

Review

Potential of some traditionally used edible plants for prevention and cure of diabesity associated comorbidities

Vikas Kumar^{1,*}, Ajit Kumar Thakur², Suruchi Verma¹, Vaishali Yadav¹, Shyam Sunder Chatterjee³

¹Neuropharmacology Research Laboratory, Department of Pharmaceutics, Indian Institute of Technology (Banaras Hindu University), Varanasi-221 005, Uttar Pradesh, India ²School of Pharmacy, IEC University, Baddi-174103, Solan, Himachal Pradesh ³Stettiner Straße 1, 76139, Karlsruhe, Germany

ABSTRACT

Medicinal uses of edible and other plants for prevention and cure of obesity and overweight associated metabolic and mental health problems have since long been known to scholars and practitioners of Ayurvedic and other traditionally known system of medicine. Modernized versions of numerous edible plant derived formulations mentioned in ancient Ayurvedic texts are at present some of the most popular, or best selling, herbal remedies in India and numerous other countries suffering from double burden of diseases caused by malnutrition and obesity. Preclinical and clinical information now available on edible plants and their bioactive constituents justify traditionally known medicinal uses of products derived from them for prevention and cure of obesity associated type-2 diabetes, psychopathologies and other health problems. Such information now available on a few edible Ayurvedic plants and their formulations and suggesting that their stress response regulating effects are involved in their broad spectrums of bioactivity profiles are summarized in this communication. Implications of recent physiological and pharmacological observations made with numerous phytochemicals isolated from edible plants for better understanding of traditionally known medicinal uses of herbal remedies are also pointed out.

Keywords Ayurveda, diabesity, neurological disorders, Brassica juncea, Emblica officinalis, Piper longum, Curcuma

INTRODUCTION

Classical Ayurvedic texts are the oldest known ones pointing out that overweight, obesity, and diabetes are closely related lifestyle disorders, and that appropriate food choices and eating habits are necessary not only for proper maintenance of physical and mental health, but also for obtaining optimal therapeutic benefits from medicines and other health care measures (Rastogi, 2014; Sharma and Chandola, 2011a; Sharma and Chandola, 2011b). Modern nutritional and other researchers have now also well recognized that diverse phytochemicals commonly consumed with everyday meals have beneficial effects against overweight or malnutrition associated metabolic disorders encountered in patients suffering from, or at risk to diabetes, hyperlipidemia, and numerous others chronic diseases (Dembinska-Kiec et al., 2008; Farooqui, 2013; Gonzalez-Castejon and Rodriguez-Casado, 2011; Manach et al., 2009). Consequently, regular consumption of fruits, vegetables and other plant derived products are now highly recommended for reducing the risks of almost all such life threatening and silently progressing diseases (Anonymous, 2003; Agrawal et al., 2014). Despite extensive efforts and considerable progress (Janero, 2014; Kar and Roy, 2012; Singh et al., 2014a), prevention and cure of such disorders still

continue to be a major challenge for both traditionally known as well as modern systems of medicine.

Epidemiological, preclinical, and some clinical evidences now available on numerous edible plants strongly suggest though, that their modulating effects on psycho-biological processes involved pathogenesis and progression of lifestyle associated medical conditions are involved in their traditionally known health benefits (Dembinska-Kiec et al., 2008; Gonzalez-Castejon and Rodriguez-Casado, 2011; Baboota et al., 2013; Chandrasekaran et al., 2012; Chang et al., 2013a; Tiwari and Rao, 2002). It is now also well recognised that structurally and functionally diverse secondary plant metabolites are biosynthesized by them for defending themselves against environmental stress (Kwon et al., 2009), and that their traditionally known health benefits are most probably due to their modulating effects on environmental and metabolic stress triggered psychobiological processes involved in pathogenesis and progression of chronic diseases (Calabrese et al., 2012; Kennedy, 2014a). Dysregulation of these processes often leads to diverse spectrums of pathologies including obesity and diabetes and other silently progressing metabolic and inflammatory disorders (Cnop et al., 2012; Ozcan et al., 2004; Zhao and Ackerman, 2006). Amongst them, diabesity, i.e. obesity triggered type-2 diabetes, is the most rapidly spreading global epidemic of the 21st century now affecting all countries irrespective of their socioeconomic status and cultural background (Farag and Gaballa, 2011). The only currently available interventions with curative potentials against diabesity are bariatric surgeries (Kahn et al., 2014; Tschop and DiMarchi, 2012), and it is now well recognized that dietary therapies in combination with some antidiabetic or antihyperglycemic drugs (especially metformin) and physical

^{*}Correspondence: Vikas Kumar

E-mail: vikas.phe@iitbhu.ac.in

Received September 5, 2014; Accepted May 12, 2015; Published May 31, 2015

doi: http://dx.doi.org/10.5667/tang.2014.0026

^{© 2015} by Association of Humanitas Medicine

This is an open access article under the CC BY-NC license.

⁽http://creativecommons.org/licenses/by-nc/3.0/)

exercise are the most effective means for preventing progression of diabesity associated physical and mental health problems (Colagiuri, 2010; Pratley and Matfin, 2007). However, due to socioeconomic, cultural, and diverse other reasons, such therapeutic possibilities and recommendations are either not available, or are not affordable and acceptable, in many economically developing and underdeveloped countries where the burden of diabesity is the utmost (Pan et al., 1997; Ramachandran et al., 2006; Ramachandran et al., 2012).

Culinary uses of numerous edible plants used in Ayurvedic and other traditionally system of medicine and health care are very popular in all such countries, and irrespective of their socioeconomic and cultural backgrounds, herbal therapies are the most affordable and acceptable health care options for a vast majority of population in most of them. Therefore efforts are now being made in many laboratories to identify edible and other plants with anti-hyperglycemic, anti-inflammatory, insulin sensitivity improving and other diabesity associated pathologies (Eddouks et al., 2014; Chang et al., 2013b; Leiherer et al., 2013). A consistent observation made with diverse types of extracts of numerous such edible and other plants in animal behavioral models has been that their stress response regulating and other efficacies increase with increasing number of days of treatments (Chatterjee and Kumar, 2012; Kumar and Chatterjee, 2014a). These and numerous observations made during efforts to define other pharmacological activity profiles of several such extracts strongly suggest that biological mechanisms and processes involved in antihyperglycemic or anti-diabetic efficacies of edible plants could as well be due to their stress response modulating effects, and that structurally diverse secondary plant metabolites ubiquitously encountered in many, if not most, edible and other plants are also involved in their such efficacies (Langstieh et al., 2014; Shivavedi et al., 2014a; Shivavedi et al., 2014b; Rauniyar et al., 2015; Verma et al., 2015; Shakya et al., 2015). In this communication, available preclinical and clinical information on a few Ayurvedic edible plants justifying these inferences are summarized and critically analyzed and discussed in light of our current understanding on stress response regulating potentials of edible and other phytochemicals.

Brassica juncea (Mustard)

Commonly known as oriental or brown or Indian mustard, Brassica juncea L. is one of the numerous edible plants of the Brassicaceae family now widely used in India and other countries for obtaining mustard seeds and edible oil. It is a draught resistant plant (also often considered as weed by agricultural industries) widely cultivated for meeting the great commercial demand of mustard seeds and oil with nutty taste and pungent aroma. Its green leaves (commonly called mustard green) are also often used as spicy vegetables, salad, and pickled condiment in many Asiatic countries. Medicinal uses of diverse varieties of mustard plants are mentioned in classical Ayurvedic texts, and Brassica juncea is one of the several plants of the family mentioned in such texts (Manohar et al., 2009). Nutritive values of mustard green and diverse medicinal and health care potentials of mustard seeds and oils have been known since long not only in India, but also in numerous other countries. Although nutritive values of edible green leaves of Brassica juncea have also been mentioned in classical Ayurvedic texts, they are seldom used in Ayurvedic pharmaceutical formulations.

Information now available on medicinal phytochemistry of *Brassica juncea* leaves reveal though, that numerous bioactive phytochemicals encountered in them are either structurally

identical or functionally similar to those identified in not only in different varieties mustard seeds, but also in many other pharmacologically and clinically better scrutinized edible plants of the Brassicaceae family. Amongst oil producing plants of this family, Brassica juncea is currently one of the major mustard oil producing crops in India, which is one of the major vegetable oils commonly consumed in the country (Mishra and Manchanda, 2012; Singh et al., 2014b). It has been suggested that regular consumption of mustard oil together with vegetarian diet could be a feasible dietary means for lowering the risk of ischemic heart diseases in Indian population (Rastogi et al., 2004). Since bitter and pungent taste of mustard oil and all condeiments and vegetables prepared from plants of the Braceacea family are not always well accepted by many consumers (Drewnowski and Gomez-Carneros, 2000), they are often not regularly consumed, or are well accepted, for culinary purposess by many consumers. Therefore, efforts are now being made by food and agricultural industries to obtain Brassica juncea products devoid of such tastes (Sindhu et al., 2012).

Another reason for obtaining different cultivars of Brassica juncea is to obtain mustard oil with lower contents of Eruic acid, i.e an omega-9 fatty acids which has been reported to possess adverse health effects (Singh et al., 2013). Since potential adverse health effects of eucric acid (Sauer and Kramer, 1983; Choudhary et al., 2014), and also those of the pungent and bitter tasting isothiocyanate and other phytochemicals encountered in Brassica juncea derived products (Tripathi and Mishra, 2007; Inyang et al., 2014) have been observed in some animal bioassays, health care authorities of several countries have issued warnings against, or have even banned, edible uses of mustard oils (Oram et al., 2005; Wendlinger et al., 2014). However, as yet no very definitive statements based on preclinical, or clinical, or epidemiological evidence on maximally tolerated daily oral doses of such acids and numerous other phytochemicals commonly consumed with diverse types of mustard derived products can be made. This is mainly because appropriate dose response and other studies necessary for estimating their pharmacologically interesting dose ranges, safety profiles, and possible interactions between them are still missing.

On the other hand, the numbers of reports suggesting potential health benefits of diverse types of *Brassica juncea* leaf and seed extracts and their bioactive constituents have continued to increase during recent decades (Kumar et al., 2011a). Apart from vitamins, minerals, and nutritive proteins and lipids, the plant is now well recognized to be a rich source of a numerous phytochemicals often encountered in diverse other edible plants and well known for their cellular stress response modulating and hormetic effects (Birringer, 2011; Calabrese, 2010; Calabrese et al., 2010). However, since in Ayurvedic system of medicine mustard derived products are always used in combination with other natural products and health care procedures, therapeutic relevance of these findings in their traditionally known medicinal uses still remain questionable or speculative only.

The most extensively studied bitter and pungent tasting secondary metabolites encountered in *Brassica juncea* are the glucosinolates. Glucobrassicin, neoglucobrassicin, 4-methoxy glucobrassin and 4-hydroxy glucobrassicin are some indole glucosinolates more often encountered in *Brassica juncea* than in other plants of the family (Schreiner et al., 2009).

Quantitatively though, sinigrin is one of the major glucosinolate encountered in Indian mustard (Sang et al., 1984), which is now often considered to be a bioactive secondary metabolite of several edible plants with anti-cancer and antimicrobial activities (Patel et al., 2012). During processing of

associated comorbidities				
S. No	No Pharmacological activity References			
1	Anxiolytic, antiamnesic and antidepressant activity in diabetic rodents	Thakur et al., 2013a; Thakur et al., 2013b; Thakur et al., 2014a		
2	Anti-diabetic, antihyperglycemic, anti-obesity	Grover et al., 2002; Grover et al., 2003; Khan et al., 1995; Yadav et al., 2004		
3	Astrocyte developing	Joardar and Das, 2007		
4	Anti-oxidant and peroxynitrite scavenging	Jung et al., 2009; Khan et al., 1997; Kim et al., 2003		
5	Antiatherogenic and cholesterol metabolism	Jo et al., 1993; Khan et al., 1996		

Table 1. Pharmacological activities of diverse types of *Brassica juncea* extracts suggesting their curative or preventive potentials against diabesity associated comorbidities

mustard seeds glucosinolates are enzymaticaly degraded to allyl-isothiocyanate, which possesses strong bactericidal activities. Therefore, mustard meal powders are often recommended as a natural antimicrobial agent (Munday and Munday, 2002; Dai and Lim, 2014). Structurally diverse glucosinolates are also well known for their preventive potentials against cancer and other chronic diseases, including diabetic neuropathy and neurodegenerative diseases (Halkier and Gershenzon, 2006; Fahey et al., 2003; Tarozzi et al., 2013; Dinkova-Kostova and Kostov, 2012)

Essential oils of *Brassica juncea* consists mainly of a group of structurally analogous isothiocyanates. Some of them, also found in edible mustard oil, are allyl isothiocyanate, diallyl trisulfide, 3-butenyl isothiocyanate, allyl isothiocyanate, diallyl trisulfide and 3-butenyl isothiocyanate (Yu et al., 2003). It has been estimated that such oils consists of ca. 11% saturated and 89% unsaturated fatty acid of which about 18% is linoleic and 15% is linolenic fatty acid (Mishra and Manchanda, 2012). Other fatty acids found in *Brassica juncea* are erucic, eicosanoic, arachidic, nonadecanoic, behenic, oleic and palmitic acids, and arachidonic and α -linolenic acids are also been encounter in its oils (Kumar et al., 2011a). Brassicasterol, campesterol, β -sitosterol, Δ 5-avenasterol and trace amounts of Δ 7-stigmasterol have also been isolated and characterized from the mustard seed oil (Li et al., 2000).

Plant polyphenolics encountered in Brassica juncea, and in numerous other edible plants now attracting major attention of modern nutritionists and herbal researchers (Wang et al., 2014: Xiao and Hogger, 2015; Pandey and Rizvi, 2009). These are structurally diverse polyphenolic acids, quercetin, kaempferol, isorhamnetin and their naturally occurring conjugates, derivatives, and analogues (Cartea et al., 2010; Kumar and Andy, 2012; Kumar et al., 2012). Sinapic acid and its conjugates are quantitatively the major polyphenolics of Brassica juncea and it has been reported that it is one of richest natural sources of the acid and its conjugates (Niciforovic and Abramovic, 2014). Like for diverse other polyphenolic of Brassica juncea leaves, sinapic acid has also been identified as an antidiabetic agent with stress response modulating and antidepressant, anxiolytic and other brain function modulating activities (Cherng et al., 2013; Yoon et al., 2007).

