.05) on day 7 of lactation and increased (linear and quadratic, p<0.05) on day 20 of lactation as the level of  $MgSO_4$  increased. However, there

Table 2. Effects of dietary supplementation of magnesium sulfate during late gestation and lactation on sow and litter performance

Item	Magnesium sulfate (mg/kg)					p-value	
item -	0	200	400	600	- SEM	Linear	Quadratic
Total piglets born	11.92	11.83	12.00	12.17	0.24	0.178	0.232
Piglets born alive	10.83	10.75	11.25	11.08	0.24	0.297	0.703
Stillborn piglets	1.08	1.08	0.75	1.08	0.18	0.742	0.775
Piglets per litter at weaning	9.42	9.17	9.25	8.83	0.19	0.120	0.432
Survival rate at weaning (%)	87.11	85.23	83.25	82.29	1.73	0.010	0.067
Litter birth weight (kg)	17.63	17.95	17.40	17.25	0.34	0.284	0.538
Average piglet birth weight (kg)	1.49	1.54	1.44	1.44	0.03	0.326	0.674
Average birth weight of all live born piglets (kg)	1.54	1.56	1.45	1.49	0.03	0.324	0.728
Litter weight at weaning (kg)	50.46	49.67	48.82	46.30	1.31	0.046	0.115
Average piglet weaning weight (kg)	5.37	5.42	5.28	5.23	0.09	0.158	0.421
Day 0 to 21 ADG (g/d)	182.8	178.3	180.7	177.1	4.0	0.252	0.656
Lactation ADFI (kg)	4.66	4.62	4.33	4.38	0.08	0.124	0.457
Sow weight at day 107 of gestation (kg)	261.2	267.8	258.4	266.3	6.1	0.826	0.981
Sow weight at weaning day 21 (kg)	221.5	228.0	217.8	220.8	5.6	0.630	0.899
Sow weight loss from day 107 of gestation to weaning (kg)	39.71	39.83	40.67	45.50	1.71	0.145	0.154
Sow backfat at post-farrowing day 0 (mm)		22.08	21.58	21.50	0.70	0.996	0.669
Sow backfat at weaning day 21 (mm)	17.67	18.42	17.58	17.58	0.64	0.648	0.770
Sow lactation backfat change (mm)	3.67	3.67	4.00	3.92	0.18	0.183	0.560
WEI by 7 day post-weaning <sup>2</sup> (d)	4.67	4.38	4.33	4.20	0.13	0.050	0.208

SEM, standard error of mean; ADG, average daily gain; ADFI, average daily feed intake; WEI, weaning-to-estrus interval.

**Table 3.** Effects of dietary supplementation of magnesium sulfate during late gestation and lactation on the fecal moisture content of sows<sup>1</sup>

Fecal moisture (%)	Magnesium sulfate (mg/kg)				SEM	p-value	
	0	200	400	600	SEM	Linear	Quadratic
Day 107 of gestation	64.80	64.12	65.66	66.34	0.46	0.182	0.409
Day 7 of lactation	65.12	65.94	66.53	68.06	0.39	0.021	0.122
Day 20 of lactation	65.42	65.88	67.02	68.41	0.40	0.021	0.042

SEM, standard error of mean. <sup>1</sup> Values are the means of 8 replicates.

was no difference (p>0.05) in the fecal moisture content among the treatments on day 107 of gestation. Moreover, the number of the sows with severe constipation in the 0, 200, 400, 600 mg/kg  $MgSO_4$  treatments was 3, 2, 2, and 1 respectively.

#### Colostrum and milk composition

The composition of the sow colostrum and milk are shown in Table 4. No differences (p>0.05) were observed in the total solid, fat or lactose levels of the milk among the treatments. However, linear trend in milk protein level (p = 0.095) was observed by increasing dietary MgSO<sub>4</sub> level. There were no differences (p>0.05) in the lactose, protein or total solid levels of colostrum among the treatments. Fat content of the sow colostrum was decreased linearly (p<0.01) with increasing level of MgSO<sub>4</sub>.