Isorhamnetin is also another Brassica juncea specific and quantitatively major flavonid of mustard green with an analogous broad spectrum of therapeutically interesting bioactivity profile. Although numerous flavonoid glycosides with oxidative stress protecting activities have been isolated from Brassica juncea Leaves (Jung et al., 2009; Kim et al., 2002), isorhamnetin glycosides have been reported to be their major antidiabetic component with such activities (Yokozawa et al., 2002; Yokozawa et al., 2003). A comparative study on flavonoid contents of 91 vegetables has revealed an unique flavonol aglycone spectrum in mustard green, and found that it has the highest amount isorhamnetin amongst all vegetables analyzed in that study (Yang et al., 2013). It must be mentioned though, that isorhamnetin and its conjugates are also human metabolites of quercetin and other naturally more abundant flavonoids (Manach et al., 2004). However, like for all other plant extracts, the pharmacological activity profiles, or antidiabetic activity, of *Brassica juncea* leaf extracts cannot be predicted from their contents of phenolic plant metabolites, or by their antioxidative potentials, only (Thakur et al., 2013a; Thakur et al., 2013b; Thakur et al., 2014a).

Information now available on preclinical pharmacology of diverse types of extracts, and some of their often discussed bioactive constituents, suggesting their preventive and curative potentials against obesity and diabetes associated comorbidities are summarized in Tables 1 and 2 respectively. These and numerous other therapeutic possibilities offered by Brassica juncea and its bioactive secondary metabolites strongly suggest that modulation or regulation of psychological and metabolic stress triggered biological processes involved in etiology, pathogenesis, and progression of diverse lifestyle associated medical conditions and accelerated aging (Dietrich and Horvath, 2012; Epel, 2009; Fontana, 2009) are involved in their modes of actions. Consequently, efforts are now being made in several laboratories, including ours, to identify stress response modulating secondary metabolites of the plant, necessary for obtaining more rationally standardized extracts of the plant suitable for further developments as phytopharmaceuticals, or nutraceuticals, against diabesity and other lifestyle and environmental stress triggered mental health problems.

For such purposes, or for dietary therapy with Brassica juncea derived food, it is necessary to cultivate, harvest and process the appropriate variety or strain of the plant. This is not only because the content of bioactive secondary metabolites of the plant vary considerably in different strains and cultivars of the plant, but also due to the fact that it is an accumulator of toxic metals like lead, arsenic, cadmium etc. (John et al., 2009; Podar et al., 2004; Jakovljevic et al., 2013). Blood levels of such metals in diabetic patients are often fairly high (Akinloye et al., 2013; Kuo et al., 2013; Afridi et al., 2008), and their adverse effect potentials are well known. This is another reason behind the efforts of agricultural and food industries, and researchers to identify Brassica juncea cultivar strains and cultivation conditions for increasing crop yields with lower heavy metal contents. Although some strains of the plant with high crop yields have been identified, many questions concerning their contents of healthy or unhealthy substances still remain to be solved.

Such questions can be more rationally answered by collaborative efforts of medicinal phytochemists, herbal pharmacologists and toxicologists only. Availability of a convenient and well validated bioassays for identifying not only the therapy relevant secondary metabolites of the plant, but also for quantifying potential synergistic, antagonistic, additive effects between them is an essential prerequisite for such purposes. Since numerous known bioactive secondary metabolites of *Brassica juncea* and almost all edible medicinal plants possess physiological stress response regulating activity (Barrajon-Catalan et al., 2014; Joven et al., 2013; Lee et al., 2014; Tomas-Menor et al., 2015), it now apparent that traditionally known medicinal uses of mustards and other edible plants derived products in Ayurvedic system of medicine

and health care is mainly due the presence of specific combinations of such substances in Ayurvedic formulations or in Ayurvedic vegetarian food menus. Therefore, it can safely be suggested that the now well validated mouse bioassay evolving from efforts to identify and pharmacologically characterize the stress response regulating constituents of *Brassica juncea* and other edible plants (Langstieh et al., 2014) could be an useful one not only for better understanding of Ayurvedic pharmacology, but also for discovering and developing urgently needed novel preventive and curative pharmaco- therapies against diabesity and other lifestyle associated physical and mental health problems.

Piper longum

Piper longum L., commonly known as Pippali or "Indian long pepper", is a wildly growing and flowering perennial climber belonging to edible plants of Piperaceae family. It is now cultivated also for its fruits, which are usually dried and used as a spice and seasonings. Piper longum fruits have a similar, but somewhat hotter or more pungent taste than its close relative Piper nigrum, from which black, green and white pepper is obtained. The oldest known references of Piper longum come from ancient Ayurvedic texts where diverse medicinal and dietary uses of different parts of the plant are described (Manoj et al., 2004). In traditionally known Chinese system of medicine, the fruits are often used for treatments of hyperlipidemia, and during recent years, at least four structurally diverse anti-hyperlipidemic constituents, i.e. piperine, piperlongumine, pipernonaline, and 7, 4'-dimethyl ether of apigenin, have been identified from the fruits. According to these reports (Jin et al., 2009; Krishna et al., 2014), anti-hyperlipidemic efficacies of some of these Piper longum constituents in animal models are comparable to the currently widely used antihyperlipidemic drug simvastatin. However, the numbers of phytochemicals isolated from different parts of Piper longum, and their diverse bioactivities suggesting preventive and curative potentials of the plants against diverse malnutrition and other environmentally triggered inflammatory pathologies are not necessarily limited to these molecules only (Kumar et al., 2011b; Zaveri et al., 2010).

Reports on structurally and functionally novel secondary metabolites of the plant as well as their novel therapeutically interesting and other bioactivities, or of their since long known secondary metabolites, have continue to appear during recent years (Yang et al., 2013; Ahmed et al., 2014; Huang et al., 2010; Jiang et al., 2013; Ku et al., 2014; Yang et al., 2014, Yadav et al., 2014). Apart from proteins, carbohydrates, starch, and other nutritive constituents, and volatilile oils, most known bioactive molecules isolated from different parts of the plant are often structurally classified as alkaloids (alkaloidal amides), saponines, lignans, and phenolics etc. However, none of them are very specific biomarkers of the plant, and all of them have also been isolated from other edible plants of Piperacea and other families. Some of the numerous alkaloids or alkaloidal amides encontered in Piper longum fruits are: piperine, methyl piperine, iperonaline, piperettine, asarinine, pellitorine, piperun decalidine, piperlongumine, piperlonguminine, refractomide A, pregumidiene, brachystamide, brachystamide-A, brachystine, pipercide, piperderidine, longamide and tetrahydropiperine, terahydro piperlongumine, dehydropipernonaline piperidine, piperine, terahydropiperlongumine and trimethoxy cinnamoylpiperidine and piperlongumine (Zaveri et al., 2010). Piperine is often considered to be quantitatively the major (ca. 3 - 5% on dry weight basis) bioactive constituent of long pepper and it is also the major pungent tasting secondary metabolite of the plant. Therefore, it is now often used as a biomarker for analytically standardizing Piper longum extracts for experimental as well as commercial purposes.

Pungency of piperine is caused by activation of the heat and acidity sensing ion channel TRPV1 on pain sensing nerve cells (McNamara et al., 2005). Such capsaicin like effects of piperine and other pungent alkaloidal amides of Piper longum are most probably also involved in pain response modulating effects of its diverse types of extracts. Altered pain sensitivity is a cardinal symptom of diabetic neuropathy and other inflammatory disorders, and crucial role of TRPV1 channel in etiology, pathogenesis, and progression of diabesity associated pathologies are now well recognized (Suri and Szallasi, 2008). However, the questions concerning the involvement of these ion channels in pain response regulating functions of brain or in observed mental function regulating effects of Piper longum and other herbal extracts, cannot yet be answered with certainty (Gunthorpe and Szallasi, 2008). It remains certain though, that piperine and other alkaloidal amides with pungent and spicy tastes are modulators of the functions of TPRV1 channel (Rios and Olivo, 2014) and that as a desensitizer of this channel piperine is more efficacious than capsaicin (Szallasi, 2005).

Another bioactivity of piperine now attracting major attention of modern researchers is its ability to enhance bioavailability of other nutrients and drugs (Patil et al., 2011). Similar or analogous effects are also known for numerous other edible phytochemicals commonly consumed with every day meals (Muttepawar et al., 2014; Tatiraju et al., 2013). Although as yet no very definitive statements on biological mechanisms and process involved in such effects of piperine can be made, it remains certain that its regular consumption can enhance oral bioavailability of other essential nutrients and edible phytochemicals commonly consumed with everyday meals, or with phyto-pharmaceuticals and nutraceuticals. It has been reported, indeed, that piperine enhances oral bioavailability of the edible antidiabetic phytochemical curcumin by 2000% in human (Shoba et al., 1998), which is the quantitatively major bioactive constituent of another Ayurvedic herb Curcuma longa

Bioactive constituent Pharmacological activity S. No. References Phenyl isothiocyanate Antitumour and antioxidant Manesh and Kuttan, 2003 1 Cognition improving and beneficial in scopolamine induced Karakida et al., 2007 amnesia Hepatoprotective Shin et al., 2013 2 Sinapic acid Roy and Stanely Mainzen Prince, Cardioprotective 2012 Neuroprotective Lee et al., 2012 Anxiolytic Yoon et al., 2007 Jung et al., 2009 3 Kaempferol glycosides Antioxidant Isorhamnetin 3,7-di-O-ß-D-glucopyranoside Yokozawa et al., 2002; Yokozawa 4 Antioxidant, antidiabetic (Isorhamnetindiglucoside) et al., 2003

Table 2. Some major bioactive constituents of Brassica juncea identified to date, and their often-cited pharmacological activities

Table 3. Some reports suggesting therapeutic potentials of *Piper longum* extracts against diabesity and associated comorbidities

S. No.	Pharmacological activity	References
1	Analgesic	Vedhanayaki et al., 2003
2	Antidiabetic and antioxidant	Nabi et al., 2013; Samudram, 2009; Thomas et al., 2009; Wakade et al., 2008
3	Antihepatotoxic and hepatoprotective	Patel and Shah, 2009; Samudram, 2009; Rajeswary et al., 2011
4	Antiinflammatory	Wakade et al., 2008; Kumar et al., 2009; Singh et al., 2008; Vaghasiya et al., 2007
5	Immunomodulatory	Sunila and Kuttan, 2004

(to be described later). These and numerous other analogous observations made with piperine and other edible phytochemicals strongly suggest that modulations of bioavailability of essential nutrients are also involved in their observed beneficial effects on metabolic disorders, and that proper understanding of their Ayurvedic pharmacology and medicinal values is possible only when due attention is paid to their such properties.

Another major alkaloidal amide encountered in Piper longum and attracting major attention of modern drug discoverers is piperlongumine, or piplartin (Bezerra et al., 2013). It is not encountered in Piper nigrum, the fruits of which are the most commonly consumed spice in the western world and elsewhere (Bezerra et al., 2013). Preclinical information now available on piplartin strongly suggest that it is most probably also one of the major bioactive secondary plant metabolites involved in diverse therapeutically interesting mental and metabolic function modulating effects of Piper longum fruits and roots (Bezerra et al., 2013), and that it could be a promising cancer therapeutic lead as well (Wu et al., 2014). Although piperlongumine and piperine are the two quantitatively the major bioactive constituents of numerous piper species commonly used in numerous Ayurvedic formulations (Meghwal and Goswami, 2013), their quantities vary considerable not only in different piper species but also in different parts of Piper longum and other plants of the species (Bao et al., 2014; Chandra et al., 2014).

Piplartin was first isolated from Piper longum roots, dried powder of which are also often used by Ayurvedic and other practitioners of herbal medicine for treatments of insomnia and debility caused by chronic fever (Murthy, 2009), and diverse types of extracts of the roots and their combinations with other herbal extracts are now also commercialized in India as Ayurvedic remedies (Rajopadhye et al., 2012). Such remedies are often used for treatment of rheumatism and other inflammatory conditions, and aqueous suspension of powdered roots of *Piper longum* root, commonly called *Pippalimula* in India, has been reported to possess ibuprofen like analgesic activity (Vedhanayaki et al., 2003). During recent years, several reports revealing antidiabetic activities of aqueous and ethanolic extracts of Piper longum roots have also appeared (Chaurasia and Das, 2013; Chaurasia et al., 2012; Nabi et al., 2013). Extensive psycho-pharmacological and some clinical observations made in our Banaras Hindu University's Ayurvedic hospital with Piper longum root powder have not only revealed its broad spectrum of therapeutically interesting neuronal function modulating activities, but also indicated that metabolic as well as psychological stress response modulating effects are also involved in its modes of actions. Since stress response regulating properties of a Piper longum fruits containing Ayurvedic formulation has been reported (Neha and Mishra, 2011), and several bioactive phytochemicals encountered in the roots and fruits used in the formulation are the same, it could as well be that the reported antidiabetic activity of the root extracts are also due to the presence of stress response regulating components in them.

Major therapeutically interesting bioactivities of diverse types of *Piper longum* extracts and their quantitatively major bioactive constituents indicating therapeutic potentials of the plant for prevention and cure of obesity associated comorbidities are summarized in Tables 3 and 4 respectively. Although several recent reports have already revealed antiobesity activity of piperine containing Piper longum oil in animal models (BrahmaNaidu et al., 2014; Choi et al., 2013; Doucette et al., 2013; Kumar et al., 2013; Noble et al., 2013), many questions concernig the roles of other bioactive constituents in such oils still remain open. Moreover, many such reports pay little attention to the fact that piperine and other constituents of such oils also possesses mental function regulating activities (Cicero Bezerra Felipe et al., 2007; Gilhotra and Dhingra, 2014; Li et al., 2007a; Li et al., 2007b; Mao et al., 2014; Pal et al., 2011), and that their modulating effects on central nervous system could as well be involved in anti-obesity and other health benefits of *Piper longum* derived products against type-2 diabetes (Sandoval et al., 2009; Patrone et al., 2014; Banks et al., 2012). It cannot be ignored though, that due to very hot taste of such products, they might not be well suited for incorporating them in everyday meals in high enough quantities necessary for obtaining health benefits from them.

Amlaki and Triphala

Amlaki is the Sanskrit name of the edible fruits of Phyllanthus emblica (synonym: Emblica officinalis; common name: Indian gooseberry) most commonly used in Ayurvedic system of medicine and health care as a tonic and for prevention and cure of physical as well as mental health problems arising from intricate disorders of digestive and excretory organs. It is also one of the most widely used edible fruits in numerous Ayurvedic formulations. One such very popular formulation is Triphala, which consists of equal parts of dried edible fruits of Embelica officinalis, Terminalia chebula, and Terminalia bellirica (Belapurkar et al., 2014; Maheshwari and Rajnee, 2014). Preventive and curative use of both Amlaki as well as of Triphala with barley and other dietary measures for treatments of diabetes is recommended in ancient Ayurvedic texts, and their such medicinal uses are now well justified by numerous preclinical and some therapeutic observations reported during recent decades (Sharma and Chandola, 2011a; Sharma and Chandola, 2011b; D'Souza et al., 2014; Mohammad and Larijani, 2013; Tiwari, 2008; Rajan and Antony, 2008). In classical Ayurvedic texts numerous other formulations containing them are mentioned also (Chulet and Pradhan, 2009; Chouhan et al., 2013), and in traditionally known Indian, Chinese, and Arabic systems of medicine Triphala is more often than not used in combination with other medicinal herbal herbs and other health care practices (Mohammad and Larijani, 2013; Zaki et al., 2014). Amongst the three edible fruits constituting Triphala formulations, the one most widely harvested and consumed for culinary purposes is Amlaki.

Recent neuropsychopharmacological observations (Kumar and Chatterjee, 2014b; Dhanalakshmi et al., 2007; Dhanalakshmi et al., 2006; Nariya et al., 2011; Rinki and Mishra, 2011; Srikumar et al., 2006) have revealed a broad spectrum of neuronal function modulating effects of *Triphala*, and suggest that its modulatory effects against glucose toxicity and diverse metabolic or mental stress triggered biological responses could be involved in its modes of actions. It is a sour, bitter, and astringent tasting fruit, which is often steeped in salt water and turmeric to make the sour fruits palatable. Gallic and Elegiac acids and their conjugates are some of the quantitatively major bioactive constituents of all the three fruits composing Triphala (Patel and Shah, 2009). However, the spectrum of known bioactive secondary plant metabolites encountered in Amlaki is not identical to those known for the other two fruits commonly used in this and numerous other Ayurvedic formulation. Available information on the bioactive constituents, and experimental evidences suggesting therapeutic potentials of Amlaki for treatments of diabesity and diverse other diseases have often been reviewed during recent years (D'Souza et al., 2014; Dasaroju and Gottumukkala, 2014; Khosla and Sharma, 2012; Krishnaveni and Mirunalini, 2010; Sri et al., 2013). Amlaki is also one of the several Ayurvedic medicinal herbs currently universally well recognized by modern herbalists and herbal researchers as an herbal adaptogen (Winston and Maimes, 2007). The concept of herbal adaptogens evolved originally in Russia during late 1940s (Brekhman and Dardymov, 1969), and the very first more systematic and detailed report on adaptogenic, or biological stress response regulating, effects of Amlaki and a few other edible and other Ayurvedic medicinal plant appeared in 1999 (Rege et al., 1999). Since then this concept has been well accepted by almost all modern scholars and practitioners of Ayurvedic system of medicine, and efforts are now being made by modern researchers to experimentally verify the efficacies of combinations of Amlaki with other edible plants and Ayurvedic therapeutic practices for prevention and cure of type-2 diabetes (Tripathi et al., 2012; Vaibhavi et al., 2013).

One of the several Amlaki formulation specially recommended in classical Ayurvedic texts for treatment of diabetes-associated comorbidities is Nishamlaki, which consists of Amlaki and Turmeric rhizome (to be described later). They are also the two major constituents of a vast majority of herbal formulations currently commonly prescribed in India for treatments of diabesity (Sarma et al., 2014). Therefore, efforts are now being made to better standardize such a formulation that could eventually be more rationally developed and used for prevention and cure of diabetes (Rao et al., 2013; Venkateshwarlu et al., 2013). A recent report have revealed though, that inappropriate formulations or doses of such preparations could as well have adverse drug-drug interaction with metformin (Puranik et al., 2014), i.e. one of the main antidiabetic drugs commonly recommended for treatments of type-2 diabetes. Efforts to answer the question whether Amlaki, or Turmeric, or inappropriate combinations of the two are involved in such herb-drug interaction will be necessary for more rational uses of the two edibles for prevention and cure of diabesity. Moreover, since both Amlaki as well turmeric are often consumed in India with every day meals, and both diabetes and obesity are also the most common metabolic disorders encountered in Indian, China, and other developing countries, efforts to better clarify the situation is urgently needed for obtaining appropriate medicinal benefits from metformin and other antidiabetic drugs in these countries (Neerati et al., 2012).

Available information on medicinal phytochemistry of the other two fruits constituting Triphala strongly suggest that they could also have adverse herb-drug interactions with metformin and other drugs currently often prescribed for prevention and cure of diabesity associated physical and mental health problems (Fasinu et al., 2012). Although a few known bioactive secondary metabolites of Emblica officinalis (D'Souza et al., 2014; Khosla and Sharma, 2012; Zhang et al., 2001; Rastogi and Mehrotra, 1995; Asolkar et al., 1992; Dasaroju and Gottumukkala, 2014), are not encountered in Terminalia chebula (Rastogi and Mehrotra, 1995; Saleem et al., 2002; Kokate et al., 2003; Rathinamoorthy and Thilagavathi, 2014; Walia and Arora, 2013; Gupta, 2012) or in Terminalia bellirica (Saleem et al., 2002; Kokate et al., 2003; Kadian et al., 2014; Kumudhavalli et al., 2010; Saxena et al., 2013), all of them are fairly rich sources of tannins, gallic acids and their conjugates, and several other secondary plant metabolites ubiquitously present in many edible and other plants and well known for their modulating effects on drug metabolizing enzymes and their bioavailability (Serrano et al., 2009; Anannarukan et al., 2012). It has been reported that drug metabolizing enzyme inhibitory properties of Triphala are involved in such observed effects of the formulation, and that in this respect all the three constituents of the formulation are almost equipotent (Ponnusankar et al., 2011). According to this report, gallic acid and its conjugates and derivatives are also their common polyphenolic constituent involved in such effects of Triphala.

It must be mentioned though, that gallic acid conjugates and other plant polyphenolics are also encountered in numerous other edible and medicinal plants, and that they are now well recognized for their protective effects against oxidative stress triggered pathologies, including diabetes and associated psychopathologies (Lee et al., 2014; Dragan et al., 2015; Khadem and Marles, 2010; Lephart, 2015; Liu et al., 2015; Santilli et al., 2015). Moreover, analgesic, anti-inflammatory, antidiabetic, anxiolytic, antidepressant, cognitive function modulating, antimicrobial, and diverse other therapeutically interesting bioactivities of diverse types of extracts of Amlaki and Triphala enriched in such phytochemicals also have often been reported. Detailed discussions and critical analysis of these reports dealing with their diverse such bioactivities is beyond the scope of this communication. For such purposes, the already cited references have to be consulted.

It must be mentioned though, that numerous observations made with *Amlaki*, *Triphala*, and numerous other edible plant derived products have consistently revealed that their oral efficacies for stress responses modulating and other bioactivities increase with increasing numbers of treatment days (Kumar and Chatterjee, 2014a), and that this is most probably due to their modulating effects on microbial ecology inside the gastrointestinal tract (Thakur et al., 2014b). Although

Table 4. Major bioactive constituents of Piper longum L. identified to date, and their often-cited pharmacological activities

S. No.	Bioactive constituent	Pharmacological activity	References
1	Piperine	Antihyperlipidemic	Maneesa et al., 2012
		Antidepressant	Li et al., 2007b
		Anticonvulsant	Mori et al., 1985
		Immunomodulatory	Pathak and Khandelwal, 2009
		Antiinflammatory	Kumar et al., 2009
		Antidiabetic and antiobesity	Lee et al., 2006
2	Methylpiperate	Antidepressant	Lee et al., 2008
3	Beta- Caryophyllene	Local anesthetic	Ghelardini et al., 2001
4	Piperine derivative	Hypolipidemic	Bao et al., 2012
5	Piperlongumine (piplartine)	Antinociceptive, anxiolytic, antidepressant, antidiabetic, antihyperlipidemic	Jin et al., 2009; Bezerra et al., 2013

our current knowledge on the bactericidal constituents of Triphala and its constituents is far from being satisfactory, evidence now available on their diverse types of formulations strongly suggest that Gallic and Ellagic acids and their conjugates and soluble and insoluble polymers (commonly referred to as tannins) are their major secondary metabolites involved in bactericidal activities of their commercialized extracts (Biradar et al., 2008). It is now well recognized that gut microbial ecology plays a crucial role in the etiology, pathogenesis and progression of diabesity (Burcelin et al., 2011; Everard and Cani, 2013), and that such and other plant phenolics and their metabolites are regulators of gut microbial ecology (Bolca et al., 2013; van Duynhoven et al., 2011). Therefore, it is now apparent that proper understanding of their modulating effects on gut microbial ecology is an essential prerequisite for their more rational uses as dietary therapies against diabesity and associated mental health problems (Dinan and Cryan, 2012; Wang and Tang, 2015).

Curcuma longa (Turmeric)

Curcuma longa (synonym: *Curcuma domestica* and commonly called turmeric) is one of the several plants of the Zingiberaceae family (genus: curcuma), well known since antiquity to the scholars and practitioners of traditionally known systems of medicine for diverse health benefits of their rhizomes. It is now widely cultivated and diversely processed in India, China, and many other countries for obtaining dried roots and other products from them for culinary as well as medicinal purposes (Li et al., 2011). In Ayurvedic and other traditionally known Indian system of medicine, turmeric is now often used as a general tonic and blood purifier and also for prevention and cure of inflammatory diseases. Currently, it is

phytochemicallly and pharmacologically one of the more extensively studied edible plant derived products of medicinal interest. Numerous reviews and monographs describing diverse therapeutic possibilities offered by the plant and its bioactive secondary metabolites have appeared during recent years (Chaudhary et al., 2010; Gupta et al., 2013; Chempakam and Parthasarathy, 2008). Available preclinical and clinical information on anti-diabetic potentials of the plant has also been summarized in a recent issue of this journal (Ponnusamy et al., 2012). Most such reviews and reports often neglect though, that modulating effects of curcumin and other turmeric curcuminoids on brain functions could as well be involved in their modes of actions and health benefits. Some of the reports suggesting such possibility for curcumin and two other Turmeric curcuminoids are summarized in Table 5.

Amongst numerous known bioactive and therapeutically interesting secondary metabolites of turmeric identified and quantified to date, the three structurally and functionally analogous ones with antioxidative properties, i.e. curcumin, desmethoxycurcumin, and bisdesmethoxycurcumine (often collectively referred to as curcuminoids), have attracted the most attention of modern herbal researchers and drug discoverers (Aggarwal et al., 2007; Ahmed and Gilani, 2014; Alappat and Awad, 2010; Bradford, 2013; Grynkiewicz and Slifirski, 2012; Javaprakasha et al., 2006; Lopresti et al., 2014; Sahebkar, 2013). Hereupon, by far a vast majority of preclinical and clinical reports deal mainly on two major obesity associated medical conditions, i.e. diabetes and cancer (Zhang et al., 2013). These efforts have not only continued to add preclinical and clinical evidences suggesting their therapeutic potentials against these and diverse other malnutrition associated health problems, but also have leaded to better

Table 5. Some reported neuro-pharmacological activities of Curcumin and of two other Turmeric curcuminoids

Turmeric constituent	Animal Species	Pharmacological activity	Dose, Duration and route of administration	References
	Mice	Anti-alzheimer's activity	7.5 mg/kg, 7 days, i.v.	Garcia-Alloza et al., 2007
	In vitro	Anti-alzheimer's activity	50 µM	Ono et al., 2004
	In vitro	Anti-alzheimer's activity	20 µM, 7 days	Qin et al., 2010
	In vitro	Anti-alzheimer's activity	$7.1 \pm 0.3 \mu g/ml$ curcumin	Kim et al., 2001
	Mice	Anti-alzheimer's activity	200 µl, single dose, i.v.	Yang et al., 2005
	Mice	Antidepressant	100 mg/kg, 14 days, p.o.	Sanmukhani et al., 201
	Mice	Antidepressant	10-80 mg/kg, i.p.	Kulkarni et al., 2009
	Rats	Antidepressant	70 mg/kg, 14 days, p.o.	Patel et al., 2014
	Rats	Antidepressant	2.5 and 5.0 mg/kg, 14 days, p.o.	Xu et al., 2005
	Mice	Antiepileptic	50, 100 and 200 mg/kg, 5 weeks, p.o.	Agarwal et al., 2011
	Rats	Antiepileptic	100 and 300 mg/kg, single dose, i.p.	Peng et al., 2009
	Rats	Antiepileptic	100 mg/kg, 7 days, p.o.	Kiasalari et al., 2013
	Mice	Antiepileptic	100 mg/kg, 21 days, p.o.	Bharal et al., 2008
	Mice	Antiepileptic	200 mg/kg, i.p.	Shin et al., 2007
Curcumin	Rats	Anxiolytic	10 mg/kg, 14 days, p.o.	Chimakurthy and Talasi 2010
	Mice	Anxiolytic	100 and 300 mg/kg, 14 days, p.o.	Abu-Taweel et al., 201
	Rats	Neuroprotective	50 and 100 mg/kg, single, i.p	Yang et al., 2009
	Rats	Neuroprotective	1-2 mg/kg, single dose, i.v.	Jiang et al., 2007
	Mice	Memory function	50 mg/kg, 21 days, i.p.	Yu et al., 2013
	Mice	Memory function	50, 100 and 200 mg/kg, 15 days, i.p.	Choudhary et al., 2013
	Mice	Memory function	40 mg/kg, single dose, i.p.	Yu et al., 2013
	In vitro	Parkinson's disease	500 nM	Liu et al., 2011
	In vitro	Parkinson's disease	20 and 25 mmol/L	Wang et al., 2009
	In vitro	Neuroprotective	5 and 10 µM	Wang et al., 2010
	Rats	Neuroprotective	100 and 300 mg/kg, single dose, i.p.	Thiyagarajan and Sharm 2004
	Rats	Neuroprotective	20 mg/kg, 6 days, i.p.	Kuo et al., 2011
	Mice	Adaptogenic	25 and 50 mg/kg, 10 days, p.o.	Bhatia et al., 2011
Desmethoxy-curcumin	In vitro	Anti-alzheimer's activity	$4.7\pm0.1~\mu g/ml$	Kim et al., 2001
is-desmethoxy-curcumin	In vitro	Anti-alzheimer's activity	$3.5 \pm 0.2 \mu g/ml$	Kim et al., 2001

understanding of medicinal phytochemistry and molecular pharmacology of curcuminoids (Grynkiewicz and Slifirski, 2012; Priyadarsini, 2014; Brodniewicz and Grynkiewicz, 2012). Some reports suggesting therapeutic potentials of *Curcuma longa* extracts against diabesity and associated comorbidities are summarized in Tables 6.