## **Blood biochemistry parameters**

The blood biochemistry parameters are shown in Table

5. Plasma magnesium concentrations were increased on day 107 of gestation (linear and quadratic, p<0.05), day of farrowing and weaning (linear, p<0.05). Plasma total protein tended to be linearly increased on day 107 of gestation (p = 0.068) and day of weaning (p = 0.057). Triglycerides level was declined with increasing level of MgSO<sub>4</sub> (linear, p<0.05) on day of farrowing.

# Plasma immunoglobulin levels of sows

The IgG, IgA, and IgM levels in the plasma of sows are presented in Table 5. There were no differences (p>0.05) in the IgA (day of farrowing and weaning) or IgM concentrations (day 107 of gestation, day of farrowing and weaning) among the treatments. Dietary supplementation of MgSO<sub>4</sub> increased IgA level quadratically (p<0.05) on day 107 of gestation. The IgG level was increased linearly (p<0.05) as the level of MgSO<sub>4</sub> increased on the day of farrowing. There were no linear and quadratic relationships (p>0.05) in plasma IgG level on day 107 of gestation and

<sup>&</sup>lt;sup>1</sup> Values are the means of 12 replicates. <sup>2</sup> 9, 8, 9, and 10 for treatments 1, 2, 3, and 4, respectively.

**Table 4.** Effects of dietary supplementation of magnesium sulfate during late gestation and lactation on the colostrum and milk composition in sows<sup>1</sup>

Item -		Magnesium sulfate (mg/kg)					p-value	
	0	200	400	600	- SEM	Linear	Quadratic	
Colostrum (%)								
Fat	4.99	4.55	4.28	3.82	0.14	0.005	0.095	
Lactose	3.66	3.57	3.68	3.78	0.06	0.296	0.315	
Protein	16.48	18.51	19.79	17.31	0.46	0.663	0.269	
Total solids	26.68	27.82	28.55	26.43	0.45	0.997	0.317	
Milk (%)								
Fat	7.68	7.18	7.56	7.42	0.19	0.760	0.841	
Lactose	6.18	6.25	5.89	5.92	0.09	0.190	0.583	
Protein	4.86	4.80	5.25	5.37	0.15	0.095	0.384	
Total solids	20.79	20.27	20.81	20.65	0.22	0.938	0.908	

SEM, standard error of mean. <sup>1</sup> Values are the means of 8 replicates.

day of weaning among the treatments.

## Immunoglobulin in colostrum and milk

The IgG, IgA, and IgM levels in the colostrum and milk are shown in Table 6. The IgM concentrations in the colostrum and milk were not affected by adding MgSO<sub>4</sub> (p>0.05). No linear and quadratic relationships (p>0.05) were observed in IgG levels of colostrum and milk among the treatments. A quadratic relationship (p<0.05) was observed in IgA level in the colostrum of sows with

increasing level of MgSO $_4$ . The IgA concentration in the milk was increased linearly (p<0.05) with increasing level of MgSO $_4$ .

### DISCUSSION

There are few studies on the effects of MgSO<sub>4</sub> supplementation on sow and litter performance during late gestation and lactation. In our study, no significant differences were observed in the pregnancy parameters

**Table 5.** Effects of dietary supplementation of magnesium sulfate during late gestation and lactation on blood biochemistry parameters and plasma immunoglobulin levels in sows<sup>1</sup>