However, curcumin and curcuminoids are not the only bioactive secondary metabolites of Curcuma longa potentially involved in diverse traditionally known, or recently identified, therapeutic potentials of turmeric (Aggarwal et al., 2013; Bhanumathy et al., 2013; Kasabri et al., 2014; Lekshmi et al., 2012). Although the list of bioactive phytochemicals, vitamins, and other micronutrients commonly consumed with turmeric, and which could also be involved in traditionally known health benefits of the spice have consistently been enlarged during the past few decades (Wang et al., 2014; Mirmiran et al., 2014; Vaidya, 2014), many questions concerning oral bioavailability, and the doses and treatment regimen necessary for obtaining therapeutic benefits from curcuminoids, or for that matter for any of the till now known bioactive secondary metabolites of turmeric, cannot yet be answered with any certainty (Anand et al., 2007). Some observations made in vitro, suggest that oral bioavailability of curcuminoids from turmeric could as well be higher than those of pure curcuminoids (Maheshwari, 2010). However, therapeutic relevance of these observations still remains questionable. This is mainly because numerous metabolic and behavioral effects observed after daily oral doses of curcuminoids seldom correlate with their blood levels observed after their single or repeated daily oral doses. Such is specially the case for their stress triggered brain function modulating effects in experimental animals observed after their oral doses commonly used for assessing their therapeutic potentials against diabesity and other metabolic disorders (Xia et al., 2011; Xia et al., 2006).

Taken together, these observations clearly suggest that health benefits of curcuminoids and other turmeric phytochemicals are either due to their bioactive metabolites, or that their observed broad spectrum of bioactivities are due to their non-systemic effects. Since turmeric is always consumed with other food ingredients containing diverse other bioactive molecules with broad spectrums of bioactivity profiles, it is almost certain that traditionally known health benefits of turmeric, cannot entirely be due to its curcuminoids contents, or on their blood levels observed after turmeric intake, only. Since analogous is the situation for almost all medicinally used edible plants and phytochemicals, attempts are now being made in several laboratories to use system-biology based pharmacological approaches for more rationally solving such problems (Goodacre, 2007; Nyanginja and Mponda, 2014; van Ommen and Stierum, 2002). However, during most such efforts attempts are made to better define the cellular and molecular mechanisms potentially involved in their modes of action, and as yet only very little attention has been paid to potential role of the so called microbiota-gut-brain axis in their observed effects in experimental animals and human volunteers and patients (Thakur et al., 2014b; Chiou et al., 2014; Greiner et al., 2014). However, the possibility that modulation of colonic microbiota could be involved in colon cancer prevention has recently been pointed out also (McFadden et al., 2014).

Some major difficulties encountered during efforts to

extrapolate preclinical findings made with turmeric extracts and their known bioactivities in terms of traditionally known medicinal uses of the turmeric arise also from the fact that depending on harvesting and processing procedures used for medicinal value and the contents of curcuminoids and other bioactive constituents vary considerably (Pal et al., 2008). A recently reported computer assisted study have revealed that at least 200 structurally and functionally diverse bioactive phytochemicals can be expected to be present in a given turmeric sample (Balaji and Chempakam, 2010), and that many of them have them have adverse effect potential as well. Results of this in silico study have revealed that out of the 200 compounds screened, 184 were predictably toxigenic, 136 mutagenic, 153 carcinogenic, and 64 hepatotoxic, and that only 16 of them are devoid of any of these adverse effects detectable by the in silico procedure used. Although this report supplies an exhaustive list of bioactive secondary metabolites of Curcuma longa, predictive validity of the observations reported there must be judged with caution (Balaji and Chempakam, 2010). Numerous toxicological and safety reports on diverse types of products derived from the plant have always pointed out that their adverse effect can be expected only after their extremely high daily oral doses that cannot be consumed with every day meals or with Ayurvedic formulations containing them (Chainani-Wu, 2003; Joshi et al., 2003; Lao et al., 2006; Madhu et al., 2013; Micucci et al., 2013; Qureshi et al., 1992; Ulbricht et al., 2011; Velusami et al., 2013; Liju et al., 2013; Hasan et al., 2014).

Some high dose adverse effect potentials of phytochemicals often cited deal with their possible effect on liver functions (Kandarkar et al., 1998; Babu and Srinivasan, 1997), and those of gastrointestinal tract and skin (Fetrow and Avila, 1999). Unfortunately, as yet only a very few scattered reports on dose response studies necessary for predicting therapeutically interesting and safety margin of Curcuma longa extracts (other than those highly concentrated in curcumin and curcuminoids) have appeared (Micucci et al., 2013; Joshi et al., 2011). Results of one such recently reported preliminary study conducted in mice have revealed not only metformin (currently the drug of first choice for prevention and cure of diabesity) like stress response suppressing effects of fairly low daily oral doses of curcumin and some Curcuma longa extracts enriched in curcuminoids, but also suggest that even more than their 50 fold higher daily oral doses are fairly well tolerated by both male and female animals (Verma et al., 2015, Verma et al., 2014). Recent observations made in our laboratories with turmeric oil devoid of curcumin indicate that analogous is also the case for such oils (manuscript in preparation). These observations, and numerous other revealing that curcumin and diverse other turmeric constituents modulate the functions of stress responses mediated by shock proteins (Ali and Rattan, 2006; Speciale et al., 2011), add experimental evidences in support of the convictions that stress response regulating effects of turmeric derived products are involved in their modes of action, and that curcuminoids are not their only antidiabetic or stress response modulating bioactive constituents.

Phytochemicals, stress and diabesity

The term "diabesity" was initially coined during early 1970s to describe strong pathogenic links between obesity and type-2

Table 6. Reports suggesting therapeutic potentials of Curcuma longa extracts against diabesity and associated comorbidities

S. No.	Pharmacological activity	References
1	Antidiabetic, antioxidant and antiobesity	Kuroda et al., 2005; Liju et al., 2011
2	Anti-inflammatory and analgesic	Liju et al., 2011
3	Antidepressant	Xia et al., 2007; Yu et al., 2002
4	Beneficial for liver functions	Soni et al., 1992

diabetes (Sims et al., 1973). Although since then it has been well established that complex interactions between obesity, insulin resistance, and pancreatic ß-cell dysfunction cause type-2 diabetes, biological mechanisms and processes regulating the interplay among these impairments still remain to be better defined (Tschop and DiMarchi, 2012; Martinez and Milagro, 2015). However, it is now well recognized that even modest reduction of body weight can lead to significant improvements in glucose homeostasis of patients suffering from, or at risk to, diabesity (Martinez et al., 2014; Pati et al., 2014), and that uncontrollable stressful events and chronic stress states have significant and positive association with weight gains and type-2 diabesity (Adam and Epel, 2007; Dallman et al., 2005; Steptoe et al., 2014). This is most probably due to behavioral alterations triggered by environmental or mental stress, which eventually leads to addiction-like eating behavior (Ahmed et al., 2013; Ginty, 2013; Ginty et al., 2012; Scott and Johnstone, 2012; Sinha and Jastreboff, 2013). Although several questions concerning individual food components involved in "addictive eating behavior" still remain open (Ahmed et al., 2013; Davis, 2014; Meule et al., 2014), it is now almost certain that proper modulation of this behavior could indeed be an effective strategy for prevention of obesity associated physical and mental health problems (Meule et al., 2014; Murray et al., 2015; Pedram and Sun, 2015).

Amongst structurally diverse secondary plant metabolites with stress response modulating and other therapeutically interesting bioactivities, polyphenolics have attracted the most attention of modern nutritional researchers and pharmacologists (Lee et al., 2014; Nowak, 2015; Pa and Gazzaley, 2014). Although their health benefits are often considered to be due to their antioxidative properties, like other diverse phytochemicals (edible or not), they also possess broad spectrums of bactericidal, anti-inflammatory, immune function modulating, and numerous other therapeutically interesting bioactivities (Kennedy, 2014a; Kennedy, 2014b). Mounting preclinical and clinical evidences accumulated during the past few decades strongly suggest that their modulating effects on gut microbial ecology and digestive functions are also involved in their modes of actions (Thakur et al., 2014b). It is now well recognized that metabolic and psychological stress responses also alter gut microbial ecology, which in turn alters the functions of the gut-brain axis (Aroniadis and Brandt, 2013; Mayer, 2011; Moloney et al., 2014). According to the postmodern eco-physiological and pharmacological concept arising from these findings (Abedon, 2014), circulating blood levels of edible phytochemicals observed after their oral intake must not necessarily be very reliable indicators of their brain function modulating effects. Available information on metabolic fate of numerous edible polyphenolics (van Duynhoven et al., 2011) is also in agreement with this inference.

Eco-physiological and other studies have now established that secondary plant metabolites afford survival benefits to plants against enviorenmental stress (Kennedy, 2014a; Kennedy, 2014b; Trowbridge, 2014), and that structurally and functionally diverse edible phytochemicals possess pleiotropic protective effects against stress responses and are potentially useful for prevention and cure of diabetes, Alzheimer's disease and other chronic silently progressing chronic diseases (Dembinska-Kiec et al., 2008; Leiherer et al., 2013; Davinelli et al., 2012; Vaiserman, 2014; Carriba and Comella, 2014; Franco and Cedazo-Minguez, 2014; Ruden and Lu, 2011). However, it has been also reported that depending on the components of whole-food some of them could as well worsen cognitive dysfunctions (Parrott et al., 2015). Available information on dose response relationship of numerous nutritive and other phytochemicals have revealed indeed that

their protective or amplifying or adverse effects on stress responses depend on their daily doses and treatment regimen used, and that hereupon their beneficial effects often predominates (Calabrese et al., 2012; Le Bourg and Rattan, 2014; McClure et al., 2014). Such dose response relationships of bioactive molecules are due to their dose dependant modulating effects on diverse cellular adaptive stress responses (Birringer, 2011; Costantini et al., 2010) which often leads to their experimentally observed inverted U or J shaped dose response curves, or hormetic effects, in bioassays (Bao et al., 2014; Lushchak, 2014a). Evaluation of dynamics of such effects of edible phytochemicals are essential prerequisite for understanding of their pharmacokinetic better and pharmacodynamic properties necessary for obtaining health benefits offered by them (Lushchak, 2014b). This is because exposures to short term stress (hormetic stress) can strengthen subsequent response to stress, and prolonged stress exposures leads to toxic stress which shorten life span, and also leads to mental health problems (Lee et al., 2014; Epel and Lithgow, 2014; Scapagnini et al., 2014)

DISCUSSION

The possibility that physiological or psychological stress is a major contributing factor to health problems triggered or caused by obesity and diabetes have often been pointed out during recent decades (Brindley and Rolland, 1989; Gastaldi and Ruiz, 2009: Kelly and Ismail, 2015). It is now well recognized also that the functions of all bodily or gas including those of the brain, and gut microbiota are altered by metabolic as well as mental and environmental stress (Steptoe et al., 2014; Carvalho et al., 2015; Cryan and Dinan, 2012; Foster and McVey Neufeld, 2013; Nicholson etal., 2012; Rhee et al., 2009). Although modern researchers have now well recognized that targeting the central nervous system is a promising approach for discovering drugs against obesity associated diabetes and other metabolic disorders (Sandoval et al., 2009; Lin and Sun, 2010; Perez-Tilve et al., 2012), most modern drug discoverers still continue to neglect that gut microbial ecology plays a crucial role in regulations of all bodily and mental functions (Baty et al., 2014; Triggle, 2012). Recent reports have pointed out though, that dietary phytochemicals are regulators of gut microbial ecology (D'Aversa et al., 2013), and that plant and other natural products libraries are valuable sources for identifying drug leads acting on proteins regulating stress responses (Davenport et al., 2014).

As summarized in this communication, many edible plants traditionally known for their medicinal values are pleiotropic stress protective agents with anti-obesity, antidiabetic, antidepressant, anxiolytic, memory function modulating, analgesic and other brain function modulating activities. Diverse combinations of their secondary metabolites with demonstrated stress response regulating and broad spectrums of pharmacologically interesting bioactivity profiles are encountered in almost all terrestrial plants (medicinal, or not) as well. These findings not only justify their medicinal uses in Ayurvedic and other traditionally known systems of medicine and health care, but also strongly suggest that more holistic pharmacological strategies will be necessary for better understanding of therapeutic potentials of traditionally known medicinal plants, or for obtaining novel therapeutic leads from edible and other plant derived products containing edible phytochemicals. During such efforts due attention has to be paid not only to their stress response regulating and microbicidal properties, but also to the fact their adaptogenics like efficacies increases with increasing number of treatment days.

ACKNOWLEDGEMENTS

None.

CONFLICT OF INTEREST

The authors do not have any conflict of interest in the present study.

REFERENCES

Abedon ST. Phage therapy: Eco-physiological pharmacology. Scientifica (Cairo). 2014;2014:581639.

Abu-Taweel GM, Ajarem JS, Ahmad M. Protective effect of curcumin on anxiety, learning behavior, neuromuscular activities, brain neurotransmitters and oxidative stress enzymes in cadmium intoxicated mice. J Behav Brain Sci. 2013;3:74-84.

Adam TC, Epel ES. Stress, eating and the reward system. Physiol Behav. 2007;91:449-458.

Afridi HI, Kazi TG, Kazi N, Jamali MK, Arain MB, Jalbani N, Baig JA, Sarfraz RA. Evaluation of status of toxic metals in biological samples of diabetes mellitus patients. Diabetes Res Clin Pract. 2008;80:280-288.

Agarwal NB, Jain S, Agarwal NK, Mediratta PK, Sharma KK. Modulation of pentylenetetrazole-induced kindling and oxidative stress by curcumin in mice. Phytomedicine. 2011;18:756-759.

Aggarwal B, Sundaram C, Malani N, Ichikawa H. Curcumin: The indian solid gold. In The molecular targets and therapeutic uses of curcumin in health and disease. Adv Exp Med Biol. 2007;595:1-75.

Aggarwal BB, Yuan W, Li S, Gupta SC. Curcumin-free turmeric exhibits anti-inflammatory and anticancer activities: Identification of novel components of turmeric. Mol Nutr Food Res. 2013;57:1529-1542.

Agrawal S, Millett CJ, Dhillon PK, Subramanian SV, EbrahimS. Type of vegetarian diet, obesity and diabetes in adult indian population. Nutr J. 2014;13:89.