Item		Magnesium s	CEM	p-value			
	0	200	400	600	- SEM	Linear	Quadratic
Day 107 of gestation							
Magnesium (mmol/L)	0.90	0.92	0.95	0.99	0.02	0.004	0.036
Total protein (g/L)	80.98	80.85	86.10	87.66	0.91	0.068	0.334
Triglycerides (mmol/L)	0.54	0.59	0.58	0.58	0.02	0.318	0.491
IgG (g/L)	5.13	5.29	7.12	5.88	0.21	0.417	0.678
IgA (g/L)	0.25	0.26	0.26	0.25	< 0.01	0.842	< 0.001
IgM (g/L)	0.30	0.36	0.35	0.36	0.01	0.201	0.388
Day of farrowing							
Magnesium (mmol/L)	0.78	0.79	0.83	0.88	0.01	0.024	0.103
Total protein (g/L)	74.58	76.98	83.78	79.08	1.06	0.330	0.525
Triglycerides (mmol/L)	0.24	0.23	0.20	0.19	0.01	0.026	0.224
IgG (g/L)	3.33	3.50	4.60	5.14	0.18	0.032	0.221
IgA (g/L)	0.21	0.21	0.25	0.24	< 0.01	0.223	0.628
IgM (g/L)	0.29	0.28	0.30	0.28	< 0.01	1.000	0.963
Day of weaning							
Magnesium (mmol/L)	0.88	0.92	0.97	0.97	0.01	0.038	0.182
Total protein (g/L)	91.75	94.98	101.60	101.48	1.23	0.057	0.267
Triglycerides (mmol/L)	0.22	0.23	0.30	0.22	0.02	0.775	0.674
IgG (g/L)	5.53	5.66	8.12	7.06	0.23	0.261	0.613
IgA (g/L)	0.20	0.25	0.23	0.24	< 0.01	0.393	0.584
IgM (g/L)	0.35	0.41	0.37	0.34	0.01	0.729	0.385

SEM, standard error of mean; IgG, immunoglobulin G; IgA, immunoglobulin A; IgM, immunoglobulin M.

<sup>1</sup> Values are the means of 8 replicates.

IgA

IgM

p-value Magnesium sulfate (mg/kg) Item SEM 0 200 400 600 Linear Quadratic Colostrum (g/L) IgG 45.03 47.76 52.38 50.88 0.98 0.127 0.315 IgA 7.16 8.60 8.79 8.01 0.23 0.517 0.049 IgM 3.22 3.66 3.92 3.49 0.12 0.530 0.224 Milk (g/L) IgG 0.73 0.79 0.01 0.195 0.74 0.76 0.122

3.69

1.56

3.87

1.51

0.04

0.02

0.014

0.812

0.108

0.689

Table 6. Effects of dietary supplementation of magnesium sulfate during late gestation and lactation on the immunoglobulin levels in the colostrum and milk of sows1

1.53 SEM, standard error of mean; IgG, immunoglobulin G; IgA, immunoglobulin A; IgM, immunoglobulin M.

3.61

(litter size, litter weight) among the treatments, which may suggest that MgSO<sub>4</sub> had no effects on the growth and development of fetuses during late gestation. Rattanatayarom et al. (2000) reported that pregnancy parameters were not adversely affected in supplemented with magnesium-L-aspartate hydrochloride daily at a dose of 13 mg Mg per kg BW starting 0 to 4 days before mating. The present data indicated that the average piglet weaning weight and ADG during the suckling period were not affected by adding MgSO<sub>4</sub>, but the survival percentage and the litter weight at weaning were declined linearly with increasing level of MgSO<sub>4</sub>. On the contrary, Trawńska et al. (2013) reported that the feeding of pregnant sows with the addition of MgCl<sub>2</sub>·6H<sub>2</sub>0 (1 g/100 kg BW/d), which constituted 120 mg of pure Mg, decreased the mortality rate of the newborn piglets and increased the survival rate of piglets until 21 days of life and the litters BW.

3.50

1.53

An adequate feed intake is important for sow performance and reproduction (Koketsu et al., 1997). Feed intake was connected with BW loss, litter performance and backfat depth (Le Cozler et al., 1999; Eissen et al., 2003). In our study, supplementation of MgSO<sub>4</sub> had no significant effects on the sow lactation ADFI, BW loss or backfat loss. Different with our results, Stockdale (2004) reported that adding 105 to 155 g MgSO<sub>4</sub> to the diet of cows during the last 3 weeks of gestation had a negative effect on intake before calving. Supplementation of MgSO<sub>4</sub> at doses of 0.255, 1.02, and 2.04 g/kg diet linearly reduced BW gain and feed intake in broiler chickens (Van der Hoeven-Hangoor et al., 2013). Stockdale (2004) interpretated that supplementation of MgSO<sub>4</sub> would seriously affect the palatability of the feed and reduce the feed intake. The possible reasons for the difference of these results are related to the animal species and MgSO<sub>4</sub> level.