Ahmed B, Tripathi K, Pandey ND, Khan M. Piperone-3 and piperone-4: Two new ketones isolated from *Piper longum* L. Dried fruits. Intl J Pharm Sci Rev Res. 2014;26:318-321.

Ahmed SH, Guillem K, Vandaele Y. Sugar addiction: Pushing the drug-sugar analogy to the limit. Curr Opin Clin Nutr Metab Care. 2013;16:434-439.

Ahmed T, Gilani AH. Therapeutic potential of turmeric in alzheimer's disease: Curcumin or curcuminoids?. Phytother Res. 2014;28:517-525.

Akinloye O, Ogunleye K, Oguntibeju OO. Cadmium, lead, arsenic and selenium levels in patients with type 2 diabetes mellitus. African J Biotechnol. 2013;9:5189-5195.

Alappat L, Awad AB. Curcumin and obesity: Evidence and mechanisms. Nutr Rev. 2010;68:729-738.

Ali RE, Rattan SI. Curcumin's biphasic hormetic response on proteasome activity and heat-shock protein synthesis in human keratinocytes. Ann N Y Acad Sci. 2006;1067:394-399.

Anand P, Kunnumakkara AB, Newman RA, Aggarwal BB. Bioavailability of curcumin: Problems and promises. Mol Pharm. 2007;4:807-818.

Anannarukan N, Niwattisaiwong N, Warisnoicharoen W, Winitthana T, Pramyothin P, Chaichantipyuth C, Lawanprasert S. Inhibition of human cytochrome p450 *in vitro* by phyllanthus amarus and *Phyllanthus emblica* aqueous extracts. Thai J Pharm Sci. 2012;36:135-143.

Anonymous. Position of the american dietetic association and dietitians of canada: Vegetarian diets. J Am Diet Assoc. 2003;103:748-765.

Aroniadis OC, Brandt LJ. Fecal microbiota transplantation: Past, present and future. Curr Opin Gastroenterol. 2013;29:79-84.

Asolkar LV, Kakkar KK, Chakre OJ. Glossary of indian medicinal plants with active principles, second supplement. (New Delhi, India; Publication and Information Directorate, CSIR), 1992.

Baboota RK, Bishnoi M, Ambalam P, Kondepudi KK, Sarma SM, Boparai RK, Podili K. Functional food ingredients for the management of obesity and associated co-morbidities - a review. J Funct Foods. 2013;5:997-1012.

Babu PS, Srinivasan K. Hypolipidemic action of curcumin, the active principle of turmeric (*Curcuma longa*) in streptozotocin induced diabetic rats. Mol Cell Biochem. 1997;166:169-175.

Balaji S, Chempakam B. Toxicity prediction of compounds from turmeric (*Curcuma longa* 1). Food Chem Toxicol. 2010;48:2951-2959.

Banks WA, Owen JB, Erickson MA. Insulin in the brain: There and back again. Pharmacol Ther. 2012;136:82-93.

Bao L, Bai S, Borijihan G. Hypolipidemic effects of a new piperine derivative gb-n from *Piper longum* in high-fat diet-fed rats. Pharm Biol. 2012;50:962-967.

Bao N, Ochir S, Sun Z, Borjihan G, Yamagishi T. Occurrence of piperidine alkaloids in piper species collected in different areas. J Nat Med. 2014;68:211-214.

Barrajon-Catalan E, Herranz-Lopez M, Joven J, Segura-Carretero A, Alonso-Villaverde C, Menendez JA, Micol V. Molecular promiscuity of plant polyphenols in the management of age-related diseases: Far beyond their antioxidant properties. Adv Exp Med Biol. 2014;824:141-159.

Baty V, Mougin B, Dekeuwer C, Carret G. Gut health in the era of the human gut microbiota: From metaphor to biovalue. Med Health Care Philos. 2014;17:579-597.

Belapurkar P, Goyal P, Tiwari-Barua P. Immunomodulatory effects of triphala and its individual constituents: A review. Indian J Pharm Sci. 2014;76:467-475.

Bezerra DP, Pessoa C, de Moraes MO, Saker-Neto N, Silveira ER, Costa-Lotufo LV. Overview of the therapeutic potential of piplartine (piperlongumine). Eur J Pharm Sci. 2013;48:453-463.

Bhanumathy M, Shivaprasad HN, Nargund LVG. Protective effect of *Curcuma longa* rhizomes against physical stress-induced perturbations in rats. J Nat Remedies. 2013;14:27-32.

Bharal N, Sahaya K, Jain S, Mediratta PK, Sharma KK. Curcumin has anticonvulsant activity on increasing current electroshock seizures in mice. Phytother Res. 2008;22:1660-1664.

Bhatia N, Jaggi A, Singh N, Anand P, Dhawan R. Adaptogenic potential of curcumin in experimental chronic stress and chronic unpredictable stress-induced memory deficits and alterations in functional homeostasis. J Nat Med. 2011;65:532-543.

Biradar YS, Jagatap S, Khandelwal KR, Singhania SS. Exploring of antimicrobial activity of triphala mashi-an ayurvedic formulation. Evid Based Complement Alternat Med. 2008;5:107-113.

Birringer M. Hormetics: Dietary triggers of an adaptive stress response. Pharm Res. 2011;28:2680-2694.

Bolca S, Van de Wiele T, Possemiers S. Gut metabotypes govern health effects of dietary polyphenols. Curr Opin Biotechnol. 2013;24:220-225.

Bradford PG. Curcumin and obesity. Biofactors. 2013;39:78-87.

BrahmaNaidu P, Nemani H, Meriga B, Mehar SK, Potana S, Ramgopalrao S. Mitigating efficacy of piperine in the physiological derangements of high fat diet induced obesity in sprague dawley rats. Chem Biol Interact. 2014;221:42-51.

Brekhman II, Dardymov IV. New substances of plant origin which increase nonspecific resistance. Annu Rev Pharmacol. 1969;9:419-430.

Brindley DN, Rolland Y. Possible connections between stress, diabetes, obesity, hypertension and altered lipoprotein metabolism that may result in atherosclerosis. Clin Sci (Lond). 1989;77:453-461.

Brodniewicz T, Grynkiewicz G. Plant phenolics as drug leads - what is missing? Acta Pol Pharm. 2012;69:1203-1217.

Burcelin R, Serino M, Chabo C, Blasco-Baque V, Amar J. Gut microbiota and diabetes: From pathogenesis to therapeutic perspective. Acta Diabetol. 2011;48:257-273.

Calabrese EJ. Hormesis is central to toxicology, pharmacology and risk assessment. Hum Exp Toxicol. 2010;29:249-261.

Calabrese V, Cornelius C, Dinkova-Kostova AT, Iavicoli I, Di Paola R, Koverech A, Cuzzocrea S, Rizzarelli E, Calabrese EJ. Cellular stress responses, hormetic phytochemicals and vitagenes in aging and longevity. Biochim Biophys Acta. 2012;1822:753-783.

Calabrese V, Cornelius C, Trovato A, Cavallaro M, Mancuso C, Di Rienzo L, Condorelli D, De Lorenzo A, Calabrese EJ. The hormetic role of dietary antioxidants in free radical-related diseases. Curr Pharm Des. 2010;16:877-883.

Carriba P, Comella JX. Amyloid beta, tnfalpha and faim-l; approaching new therapeutic strategies for ad. Front Neurol. 2014;5:276.

Cartea ME, Francisco M, Soengas P, Velasco P. Phenolic compounds in brassica vegetables. Molecules. 2010;16:251-280.

Carvalho LA, Urbanova L, Hamer M, Hackett RA, Lazzarino AI, Steptoe A. Blunted glucocorticoid and mineralocorticoid sensitivity to stress in people with diabetes. Psychoneuroendocrinology. 2015;51:209-218.

Chainani-Wu N. Safety and anti-inflammatory activity of curcumin: A component of tumeric (*Curcuma longa*). J Altern Complement Med. 2003;9:161-168.

Chandra P, Bajpai V, Srivastva M, Kumar KBR, Kumar B. Metabolic profiling of piper species by direct analysis using real time mass spectrometry combined with principal component analysis. Anal Methods. 2014;6:4234-4239.

Chandrasekaran C, Vijayalakshmi M, Prakash K, Bansal V, Meenakshi J, Amit A. Review article: Herbal approach for obesity management. Am J Plant Sci. 2012;3:1003-1014.

Chang CL, Chen YC, Chen HM, Yang NS, Yang WC. Natural cures for type 1 diabetes: A review of phytochemicals, biological actions, and clinical potential. Curr Med Chem. 2013a;20:899-907.

Chang CL, Lin Y, Bartolome AP, Chen YC, Chiu SC, Yang WC. Herbal therapies for type 2 diabetes mellitus: Chemistry, biology, and potential application of selected plants and compounds. Evid Based Complement Alternat Med. 2013b;2013:378657.

Chatterjee SS, Kumar V. Holistic psychopharmacology and promiscuous plants and principles of ayurveda. Am J Plant Sci. 2012;3:1015-1021.

Chaudhary SA, Gadhvi KV, Chaudhary AB. Comprehensive review on world herb trade and most utilized medicinal plant. Int J Appl Biol Pharm Tech. 2010;1:510-517.

Chaurasia A, Das D, From 5th World Ayurveda Congress Bhopal MPID. Pa01.07. Evaluation of anti hyperglycemic potential of *Piper longum* root (linn.) on alloxan induced diabetic mice. Anc Sci Life. 2012;32:S56.

Chaurasia A, Das D. Evaluation of antihyperglycemic potential of *Piper longum* root (linn.) on alloxan induced diabetic mice. Adv Pharmacol Toxicol. 2013;14:1-6.

Chempakam B, Parthasarathy VA. Turmeric. In Chemistry of spices. Parthasarathy VA, Chempakam B, Zachariah TJ eds. (Egham, UK; CAB International), pp. 97-123, 2008.

Cherng YG, Tsai CC, Chung HH, Lai YW, Kuo SC, Cheng JT. Antihyperglycemic action of sinapic acid in diabetic rats. J Agric Food Chem. 2013;61:12053-12059.

Chimakurthy J, Talasila M. Effects of curcumin on pentylenetetrazole-induced anxiety-like behaviors and associated changes in cognition and monoamine levels. Psychol Neurosci. 2010;3:239-244.

Chiou YS, Wu JC, Huang Q, Shahidi F, Wang YJ, Ho CT, Pan MH. Metabolic and colonic microbiota transformation may enhance the bioactivities of dietary polyphenols. J Func Foods. 2014;7:3-25.

Choi S, Choi Y, Choi Y, Kim S, Jang J, Park T. Piperine reverses high fat diet-induced hepatic steatosis and insulin resistance in mice. Food Chem. 2013;141:3627-3635.

Choudhary KM, Mishra A, Poroikov VV, Goel RK. Ameliorative effect of curcumin on seizure severity, depression like behavior, learning and memory deficit in postpentylenetetrazole-kindled mice. Eur J Pharmacol. 2013;704:33-40.

Choudhary M, Sangha J, Grover K. Conventional and nonconventional edible oils: An indian perspective. J Am Oil Chemists' Soc. 2014;91:179-206.

Chouhan B, Kumawat RC, Kotecha M, Ramamurthy A, Nathani S. Triphala: A comprehensive ayurvedic review. Int J Res Ayurveda Pharm. 2013;4:612-617.

Chulet R, Pradhan P. A review on rasayana. Pharmacogn Rev. 2009;3:229-234.

Cicero Bezerra Felipe F, Trajano Sousa Filho J, de Oliveira Souza LE, Alexandre Silveira J, Esdras de Andrade Uchoa D, Rocha Silveira E, Deusdenia Loiola Pessoa O, de Barros Viana GS. Piplartine, an amide alkaloid from piper tuberculatum, presents anxiolytic and antidepressant effects in mice. Phytomedicine. 2007;14:605-612.

Cnop M, Foufelle F, Velloso LA. Endoplasmic reticulum stress, obesity and diabetes. Trends Mol Med. 2012;18:59-68.

Colagiuri S. Diabesity: Therapeutic options. Diabetes Obes Metab. 2010;12:463-473.

Costantini D, Metcalfe NB, Monaghan P. Ecological processes in a hormetic framework. Ecol Lett. 2010;13:1435-1447.

Cryan JF, Dinan TG. Mind-altering microorganisms: The impact of the gut microbiota on brain and behaviour. Nat Rev Neurosci. 2012;13:701-712.

D'Souza JJ, D'Souza PP, Fazal F, Kumar A, Bhat HP, Baliga MS. Anti-diabetic effects of the indian indigenous fruit *Emblica officinalis* gaertn: Active constituents and modes of action. Food Funct. 2014;5:635-644.

D'Aversa F, Tortora A, Ianiro G, Ponziani F, Annicchiarico B, Gasbarrini A. Gut microbiota and metabolic syndrome. Intern Emerg Med. 2013;8:11-15.

Dai R, Lim LT. Release of allyl isothiocyanate from mustard seed meal powder. J Food Sci. 2014;79:E47-53.

Dallman MF, Pecoraro NC, la Fleur SE. Chronic stress and comfort foods: Self-medication and abdominal obesity. Brain Behav Immun. 2005;19:275-280.

Dasaroju S, Gottumukkala KM. Current trends in the research of *Emblica officinalis* (amla): A pharmacological perspective. Int J Pharm Sci Rev Res. 2014;24:150-159.

Davenport J, Balch M, Galam L, Girgis A, Hall J, Blagg BS, Matts RL. High-throughput screen of natural product libraries for hsp90 inhibitors. Biology (Basel). 2014;3:101-138.

Davinelli S, Sapere N, Zella D, Bracale R, Intrieri M, Scapagnini G. Pleiotropic protective effects of phytochemicals in alzheimer's disease. Oxid Med Cell Longev. 2012;2012:386527.

Davis C. Evolutionary and neuropsychological perspectives on addictive behaviors and addictive substances: Relevance to the "food addiction" construct. Subst Abuse Rehabil. 2014;5:129-137.

Dembinska-Kiec A, Mykkanen O, Kiec-Wilk B, Mykkanen H. Antioxidant phytochemicals against type 2 diabetes. Br J Nutr. 2008;99:ES109-ES117.

Dhanalakshmi S, Devi RS, Srikumar R, Manikandan S, Thangaraj R. Protective effect of triphala on cold stressinduced behavioral and biochemical abnormalities in rats. Yakugaku Zasshi. 2007;127:1863-1867.

Dhanalakshmi S, Srikumar R, Manikandan S, Parthasarathy NJ, Devi RS. Antioxidant property of triphala on cold stress induced oxidative stress in experimental rats. J Health Sci. 2006;52:843-847.