Constipation is a common state around farrowing (Tabeling et al., 2003; Oliviero et al., 2009). In clinical practice, osmotic laxatives are prescribed as the first-line agents for patients with severe constipation (Ikarashi et al., 2012). It is believed that osmotic laxatives such as magnesium oxide (MgO) and MgSO<sub>4</sub> induce diarrhea by increasing the osmotic pressure in the intestinal tract (Izzo et al., 1996). The laxative effect produced by MgSO<sub>4</sub> is not simply a result of changes in osmotic pressure but is also associated with the increased expression of a water channel protein, aquaporin-3 (AQP<sub>3</sub>), in the mucosal epithelial cells of the colon (Ikarashi et al., 2011a,b). The present study showed that supplementation of MgSO<sub>4</sub> increased the fecal moisture content linearly during lactation. However, there was no difference in the fecal moisture content on day 107 of gestation among the treatments. This might be due to the higher concentration of the wheat bran in the diet of gestation sows compared to diet of lactation sows. Wheat bran contains adequate dietary fiber and can promote bowel movements to relieve constipation due to the waterretaining properties of insoluble fiber (Shen, 1998). Additionally, water consumption also has a great impact on fecal moisture content. Moreover, the number of the sows with severe constipation in the MgSO<sub>4</sub> groups was less compared with the control group. In summary, the addition of 600 mg/kg MgSO<sub>4</sub> to the diet during late gestation and lactation may have the potential to prevent sow constipation to some extent. Similar finding on finishing pig constipation with the addition of MgSO<sub>4</sub> to diets was reported by Tang (2000). According to Van der Hoeven-Hangoor et al. (2013), adding Mg (MgSO<sub>4</sub>, MgO, and magnesium chloride (MgCl<sub>2</sub>), each at 3 levels (0.255, 1.02, and 2.04 g/kg diet)) to the diet of broilers linearly increased the excreta moisture content, following the pattern MgCl<sub>2</sub>>MgSO<sub>4</sub> = MgO, which has confirmed that Mg can prevent reabsorption of water from the digesta, and as a consequence, more moisture will be present in the excreta.

In the present study, the plasma Mg concentrations were significantly increased with the supplementation of MgSO<sub>4</sub>. This was in agreement with previous observations that supplementation with dietary Mg increased the serum Mg

<sup>&</sup>lt;sup>1</sup> Values are the means of 8 replicates.

concentration in pigs (Rattanatayarom et al., 2000; Lahucky et al., 2004). The change of plasma total protein suggested that the supplementation of MgSO<sub>4</sub> in sow diets could promote protein synthesis and strengthen protein deposition. The increase of plasma total protein was most likely caused by the fact that Mg participates in the metabolism of proteins due to its role in the action of many enzymes (Wolf and Cittadini, 2003).

The growth of suckling pigs depends on the availability and composition of colostrum and milk and the efficiency of their conversion into BW gain (Ariza-Nieto et al., 2011). A strong association exists between preweaning growth and the sow milk nutrient output (Noblet and Etienne, 1989). Reductions in the colostrum fat percentage were found in sows supplemented with MgSO<sub>4</sub> during late gestation. In addition, the MgSO<sub>4</sub> intake resulted in a higher milk protein content. However, Stockdale (2004) reported that MgSO<sub>4</sub> had no effect on either milk yield or milk composition in cows. The fat in milk is a major source of energy, accounting for 55% to 60% of the total energy (Le Dividich and Seve, 2001). The high fat content of colostrum is essential to the survival and health of piglets (Jackson et al., 1995). Triglycerides are a primary component of the fat in mammary milk, and they mainly provide fatty acids (Boyd and Kensinger, 1998). The decrease in the colostrum fat content in this experiment may be linked to the lower plasma triglycerides in sows supplemented with MgSO<sub>4</sub> on day of farrowing. According to Haenni et al. (1998), plasma Mg could significantly reduce plasma concentration of the fat. Moreover, supplementation of Mg in the diet improved lipid metabolism (Apple et al., 2000). The fat in milk is one of the important factors affecting piglet weight gain (Miller et al., 1994). This may be the reason for the decrease of survival percentage and the litter weight at weaning.