Dietrich MO, Horvath TL. Limitations in anti-obesity drug development: The critical role of hunger-promoting neurons. Nat Rev Drug Discov. 2012;11:675-691.

Dinan TG, Cryan JF. Regulation of the stress response by the gut microbiota: Implications for psychoneuroendocrinology. Psychoneuroendocrinology. 2012;37:1369-1378.

Dinkova-Kostova AT, Kostov RV. Glucosinolates and isothiocyanates in health and disease. Trends Mol Med. 2012;18:337-347.

Doucette CD, Hilchie AL, Liwski R, Hoskin DW. Piperine, a dietary phytochemical, inhibits angiogenesis. J Nutr Biochem. 2013;24:231-239.

Dragan S, Andrica F, Serban MC, Timar R. Polyphenols-rich natural products for treatment of diabetes. Curr Med Chem. 2015;22:14-22.

Drewnowski A, Gomez-Carneros C. Bitter taste, phytonutrients, and the consumer: A review. Am J Clin Nutr. 2000;72:1424-1435.

Eddouks M, Bidi A, El Bouhali B, Hajji L, Zeggwagh NA. Antidiabetic plants improving insulin sensitivity. J Pharm Pharmacol. 2014;66:1197-1214.

Epel ES, Lithgow GJ. Stress biology and aging mechanisms: Toward understanding the deep connection between adaptation to stress and longevity. J Gerontol A Biol Sci Med Sci. 2014;69:S10-S16.

Epel ES. Psychological and metabolic stress: A recipe for accelerated cellular aging? Hormones (Athens). 2009;8:7-22.

Everard A, Cani PD. Diabetes, obesity and gut microbiota. Best Pract Res Clin Gastroenterol. 2013;27:73-83.

Fahey JW, Wade KL, Stephenson KK, Chou FE. Separation and purification of glucosinolates from crude plant homogenates by high-speed counter-current chromatography. J Chromatogr A. 2003;996:85-93.

Farag YM, Gaballa MR. Diabesity: An overview of a rising epidemic. Nephrol Dial Transplant. 2011;26:28-35.

Farooqui AA. Effect of dietary phytochemicals on metabolic syndrome and neurological disorders. In Metabolic syndrome. (New York, USA; Springer), pp. 191-234, 2013.

Fasinu PS, Bouic PJ, Rosenkranz B. An overview of the evidence and mechanisms of herb-drug interactions. Front Pharmacol. 2012;3:69.

Fetrow CW, Avila JR. Professional's handbook of complementary and alternative medicines. (Pennsylvania, USA; Springhouse Corporation), 1999.

Fontana L. Modulating human aging and age-associated diseases. Biochim Biophys Acta. 2009;1790:1133-1138.

Foster JA, McVey Neufeld KA. Gut-brain axis: How the microbiome influences anxiety and depression. Trends Neurosci. 2013;36:305-312.

Franco R, Cedazo-Minguez A. Successful therapies for alzheimer's disease: Why so many in animal models and none in humans? Front Pharmacol. 2014;5:146.

Garcia-Alloza M, Borrelli LA, Rozkalne A, Hyman BT, Bacskai BJ. Curcumin labels amyloid pathology *in vivo*, disrupts existing plaques, and partially restores distorted neurites in an alzheimer mouse model. J Neurochem. 2007;102:1095-1104.

Gastaldi G, Ruiz J. Metabolic dysfunction and chronic stress: A new sight at "diabesity" pandemic. Rev Med Suisse. 2009;5:1273-1277.

Ghelardini C, Galeotti N, Di Cesare Mannelli L, Mazzanti G, Bartolini A. Local anaesthetic activity of beta-caryophyllene. Farmaco. 2001;56:387-389.

Gilhotra N, Dhingra D. Possible involvement of gabaergic and nitriergic systems for antianxiety-like activity of piperine in unstressed and stressed mice. Pharmacol Rep. 2014;66:885-891.

Ginty AT, Phillips AC, Higgs S, Heaney JL, Carroll D. Disordered eating behaviour is associated with blunted cortisol and cardiovascular reactions to acute psychological stress. Psychoneuroendocrinology. 2012;37:715-724.

Ginty AT. Blunted responses to stress and reward: Reflections on biological disengagement? Int J Psychophysiol. 2013;90:90-94.

Gonzalez-Castejon M, Rodriguez-Casado A. Dietary phytochemicals and their potential effects on obesity: A review. Pharmacol Res. 2011;64:438-455.

Goodacre R. Metabolomics of a superorganism. J Nutr. 2007;137:259S-266S.

Greiner AK, Papineni RV, Umar S. Chemoprevention in gastrointestinal physiology and disease. Natural products and

microbiome. Am J Physiol Gastrointest Liver Physiol. 2014;307:G1-15.

Grover JK, Yadav S, Vats V. Hypoglycemic and antihyperglycemic effect of *Brassica juncea* diet and their effect on hepatic glycogen content and the key enzymes of carbohydrate metabolism. Mol Cell Biochem. 2002;241:95-101.

Grover JK, Yadav SP, Vats V. Effect of feeding murraya koeingii and *Brassica juncea* diet on [correction] kidney functions and glucose levels in streptozotocin diabetic mice. J Ethnopharmacol. 2003;85:1-5.

Grynkiewicz G, Slifirski P. Curcumin and curcuminoids in quest for medicinal status. Acta Biochim Pol. 2012;59(2):201-212.

Gunthorpe MJ, Szallasi A. Peripheral trpv1 receptors as targets for drug development: New molecules and mechanisms. Curr Pharm Des. 2008;14:32-41.

Gupta PC. Biological and pharmacological properties of *Terminalia chebula* retz. (haritaki)-an overview. Int J Pharm Pharm Sci. 2012;4:62-68.

Gupta SC, Sung B, Kim JH, Prasad S, Li S, Aggarwal BB. Multitargeting by turmeric, the golden spice: From kitchen to clinic. Mol Nutr Food Res. 2013;57:1510-1528.

Halkier BA, Gershenzon J. Biology and biochemistry of glucosinolates. Annu Rev Plant Biol. 2006;57:303-333.

Hasan MN, Ferdoushi A, Ara N, Rahman S, Hossan MS, Rahmatullah M. Preliminary phytochemical screening, toxicity, antihyperglycemic and analgesic activity studies with *Curcuma longa* leaves. World J Pharm Pharm Sci. 2014;3:81-91.

Huang H, Morgan CM, Asolkar RN, Koivunen ME, Marrone PG. Phytotoxicity of sarmentine isolated from long pepper (*Piper longum*) fruit. J Agric Food Chem. 2010;58:9994-10000.

Inyang IJ, Eyo AAO, Olajide TM, Essien A. Effects of ethanolic extract of *Brassica juncea* (mustard seed) on the brain and kidney tissues of albino wistar rats. J Biol Agric Healthc. 2014;4:75-82.

Jakovljevic T, Cvjetko M, Sedak M, Dokic M, Bilandzic N, Vorkapic-Furac J, Redovnikovic IR. Balance of glucosinolates content under cd stress in two brassica species. Plant Physiol Biochem. 2013;63:99-106.

Janero DR. Relieving the cardiometabolic disease burden: A perspective on phytometabolite functional and chemical annotation for diabetes management. Expert Opin Pharmacother. 2014;15:5-10.

Jayaprakasha GK, Rao LJ, Sakariah KK. Antioxidant activities of curcumin, demethoxycurcumin and bisdemethoxycurcumin. Food Chem. 2006;98:720-724.

Jiang J, Wang W, Sun YJ, Hu M, Li F, Zhu DY. Neuroprotective effect of curcumin on focal cerebral ischemic rats by preventing blood-brain barrier damage. Eur J Pharmacol. 2007;561:54-62.

Jiang ZY, Liu WF, Huang CG, Huang XZ. New amide alkaloids from *Piper longum*. Fitoterapia. 2013;84:222-226.

Jin Z, Borjihan G, Zhao R, Sun Z, Hammond GB, Uryu T. Antihyperlipidemic compounds from the fruit of *Piper longum* L. Phytother Res. 2009;23:1194-1196.

Jo YS, Park JR, Park SK, Chun SS, Chung SY, Ha BS. Effects of mustard leaf (*Brassica juncea*) on cholesterol metabolism in rats. Korean J Nutr. 1993;26:13-20.

Joardar A, Das S. Effect of fatty acids isolated from edible oils like mustard, linseed or coconut on astrocytes maturation. Cell Mol Neurobiol. 2007;27:973-983.

John R, Ahmad P, Gadgil K, Sharma S. Heavy metal toxicity: Effect on plant growth, biochemical parameters and metal accumulation by *Brassica juncea* L. Int J Plant Prod. 2009;3:65-76.

Joshi J, Ghaisas S, Vaidya A, Vaidya R, Kamat DV, Bhagwat AN, Bhide S. Early human safety study of turmeric oil (*Curcuma longa* oil) administered orally in healthy volunteers. J Assoc Physicians India. 2003;51:1055-1060.

Joshi JV, Paradkar PH, Jagtap SS, Agashe SV, Soman G, Vaidya AB. Chemopreventive potential and safety profile of a *Curcuma longa* extract in women with cervical low-grade squamous intraepithelial neoplasia. Asian Pac J Cancer Prev. 2011;12:3305-3311.

Joven J, Rull A, Rodriguez-Gallego E, Camps J, Riera-Borrull M, Hernandez-Aguilera A, Martin-Paredero V, Segura-Carretero A, Micol V, Alonso-Villaverde C, Menendez JA. Multifunctional targets of dietary polyphenols in disease: A case for the chemokine network and energy metabolism. Food Chem Toxicol. 2013;51:267-279.

Jung HA, Woo JJ, Jung MJ, Hwang GS, Choi JS. Kaempferol glycosides with antioxidant activity from *Brassica juncea*. Arch Pharm Res. 2009;32:1379-1384.

Kadian R, Parle M, Yadav M. Therapeutic potential and phytopharmacology of *Terminalia bellerica*. World J Pharm Pharm Sci. 2014;3:804-819.

Kahn SE, Cooper ME, Del Prato S. Pathophysiology and treatment of type 2 diabetes: Perspectives on the past, present, and future. Lancet. 2014;383:1068-1083.

Kandarkar SV, Sawant SS, Ingle AD, Deshpande SS, Maru GB. Subchronic oral hepatotoxicity of turmeric in mice-histopathological and ultrastructural studies. Indian J Exp Biol. 1998;36:675-679.

Kar S, Roy K. Qsar of phytochemicals for the design of better drugs. Expert Opin Drug Discov. 2012;7:877-902.

Karakida F, Ikeya Y, Tsunakawa M, Yamaguchi T, Ikarashi Y, Takeda S, Aburada M. Cerebral protective and cognitionimproving effects of sinapic acid in rodents. Biol Pharm Bull. 2007;30:514-519.

Kasabri V, Flatt PR, Abdel-Wahab YHA. *In vitro* modulation of pancreatic insulin secretion, extrapancreatic insulin action and peptide glycation by *Curcuma longa* aqueous extracts. J Exp Integr Med. 2014;4:187-193.

Kelly SJ, Ismail M. Stress and type 2 diabetes: A review of

how stress contributes to the development of type 2 diabetes. Annu Rev Public Health. 2015;36:441-462.

Kennedy DO. Plants and the human brain. (New York, USA; Oxford University Press), 2014a.

Kennedy DO. Polyphenols and the human brain: Plant "secondary metabolite" ecologic roles and endogenous signaling functions drive benefits. Adv Nutr. 2014b;5:515-533.

Khadem S, Marles RJ. Monocyclic phenolic acids; hydroxyand polyhydroxybenzoic acids: Occurrence and recent bioactivity studies. Molecules. 2010;15:7985-8005.

Khan BA, Abraham A, Leelamma S. Anti-oxidant effects of curry leaf, murraya koenigii and mustard seeds, *Brassica juncea* in rats fed with high fat diet. Indian J Exp Biol. 1997;35:148-150.

Khan BA, Abraham A, Leelamma S. Hypoglycemic action of murraya koenigii (curry leaf) and *Brassica juncea* (mustard): Mechanism of action. Indian J Biochem Biophys. 1995;32:106-108.

Khan BA, Abraham A, Leelamma S. Murraya koenigii and *Brassica juncea*-alterations on lipid profile in 1-2 dimethyl hydrazine induced colon carcinogenesis. Invest New Drugs. 1996;14:365-369.

Khosla S, Sharma S. A short description on pharmacogenetic properties of *Emblica officinalis*. Spatula DD. 2012;2:187-193.

Kiasalari Z, Roghani M, Khalili M, Rahmati B, Baluchnejadmojarad T. Antiepileptogenic effect of curcumin on kainate-induced model of temporal lobe epilepsy. Pharm Biol. 2013;51:1572-1578.

Kim DSHL, Park S-Y, Kim J-Y. Curcuminoids from *Curcuma* longa l. (zingiberaceae) that protect pc12 rat pheochromocytoma and normal human umbilical vein endothelial cells from $\beta a(1-42)$ insult. Neurosci Lett. 2001;303:57-61.

Kim HY, Yokozawa T, Cho EJ, Cheigh HS, Choi JS, Chung HY. *In vitro* and *in vivo* antioxidant effects of mustard leaf (*Brassica juncea*). Phytother Res. 2003;17:465-471.

Kim JE, Jung MJ, Jung HA, Woo JJ, Cheigh HS, Chung HY, Choi JS. A new kaempferol 7-o-triglucoside from the leaves of *Brassica juncea* L. Arch Pharm Res. 2002;25:621-624.

Kokate CK, Purohit AP, Gokhale SB. Practical Pharmacognosy, Twenty second edition. (Pune, India: Nirali Prakashan), 2003.

Krishna M, Joy B, Sundaresan A. Effect on oxidative stress, glucose uptake level and lipid droplet content by apigenin 7, 4'-dimethyl ether isolated from *Piper longum* L. J Food Sci Tech. 2014:1-10.

Krishnaveni M, Mirunalini S. Therapeutic potential of *Phyllanthus emblica* (amla): The ayurvedic wonder. J Basic Clin Physiol Pharmacol. 2010;21:93-105.

Ku SK, Kim JA, Bae JS. Vascular barrier protective effects of piperlonguminine *in vitro* and *in vivo*. Inflamm Res. 2014;63:369-379.

Kulkarni S, Dhir A, Akula KK. Potentials of curcumin as an antidepressant. ScientificWorldJournal. 2009;9:1233-1241.

Kumar A, Panghal S, Mallapur SS, Kumar M, Ram V, Singh BK. Antiinflammatory activity of *Piper longum* fruit oil. Indian J Pharm Sci. 2009;71:454-456.

Kumar S, Andy A. Health promoting bioactive phytochemicals from brassica. Int Food Res J. 2012;19:141-152.

Kumar S, Kamboj J, Suman, Sharma S. Overview for various aspects of the health benefits of *Piper longum* Linn. Fruit. J Acupunct Meridian Stud. 2011b;4:134-140.

Kumar S, Sharma S, Vasudeva N. Screening of antidiabetic and antihyperlipidemic potential of oil from *Piper longum* and piperine with their possible mechanism. Expert Opin Pharmacother. 2013;14:1723-1736.

Kumar V, Chatterjee SS. A holistic approach for evaluating the potential of triphala extract against neurological disorders. PharmaNutrition. 2014b;2:90.

Kumar V, Chatterjee SS. Single and repeated dose effects of phytochemicals in rodent behavioural models. EC Pharm Sci. 2014a;1:16-18.

Kumar V, Thakur A, Chatterjee S. Obesity, cancer and psychopathology: Can vegetarian diet be of help? In Nutrition, diet and cancer. Shankar S, Srivastava RK eds. (Houten, Netherlands; springer), pp. 459-491, 2012.

Kumar V, Thakur AK, Barothia ND, Chatterjee SS. Therapeutic potentials of *Brassica juncea*: An overview. TANG (Humanitas Medicine). 2011a;1:1-17.

Kumudhavalli MV, Mohit V, Jayakar B. Phytochemical and pharmacological evaluation of the plant fruit of *Terminalia belerica* roxb. Int J Pharm Life Sci. 2010;1:1-11.

Kuo CC, Moon K, Thayer KA, Navas-Acien A. Environmental chemicals and type 2 diabetes: An updated systematic review of the epidemiologic evidence. Curr Diab Rep. 2013;13:831-849.

Kuo CP, Lu CH, Wen LL, Cherng CH, Wong CS, Borel CO, Ju DT, Chen CM, Wu CT. Neuroprotective effect of curcumin in an experimental rat model of subarachnoid hemorrhage. Anesthesiology. 2011;115:1229-1238.

Kuroda M, Mimaki Y, Nishiyama T, Mae T, Kishida H, Tsukagawa M, Takahashi K, Kawada T, Nakagawa K, Kitahara M. Hypoglycemic effects of turmeric (*Curcuma longa* l. Rhizomes) on genetically diabetic kk-ay mice. Biol Pharm Bull. 2005;28:937-939.

Kwon DY, Kim HJ, Yoon SH. Induction of phenolics and terpenoids in edible plants using plant stress responses. In Biocatalysis and agricultural biotechnology. Hou CT, Shaw JF eds. (New York, USA; CRC Press), pp. 249–258, 2009.

Langstieh AJ, Verma P, Thakur AK, Chatterjee SS, Kumar V. Desensitization of mild stress triggered responses in mice by a *Brassica juncea* Leaf extract and some ubiquitous secondary plant metabolites. Pharmacologia. 2014;5:326-338.

Lao CD, Ruffin MTt, Normolle D, Heath DD, Murray SI,

Bailey JM, Boggs ME, Crowell J, Rock CL, Brenner DE. Dose escalation of a curcuminoid formulation. BMC Complement Altern Med. 2006;6:10.

Le Bourg E, Rattan SI. Hormesis and trade-offs: A comment. Dose Response. 2014;12:522-524.

Lee HE, Kim DH, Park SJ, Kim JM, Lee YW, Jung JM, Lee CH, Hong JG, Liu X, Cai M, Park KJ, Jang DS, Ryu JH. Neuroprotective effect of sinapic acid in a mouse model of amyloid beta(1-42) protein-induced alzheimer's disease. Pharmacol Biochem Behav. 2012;103:260-266.

Lee J, Jo DG, Park D, Chung HY, Mattson MP. Adaptive cellular stress pathways as therapeutic targets of dietary phytochemicals: Focus on the nervous system. Pharmacol Rev. 2014;66:815-868.

Lee SA, Hwang JS, Han XH, Lee C, Lee MH, Choe SG, Hong SS, Lee D, Lee MK, Hwang BY. Methylpiperate derivatives from *Piper longum* and their inhibition of monoamine oxidase. Arch Pharm Res. 2008;31:679-683.

Lee SW, Rho MC, Park HR, Choi JH, Kang JY, Lee JW, Kim K, Lee HS, Kim YK. Inhibition of diacylglycerol acyltransferase by alkamides isolated from the fruits of *Piper longum* and piper nigrum. J Agric Food Chem. 2006;54:9759-9763.

Leiherer A, Mundlein A, Drexel H. Phytochemicals and their impact on adipose tissue inflammation and diabetes. Vascul Pharmacol. 2013;58:3-20.

Lekshmi PC, Arimboor R, Indulekha PS, Menon AN. Turmeric (*Curcuma longa* l.) volatile oil inhibits key enzymes linked to type 2 diabetes. Int J Food Sci Nutr. 2012;63:832-834.

Lephart ED. Polyphenols and cognitive function. In Diet and exercise in cognitive function and neurological diseases. Farooqui T, Farooqui AA eds. (Hoboken, New Jersey, USA; John Wiley & Sons, Inc), pp. 143-161, 2015.

Li J, Ho C-T, Li HE, Tao H, Tao L. Separation of sterols and triterpene alcohols from unsaponifiable fractions of three plant seed oils. J Food Lipids. 2000;7:11-20.

Li S, Wang C, Li W, Koike K, Nikaido T, Wang MW. Antidepressant-like effects of piperine and its derivative, antiepilepsirine. J Asian Nat Prod Res. 2007a;9:421-430.

Li S, Wang C, Wang M, Li W, Matsumoto K, Tang Y. Antidepressant like effects of piperine in chronic mild stress treated mice and its possible mechanisms. Life Sci. 2007b;80(15):1373-1381.

Li S, Yuan W, Deng G, Wang P, Yang P, Aggarwal BB. Chemical composition and product quality control of turmeric (*Curcuma longa* L.). Phytochemistry. 2011;2:28-54.

Liju VB, Jeena K, Kuttan R. Acute and subchronic toxicity as well as mutagenic evaluation of essential oil from turmeric (*Curcuma longa* L). Food Chem Toxicol. 2013;53:52-61.

Liju VB, Jeena K, Kuttan R. An evaluation of antioxidant, antiinflammatory, and antinociceptive activities of essential oil from *Curcuma longa*. L. Indian J Pharmacol. 2011;4:526-531. Lin Y, Sun Z. Current views on type 2 diabetes. J Endocrinol. 2010;204:1-11.

Liu J-Y, Chen X-X, Tang SC-W, Lao L-X, Cho-Wing Sze S, Lee K-F, Zhang KY-B. Edible plants from traditional chinese medicine is a promising alternative for the management of diabetic nephropathy. J Func Foods. 2015;14:12-22.

Liu Z, Yu Y, Li X, Ross CA, Smith WW. Curcumin protects against a53t alpha-synuclein-induced toxicity in a pc12 inducible cell model for parkinsonism. Pharmacol Res. 2011;63:439-444.

Lopresti AL, Maes M, Maker GL, Hood SD, Drummond PD. Curcumin for the treatment of major depression: A randomised, double-blind, placebo controlled study. J Affect Disord. 2014;167:368-375.

Lushchak VI. Dissection of the hormetic curve: Analysis of components and mechanisms. Dose Response. 2014a;12:466-479.

Lushchak VI. Hormesis in biology and pharmacology. Biochem Pharmacol. 2014b;3:1-2.

Madhu K, Chanda K, Saji MJ. Safety and efficacy of *Curcuma longa* extract in the treatment of painful knee osteoarthritis: A randomized placebo-controlled trial. Inflammopharmacology. 2013;21:129-136.

Maheshwari M. Comparative bioavailability of curcumin, turmeric and biocurcumaxTM in traditional vehicles using noneverted rat intestinal sac model. J Func Foods. 2010;2:60-65.

Maheshwari RK, Rajnee K. Ingenious triphala: A curative preparation for health care. Asian J Chem Pharm Res. 2014;2:175 -185.

Manach C, Hubert J, Llorach R, Scalbert A. The complex links between dietary phytochemicals and human health deciphered by metabolomics. Mol Nutr Food Res. 2009;53:1303-1315.

Manach C, Scalbert A, Morand C, Remesy C, Jimenez L. Polyphenols: Food sources and bioavailability. Am J Clin Nutr. 2004;79:727-747.

Maneesa P, Scholfield CN, Chootip K. Piperine is antihyperlipidemic and improves endothelium-dependent vasorelaxation in rats on a high cholesterol diet. J Physiol Biomed Sci. 2012;25:27-30.

Manesh C, Kuttan G. Anti-tumour and anti-oxidant activity of naturally occurring isothiocyanates. J Exp Clin Cancer Res. 2003;22:193-199.

Manohar PR, Pushpan R, Rohini S. Mustard and its uses in ayurveda. Indian J Tradit Knowl. 2009;8:400-404.

Manoj P, Soniya EV, Banerjee NS, Ravichandran P. Recent studies on well-known spice, *Piper longum* Linn. Nat Prod Radiance. 2004;3:222-227.

Mao QQ, Huang Z, Zhong XM, Xian YF, Ip SP. Brain-derived neurotrophic factor signalling mediates the antidepressant-like effect of piperine in chronically stressed mice. Behav Brain Res. 2014;261:140-145.

Martinez JA, Milagro FI. Genetics of weight loss: A basis for personalized obesity management. Trends Food Sci Tech. 2015;42:97-115.

Martinez JA, Navas-Carretero S, Saris WH, Astrup A. Personalized weight loss strategies-the role of macronutrient distribution. Nat Rev Endocrinol. 2014;10:749-760.

Mayer EA. Gut feelings: The emerging biology of gut-brain communication. Nat Rev Neurosci. 2011;12:453-466.

McClure CD, Zhong W, Hunt VL, Chapman FM, Hill FV, Priest NK. Hormesis results in trade-offs with immunity. Evolution. 2014;68:2225-2233.

McFadden RMT, Larmonier CB, Midura-Kiela MT, Ramalingam R, Harrison CA, Besselsen DG, Chase J, Caporaso G, Ghishan FK, Kiela PR. The role of curcumin in modulating colonic microbiota during colitis and colon cancer prevention. Gastroenterology. 2014;146:S66.

McNamara FN, Randall A, Gunthorpe MJ. Effects of piperine, the pungent component of black pepper, at the human vanilloid receptor (trpv1). Br J Pharmacol. 2005;144:781-790.

Meghwal M, Goswami TK. Piper nigrum and piperine: An update. Phytother Res. 2013;27:1121-1130.

Meule A, Heckel D, Jurowich CF, Vogele C, Kubler A. Correlates of food addiction in obese individuals seeking bariatric surgery. Clin Obes. 2014;4:228-236.

Micucci M, Aldini R, Cevenini M, Colliva C, Spinozzi S, Roda G, Montagnani M, Camborata C, Camarda L, Chiarini A, Mazzella G, Budriesi R. *Curcuma longa* I. As a therapeutic agent in intestinal motility disorders. 2: Safety profile in mouse. PLoS One. 2013;8:e80925.

Mirmiran P, Bahadoran Z, Azizi F. Functional foods-based diet as a novel dietary approach for management of type 2 diabetes and its complications: A review. World J Diabetes. 2014;5:267-281.

Mishra S, Manchanda SC. Cooking oils for heart health. J Prev Cardiol. 2012;1:123-131.

Mohammad K, Larijani B. A systematic review of the antioxidant, anti-diabetic, and anti-obesity effects and safety of triphala herbal formulation. J Med Plants Res. 2013;7:831-844.

Moloney RD, Desbonnet L, Clarke G, Dinan TG, Cryan JF. The microbiome: Stress, health and disease. Mamm Genome. 2014;25:49-74.

Mori A, Kabuto H, Pei YQ. Effects of piperine on convulsions and on brain serotonin and catecholamine levels in e1 mice. Neurochem Res. 1985;10:1269-1275.

Munday R, Munday CM. Selective induction of phase ii enzymes in the urinary bladder of rats by allyl isothiocyanate, a compound derived from brassica vegetables. Nutr Cancer. 2002;44:52-59.

Murray S, Kroll C, Avena NM. Food and addiction among the ageing population. Ageing Res Rev. 2015;20:79-85.

Murthy KRS. Bhava prakasha of bhavamisra-english

translation. (Varanasi, India; Chowkhamba Krishnadas Academy), 2009.

Muttepawar SS, Jadhav SB, Kankudate AD, Sanghai SD, Usturge DR, Chavare SS. A review on bioavailability enhancers of herbal origin. World J Pharm Pharm Sci. 2014;3:667-677.

Nabi SA, Kasetti RB, Sirasanagandla S, Tilak TK, Kumar MV, Rao CA. Antidiabetic and antihyperlipidemic activity of *Piper longum* root aqueous extract in STZ induced diabetic rats. BMC Complement Altern Med. 2013;13:37.

Nariya MB, Shukla VJ, Ravishankar B, Jain SM. Comparison of gastroprotective effects of triphala formulations on stressinduced ulcer in rats. Indian J Pharm Sci. 2011;73:682-687.

Neerati P, Ravi Karan M, Kanwar JR. Influence of curcumin on pioglitazone metabolism and pk/pd: Diabetes mellitus. J Diabetes Metab. 2012;S6:1-6.

Neha J, Mishra RN. Adaptogenic activity of Trikatu megaExt. Int J Res Pharm Biomed Sci. 2011;2:570-574.

Nicholson JK, Holmes E, Kinross J, Burcelin R, Gibson G, Jia W, Pettersson S. Host-gut microbiota metabolic interactions. Science. 2012;336:1262-1267.

Niciforovic N, Abramovic H. Sinapic acid and its derivatives: Natural sources and bioactivity. Compr Rev Food Sci Food Saf. 2014;13:34-51.

Noble T, Zingg JM, Paul L, Smith D, Meydani M. The effect of curcumin plus piperine on body weight and fat loss as well as on the plasma levels of inflammatory cytokines in obese mice. FASEB J. 2013;27:636.

Nowak D. Antioxidant plant polyphenols and cognitive disorders. In Studies on psychiatric disorders. Dietrich-Muszalska A, Chauhan V, Grignon S eds. (New York, USA; Springer), pp. 521-552, 2015.

Nyanginja RA, Mponda J. Nutrigenomic approach in understanding the antiallergic effects of curcumin. Asian J Biomed Pharma Sci. 2014;4:1-5.