Several studies relating Mg to various aspects of the immune response have been published (Larvor, 1980; Galland, 1988; McCoy and Kenney, 1992). Magnesium is essential for the normal functioning of many components of the innate and adaptive immune defenses (Zimowska et al., 2002). The harmful effects of experimentally induced severe Mg deficiency on the immune system are well known, and experimental Mg deficiency has been shown to affect the humoral immune system (Alcock and Shils, 1974; Kubena et al., 1989; Windhauser et al., 1991). Regarding the effects of dietary MgSO<sub>4</sub> on immunoglobulin levels, dietary supplementation of MgSO4 during gestation and lactation resulted in the linear increase of plasma IgG concentrations in sows on the day of farrowing, but no linear and quadratic relationships were found in the colostrum and milk. We also found that the treaments supplemented with MgSO<sub>4</sub> resulted in the increase of IgA concentrations in the colostrum and milk. The data suggested that a higher MgSO<sub>4</sub> intake induced elevated IgG

production in the blood of the sows, which could improve the host defense against invading pathogens. According to Galland (1988), Mg is involved in the synthesis of immunoglobulins and the activation of complement, and it impacts the function of phagocytes and regulates the maturation and function of T lymphocytes. Maternal immunity is very important to maintain the health status of pigs and their production levels (Le Dividich, 2006). The survival of neonatal pigs is totally dependent on maternal immunity via the colostrum and milk due to the epitheliochorial placenta (Kim, 1975). The colostrum provides newborn piglets with maternal serum antibodies, the majority of which are IgG (Bourne and Curtis, 1973). Colostral IgG is translocated from the blood to the mammary gland (Salmon et al., 2009). This process nearly stops just after parturition, resulting in a sharp decrease in the colostral IgG concentration when the colostrum becomes milk (Klobasa et al., 1987). As lactation proceeds, IgG concentrations decrease and IgA becomes the major immunoglobulin in sow milk (Gaskins, Immunoglobulin G is predominantly involved in the secondary antibody response, whereas IgA plays a critical role in mucosal immunity (Ariza-Nieto et al., 2011). In our study, the MgSO<sub>4</sub> treatments improved the milk IgA levels. In pigs, nearly 100% of milk IgA originated from IgA synthesized locally into the mammary gland (Bourne and Curtis, 1973). These results showed that MgSO<sub>4</sub> could stimulate the mammary tissue to synthesize IgA. The milk antibody (especially IgA) cannot be absorbed into the blood, but it protects against local pathogens, commensal bacteria and food antigens in the topical digestive tract (Brandtzaeg, 2010). Our study demonstrated that supplementation with MgSO<sub>4</sub> enhanced the IgA level in the colostrum and milk, which may support the better protection of piglets. However, the increases of immunoglobulin levels in the colostrum and milk did not result in gains by the piglets while suckling.

## CONCLUSION

The results suggest that supplementation with 200, 400, or 600 mg/kg of MgSO<sub>4</sub> decreased the survival percentage and the litter weight at weaning linearly and did not affect other parameters of the sow or litter performance. The addition of MgSO<sub>4</sub> to the diet during late gestation and lactation increased the fecal moisture content linearly during lactation. Supplementation with MgSO<sub>4</sub> linearly increased the concentrations of plasma Mg and total protein. Supplementation with MgSO<sub>4</sub> increased the plasma IgG level of sows on the day of farrowing and IgA level in the milk. Additionally, an increase in the dietary MgSO<sub>4</sub> level during late gestation and lactation resulted in a linear decrease of colostrum fat and an increase of milk protein.

We conclude that supplementation with MgSO<sub>4</sub> during late gestation and lactation may have the potential to prevent sow constipation, but result in some negative effects.

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#### REFERENCES

- Alcock, N. W. and M. E. Shils. 1974. Serum immunoglobulin G in the magnesium-depleted rat. Proc. Soc. Exp. Biol. Med. 145: 855-858.
- AOAC. 2006. Official Methods of Analysis. 18th edn. Association of Official Analytical Chemists, Arlington, VA, USA.
- Apple, J. K., C. V. Maxwell, B. DeRodas, H. B. Watson, and Z. B. Johnson. 2000. Effect of magnesium mica on performance and carcass quality of growing-finishing swine. J. Anim. Sci. 78: 2135-2143.
- Apple, J. K., E. B. Kegley, C. V. Maxwell, L. K. Rakes, D. Galloway, and T. J. Wistuba. 2005. Effects of dietary magnesium and short-duration transportation on stress response, postmortem muscle metabolism, and meat quality of finishing swine. J. Anim. Sci. 83:1633-1645.
- Ariza-Nieto, C., M. Bandrick, S. K. Baidoo, L. Anil, T. W. Molitor, and M. R. Hathaway. 2011. Effect of dietary supplementation of oregano essential oils to sows on colostrum and milk composition, growth pattern and immune status of suckling pigs. J. Anim. Sci. 89:1079-1089.
- Bourne, F. J. and J. Curtis. 1973. The transfer of immunoglobulins IgG, IgA and IgM from serum to colostrum and milk in the sow. Immunology. 24:157-162.
- Boyd, R. D. and R. S. Kensinger. 1998. Metabolic precursors for milk synthesis. In: Lactating Sow, 1st Ed. (Eds. M. W. A. Verstegen, P. J. Morgan, and J. W. Schrama). Wageningen Press, Wageningen, Netherlands. pp.71-95.
- Brandtzaeg, P. 2010. The mucosal immune system and its integration with the mammary glands. J. Pediatr. 156:S8-S15.
- Cowart, R. P. 2007. Parturition and dystocia in swine. In: Large Animal Theriogenology (Eds. R. S. Youngquist and W. R. Threlfall). Saunders, St. Louis, MI, USA. pp. 778-784.
- D'souza, D. N., R. D. Warner, B. J. Leury, and F. R. Dunshea. 1998. The effect of dietary magnesium aspartate supplementation on pork quality. J. Anim. Sci. 76:104-109.
- Eissen, J. J., E. J. Apel doorn, E. Kanis, M. W. A. Verstegen, and K. H. de Greef. 2003. The importance of a high feed intake during lactation of primiparous sows nursing large litters. J. Anim. Sci. 81:594-603.
- Galland, L. 1988. Magnesium and immune function: An overview. Magnesium 7:290-299.
- Gaskins, H. R. 1998. Immunological development and mucosal defence in the pig intestine. Pages 81-10 In: Progress in Pig Science (Eds. J. Wiseman, M. A. Varley, and J. P. Chadwick).

- Nottingham University Press, Nottingham, UK. pp. 81-10.
- Guerrera, M. P., S. L. Volpe, and J. J. Mao. 2009. Therapeutic uses of magnesium. Am. Fam. Physician. 80:157-162.
- Haenni, A., M. Ohrvall, and H. Lithell. 1998. Atherogenic lipid fractions are related to ionized magnesium status. Am. J. Clin. Nutr. 67:202-207.
- Hermansson, I., S. Einarsson, K. Larsson, and L. Backstrom. 1978. On the agalactia postpartum in the sow. A clinical study. Nord. Vet. Med. 30: 465-473.
- Ikarashi, N., A. Mimura, R. Kon, T. Iizasa, M. Omodaka, C. Nagoya, M. Ishii, T. Toda, W. Ochiai, and K. Sugiyama. 2012. The concomitant use of an osmotic laxative, magnesium sulphate, and a stimulant laxative, bisacodyl, does not enhance the laxative effect. Eur. J. Pharm. Sci. 45:73-78.
- Ikarashi, N., T. Mochiduki, A. Takasaki, T. Ushiki, K. Baba, M. Ishii, T. Kudo, K. Ito, T. Toda, W. Ochiai, and K. Sugiyama. 2011a. A mechanism by which the osmotic laxative magnesium sulphate increases the intestinal aquaporin 3 expression in HT-29 cells. Life Sci. 88: 194-200.
- Ikarashi, N., T. Ushiki, T. Mochizuki, T. Toda, T. Kudo, K. Baba, M. Ishii, K. Ito, W. Ochiai, and K. Sugiyama. 2011b. Effects of magnesium sulphate administration on aquaporin 3 in rat gastrointestinal tract. Biol. Pharm. Bull. 34:238-242.
- Izzo, A. A., T. S. Gaginella, and F. Capasso. 1996. The osmotic and intrinsic mechanisms of the pharmacological laxative action of oral high doses of magnesium sulphate. Importance of the release of digestive polypeptides and nitric oxide. Magnes. Res. 9:133-138.
- Jackson, J. R., W. L. Hurley, R. A. Easter, A. H. Jensen, and J. Odle. 1995. Effects of induced or delayed parturition and supplemental dietary fat on colostrum and milk composition in sows. J. Anim. Sci. 73:1906-1913.
- Kim, Y. B. 1975. Developmental immunity in the piglet. Birth Defects Orig. Artic. Ser. 11:549-557.
- Klobasa, F., E. Werhahn, and J. E. Butler. 1987. Composition of sow milk during lactation. J. Anim. Sci. 64:1458-1466.
- Koketsu, Y., G. D. Dial, J. E. Pettigrew, and V. L. King. 1997. Influence of feed intake during individual weeks of lactation on reproductive performance of sows on commercial farms. Livest. Prod. Sci. 49:217-225.
- Kubena, K. S., D. T. Cohill, and D. N. McMurray. 1989. Effect of varying levels of magnesium during gestation and lactation on humoral immune response and tissue minerals in rats. Ann. Nutr. Metab. 33:7-14.
- Lahucky, R., K. Nuernberg, U. Kuechenmeister, I. Bahelka, J. Mojto, and G. Nuernberg. 2004. The effect of dietary magnesium oxide supplementation on fatty acid composition, antioxidative capacity and meat quality of heterozygous and normal malignant hyperthermia pigs. Arch. Tierz., Dummerstorf. 47:183-191.
- Larvor, P. 1980. Magnesium, humoral immunity, and allergy. In: Magnesium in Health and Disease Proceedings of the 2nd International Symposium on Magnesium (Eds. M. Cantin and M. S. Seelig). SP Medical and Scientific Books, New York, USA. pp. 201-224.
- Le Cozler, Y., E. Ringmar-Cederberg, L. Rydhmer, N. Lundeheim, J. Y. Dourmad, and M. Neil. 1999. Effect of feedings level during rearing and mating strategy on performance of Swedish Yorkshire sows: 2. Reproductive performance, food intake,

- backfat changes and culling rate during the first two parities. Anim. Sci. 68:365-377.
- Le Dividich, J. 2006. The issue of colostrum in piglet survival: energy and immunity. In: Biotechnology in the Feed Industry: Proceedings of Alltech's 22 Annual Symposium (Eds. T. P. Lyons and K. A. Jacques). Nottingham Press University, Nottingham, UK. pp. 89-102.
- Le Dividich, J. and B. Seve. 2001. Energy requirement of the young pig. In: The Weaner Pig (Eds. M. A. Varley and J. Wiseman). CABI Publishing, Wallingford, UK. pp. 17-44.
- Lee, S. and W. M. Britton. 1980. Magnesium toxicity: Effect on phosphorus utilization by broiler chicks. Poult. Sci. 59:1989-
- Liu, Z. K. and B. R. Luo. 2010. Investigation of sow reproductive condition in part scale pig farms in seven provinces of China. Swine. Prod. 5:65-68.
- Maguire, M. E. and J. A. Cowan. 2002. Magnesium chemistry and biochemistry. Biometals. 15:203-210.
- McCoy, H. and M. A. Kenney. 1992. Magnesium and immune function: recent findings. Magnes. Res. 5:281-293.
- Miller, M. B., T. G. Hartsock, B. Erez, L. Douglass, and B. Alston-Mills. 1994. Effect of dietary calcium concentrations during gestation and lactation in the sow on milk composition and litter growth. J. Anim. Sci. 72:1315-1319.
- National Research Council. 1998. Nutrient Requirements of Swine. 10th Ed. National Academy Press, Washington, DC, USA.
- Noblet, J. and M. Etienne. 1989. Estimation of sow milk nutrient output. J. Anim. Sci. 67:3352-3359.
- Oliviero, C., T. Kokkonen, M. Heinonen, S. Sankari, and O. Peltoniemi. 2009. Feeding sows with high fibre diet around farrowing and early lactation: impact on intestinal activity, energy balance related parameters and litter performance. Res. Vet. Sci. 86:314-319.
- Rattanatayarom, W., K. Korteerakul, H. G. Classen, and L. Spatling. 2000. Effects of magnesium-L-aspartate hydrochloride supplements in pregnant sows. Magnes. B. 22:39-44.
- Salmon, H., M. Berri, V. Gerdts, and F. Meurens. 2009. Humoral and cellular factors of maternal immunity in swine. Dev. Comp. Immunol. 33:384-393.
- Shen, Y. M. 1998. Characteristics and application of dietary fiber. Cereals and Oils 4:2-4.

- Shi, M. K. 2010. Reasons and prevention measures for sow constipation in intensive farms. Chinese J. Anim. Quar. 27:59-60.
- Siener, R., A. Jahnen, and A. Hesse. 2011. Bioavailability of magnesium from different pharmaceutical formulations. Urol. Res. 39:123-127.
- Smith, B. B. 1985. Pathogenesis and therapeutic management of lactation failure in periparturient sows. Compend. Contin. Educ. Prac. Vet. 7: S523-S532.
- Stockdale, C. R. 2004. Effects of feeding magnesium sulfate to dry pregnant dairy cows with different body condition scores on intake in late gestation, periparturient blood calcium concentrations and production in early lactation. Anim. Prod. Sci. 44:539-546.
- Tabeling, R., S. Schwier, and J. Kamphues. 2003. Effects of different feeding and housing conditions on dry matter content and consistency of faeces in sows. J. Anim. Physiol. Anim. Nutr. 87:116-121.
- Tang, D. H. 2000. Effect of feeding magnesium sulfate to fattening pigs on constipation. Lives. Poult. Ind. 6:24-25.
- Trawńska, B., A. Polonis, J. Lechowski, L. Tymczyna, R. Borowski, and K. Gizińska. 2013. Effect of the addition of magnesium salt to a feed mixture on intestinal microflora, health, and production of sows. Bull. Vet. Inst. Pulawy 57:69-72.
- Van der Hoeven-Hangoor, E., I. B. Van de Linde, N. D. Paton, M. W. A. Verstegen, and W. H. Hendriks. 2013. Effect of different magnesium sources on digesta and excreta moisture content and production performance in broiler chickens. Poult. Sci. 92: 382-391.
- Windhauser, M. M., L. C. Kappel, and M. Hegsted. 1991. The humoral immune response in marginally and severely magnesium deficient rats. J. Nutr. Biochem. 2:541-546.
- Wolf, F. I. and A. Cittadini. 2003. Chemistry and biochemistry of magnesium. Mol. Aspects Med. 24:3-9.
- Yang, Y., M. Y. Gao, W. Nie, J. M. Yuan, B. K. Zhang, Z. Wang, and Z. L. Wu. 2012. Dietary magnesium sulfate supplementation protects heat stress-induced oxidative damage by restoring the activities of anti-oxidative enzymes in broilers. Biol. Trace Elem. Res. 146:53-58.
- Zimowska, W., J. P. Girardeau, J. Kuryszko, D. Bayle, Y. Rayssiguier, and A. Mazur. 2002. Morphological and immune response alterations in the intestinal mucosa of the mouse after short periods on a low-magnesium diet. Br. J. Nutr. 88:515-522.