Ono K, Hasegawa K, Naiki H, Yamada M. Curcumin has potent anti-amyloidogenic effects for alzheimer's beta-amyloid fibrils *in vitro*. J Neurosci Res. 2004;75:742-750.

Oram RN, Kirk JTO, Veness PE, Hurlstone CJ, Edlington JP, Halsall DM. Breeding indian mustard [*Brassica juncea* (L.) czern.] for cold-pressed, edible oil production-a review. Australian J Agric Res. 2005;56:581-596.

Ozcan U, Cao Q, Yilmaz E, Lee AH, Iwakoshi NN, Ozdelen E, Tuncman G, Gorgun C, Glimcher LH, Hotamisligil GS. Endoplasmic reticulum stress links obesity, insulin action, and type 2 diabetes. Science. 2004;306:457-461.

Pa J, Gazzaley A. Flavanol-rich food for thought. Nat Neurosci. 2014;17:1624-1625.

Pal A, Nayak S, Sahu PK, Swain T. Piperine protects epilepsy associated depression: A study on role of monoamines. Eur Rev Med Pharmacol Sci. 2011;15:1288-1295.

Pal US, Khan K, Sahoo NR, Sahoo G. Development and evaluation of farm level turmeric processing equipment. AMA. 2008;39:46-50.

Pan XR, Li GW, Hu YH, Wang JX, Yang WY, An ZX, Hu ZX, Lin J, Xiao JZ, Cao HB, Liu PA, Jiang XG, Jiang YY, Wang JP, Zheng H, Zhang H, Bennett PH, Howard BV. Effects of diet and exercise in preventing niddm in people with impaired glucose tolerance. The da qing igt and diabetes study. Diabetes Care. 1997;20:537-544.

Pandey KB, Rizvi SI. Plant polyphenols as dietary antioxidants in human health and disease. Oxid Med Cell Longev. 2009;2:270-278.

Parrott MD, Winocur G, Bazinet RP, Ma DW, Greenwood CE. Whole-food diet worsened cognitive dysfunction in an alzheimer's disease mouse model. Neurobiol Aging. 2015;36:90-99.

Patel D, Patel K, Gadewar M, Tahilyani V. A concise report on pharmacological and bioanalytical aspect of sinigrin. Asian Pac J Trop Biomed. 2012;2:S446-S448.

Patel JA, Shah US. Hepatoprotective activity of *Piper longum* traditional milk extract on carbon tetrachloride induced liver toxicity in wistar rats. Bolet ín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas. 2009;8:121-129.

Patel P, Gohil KJ, Roy SP, Patel N. Investigation of antidepressant and anxiolytic activity of curcumin given alone and in combination with amitriptyline in rats. Indian J Res Pharm Biotechnol. 2014;2:1173-1178.

Pathak N, Khandelwal S. Immunomodulatory role of piperine in cadmium induced thymic atrophy and splenomegaly in mice. Environ Toxicol Pharmacol. 2009;28:52-60.

Pati S, Hussain M, Swain S. Characteristics and correlates of diabesity in india: A secondary data analysis of world health survey. Obes Rev. 2014;15:67-68.

Patil UK, Singh A, Chakraborty AK. Role of piperine as a bioavailability enhancer. Int J Recent Adv Pharm Res. 2011;4:16-23.

Patrone C, Eriksson O, Lindholm D. Diabetes drugs and neurological disorders: New views and therapeutic possibilities. Lancet Diabetes Endocrinol. 2014;2:256-262.

Pedram P, Sun G. Hormonal and dietary characteristics in obese human subjects with and without food addiction. Nutrients. 2015;7:223-238.

Peng D, Xin L, Hao-jie L, Wei-feng P, Jian-ying L, Yu M, Wei F, Xin W. Curcumin inhibits amygdaloid kindled seizures in rats. Chin Med J. 2009;122:1435-1438.

Perez-Tilve D, Davidson WS, Tschop M, Hofmann SM. Cns regulation of plasma cholesterol. Ann Med. 2012;44:656-663.

Podar D, Ramsey MH, Hutchings MJ. Effect of cadmium, zinc and substrate heterogeneity on yield, shoot metal concentration and metal uptake by *Brassica juncea*: Implications for human health risk assessment and phytoremediation. New Phytologist. 2004;163:313-324.

Ponnusamy S, Zinjarde S, Bhargava S, Kumar AR. Role of *Curcuma longa*, a traditional ayurvedic medicinal plant, in diabetes. TANG (Humanitas Medicine). 2012;2:5-11.

Ponnusankar S, Pandit S, Babu R, Bandyopadhyay A, Mukherjee PK. Cytochrome p450 inhibitory potential of triphala-a rasayana from ayurveda. J Ethnopharmacol. 2011;133:120-125.

Pratley RE, Matfin G. Pre-diabetes: Clinical relevance and therapeutic approach. Br J Diabetes Vasc Dis. 2007;7:120–129.

Priyadarsini KI. The chemistry of curcumin: From extraction to therapeutic agent. Molecules. 2014;19:20091-20112.

Puranik A, Nabar N, Joshi J, Amonkar A, Shah S, Menon S, Vaidya R, Vaidya ADB. Single dose metformin kinetics after co-administration of nisha-amalaki powder or mamejwa ghanavati, ayurvedic anti-diabetic formulations: A randomized crossover study in healthy volunteers. J Obes Metab Res. 2014;1:99-104.

Qin XY, Cheng Y, Yu LC. Potential protection of curcumin against intracellular amyloid beta-induced toxicity in cultured rat prefrontal cortical neurons. Neurosci Lett. 2010;480:21-24.

Qureshi S, Shah AH, Ageel AM. Toxicity studies on alpinia galanga and *Curcuma longa*. Planta Med. 1992;58:124-127.

Rajan SS, Antony S. Hypoglycemic effect of triphala on selected non insulin dependent diabetes mellitus subjects. Anc Sci Life. 2008;27:45-49.

Rajeswary H, Vasuki R, Samudram P, Geetha A. Hepatoprotective action of ethanolic extracts of melia azedarach linn. And *Piper longum* Linn and their combination on ccl4 induced hepatotoxicity in rats. Indian J Exp Biol. 2011;49:276-281.

Rajopadhye AA, Namjoshi TP, Upadhye AS. Rapid validated hptlc method for estimation of piperine and piperlongumine in root of *Piper longum* extract and its commercial formulation. Revista Brasileira de Farmacognosia. 2012;22:1355-1361.

Ramachandran A, Snehalatha C, Mary S, Mukesh B, Bhaskar AD, Vijay V, Indian Diabetes Prevention P. The indian diabetes prevention programme shows that lifestyle modification and metformin prevent type 2 diabetes in asian indian subjects with impaired glucose tolerance (idpp-1). Diabetologia. 2006;49:289-297.

Ramachandran A, Snehalatha C, Shetty AS, Nanditha A. Trends in prevalence of diabetes in asian countries. World J Diabetes. 2012;3:110-117.

Rao G, Bhat S, Rao GS, Bhat GP. Antidiabetic and antioxidant efficacy of a powdered mixture of *Curcuma longa* and *Emblica officinalis* in diabetic rats in comparison with glyburide. WebmedCentral Diabetes. 2013;4:1-13.

Rastogi RP, Mehrotra BN. Compendium of Indian medicinal plants, vol. 4. (New Delhi, India; Publication and Information Directorate, CSIR), 1995.

Rastogi S. Ayurvedic science of food and nutrition. (New York, USA; Springer), 2014.

Rastogi T, Reddy KS, Vaz M, Spiegelman D, Prabhakaran D, Willett WC, Stampfer MJ, Ascherio A. Diet and risk of ischemic heart disease in india. Am J Clin Nutr. 2004;79:582-592.

Rathinamoorthy R, Thilagavathi G. *Terminalia chebula*-review on pharmacological and biochemical studies. Int J PharmTech Res. 2014;6:97-116.

Rauniyar BK, Shakya A, Thakur AK, Chatterjee SS, Kumar V. Anti-stress activity of phloroglucinol: A transient metabolite of some plant polyphenolics. Pharmacologia. 2015;6:21-30.

Rege NN, Thatte UM, Dahanukar SA. Adaptogenic properties of six rasayana herbs used in ayurvedic medicine. Phytother Res. 1999;13:275-291.

Rhee SH, Pothoulakis C, Mayer EA. Principles and clinical implications of the brain-gut-enteric microbiota axis. Nat Rev Gastroenterol Hepatol. 2009;6:306-314.

Rinki S, Mishra RN. Adaptogenic activity of triphala megaext. Int J Res Pharm Biomed Sci. 2011;2:579-582.

Rios MY, Olivo HF. Chapter 3 - natural and synthetic alkamides: Applications in pain therapy. In Studies in natural products chemistry. Atta ur R ^{ed}. (Oxford, UK: Elsevier), pp. 79-121, 2014.

Roy SJ, Stanely Mainzen Prince P. Protective effects of sinapic acid on lysosomal dysfunction in isoproterenol induced myocardial infarcted rats. Food Chem Toxicol. 2012;50:3984-3989.

Ruden D, Lu X. Personalized nutrigenomics: Tailoring the diet to the aging diabesity population. Nutr Diet Suppl. 2011;3:31-41.

Sahebkar A. Why it is necessary to translate curcumin into clinical practice for the prevention and treatment of metabolic syndrome? Biofactors. 2013;39:197-208.

Saleem A, Husheem M, Harkonen P, Pihlaja K. Inhibition of cancer cell growth by crude extract and the phenolics of *Terminalia chebula* retz. Fruit. J Ethnopharmacol. 2002;81:327-336.

Samudram P, Vasuki, R., Rajeshwari, H., Geetha, A., Moorthi, P.S. Antioxidant and antihepatotoxic activities of ethanolic crude extract of melia azedarach and *Piper longum*. J Med Plants Res. 2009;3:1078-1083.

Sandoval DA, Obici S, Seeley RJ. Targeting the cns to treat type 2 diabetes. Nat Rev Drug Discov. 2009;8:386-398.

Sang JP, Minchinton IR, Johnstone PK, Truscott RJ. Glucosinolates profiles in the seed, root and leaf tissue of cabbage, mustard, rapeseed, radish and swede. Can J Plant Sci. 1984;64:77-93.

Sanmukhani J, Anovadiya A, Tripathi CB. Evaluation of antidepressant like activity of curcumin and its combination with fluoxetine and imipramine: An acute and chronic study. Acta Pol Pharm. 2011;68:769-775.

Santilli F, Guagnano MT, Vazzana N, La Barba S, Davi G. Oxidative stress drivers and modulators in obesity and

cardiovascular disease: From biomarkers to therapeutic approach. Curr Med Chem. 2015;22:582-595.

Sarma GK, Kalita P, Barthakur M, Sarma RK. Importance of traditionally used medicinal plants: *Emblica officinalis* (L), *Curcuma longa* (L) & *Syzygium cumini* (L) in the management of diabetes mellitus. Int Ayurvedic Med J. 2014;2:356-361.

Sauer FD, Kramer JKG. The problems associated with the feeding of high erucic acid rapeseed oils and some fish oils to experimental animals. In High and low erucic acid rapeseed oils production, usage, chemistry, and toxicological examination. Kramer JKG, Sauer FD, Pigden WJ eds. (Toronto, Canada; Academic Press), pp. 253–292, 1983.

Saxena V, Mishra G, Vishwakarma KK, Saxena A. A comparative study on quantitative estimation of tannins in *Terminalia chebula*, *Terminalia belerica*, *Terminalia arjuna* and saraca indica using spectrophotometer. Asian J Pharm Clin Res. 2013;6:1-2.

Scapagnini G, Davinelli S, Kaneko T, Koverech G, Koverech A, Calabrese EJ, Calabrese V. Dose response biology of resveratrol in obesity. J Cell Commun Signal. 2014;8:385-391.

Schreiner M, Krumbein A, Ruppel S. Interaction between plants and bacteria: Glucosinolates and phyllospheric colonization of cruciferous vegetables by enterobacter radicincitans dsm 16656. J Mol Microbiol Biotechnol. 2009;17:124-135.

Scott C, Johnstone AM. Stress and eating behaviour: Implications for obesity. Obes Facts. 2012;5:277-287.

Serrano J, Puupponen-Pimia R, Dauer A, Aura AM, Saura-Calixto F. Tannins: Current knowledge of food sources, intake, bioavailability and biological effects. Mol Nutr Food Res. 2009;53:S310-329.

Shakya A, Chatterjee SS, Kumar V. Role of fumarates in adaptogenics like efficacies of traditionally used fumaria indica extracts. TANG (Humanitas Medicine). 2015;5:28-37

Sharma H, Chandola HM. Prameha in ayurveda: Correlation with obesity, metabolic syndrome, and diabetes mellitus. Part 1-etiology, classification, and pathogenesis. J Altern Complement Med. 2011a;17:491-496.

Sharma H, Chandola HM. Prameha in ayurveda: Correlation with obesity, metabolic syndrome, and diabetes mellitus. Part 2-management of prameha. J Altern Complement Med. 2011b;17:589-599.

Shin DS, Kim KW, Chung HY, Yoon S, Moon JO. Effect of sinapic acid against dimethylnitrosamine-induced hepatic fibrosis in rats. Arch Pharm Res. 2013;36:608-618.

Shin HJ, Lee JY, Son E, Lee DH, Kim HJ, Kang SS, Cho GJ, Choi WS, Roh GS. Curcumin attenuates the kainic acidinduced hippocampal cell death in the mice. Neuroscience Letters. 2007;416:49-54.

Shivavedi N, Chatterjee SS, Kumar V. Evaluation of pharmacologically interesting dose range of ascorbic acid in mice. SAJ Neurol. 2014a;1:101.

Shivavedi N, Chatterjee SS, Kumar V. Stress response

modulating effects of lactic acid in mice. Ther Targets Neurol Dis. 2014b;1:e418.

Shoba G, Joy D, Joseph T, Majeed M, Rajendran R, Srinivas PS. Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. Planta Med. 1998;64:353-356.

Sims EA, Danforth E, Jr., Horton ES, Bray GA, Glennon JA, Salans LB. Endocrine and metabolic effects of experimental obesity in man. Recent Prog Horm Res. 1973;29:457-496.

Sindhu S, Maya P, Indira TN. A method for preparation of mustard (*Brassica juncea*) powder with retained pungency and reduced bitterness. LWT - Food Sci Tech. 2012;49:42-47.

Singh J, Sharma PC, Sharma SK, Kumar A. Standardization of the fourier transform near-infrared reflectance spectroscopy for estimation of some oil quality parameters in mustard (brassica spp.). Plant Soil Environ. 2013;59:478-483.

Singh K, Shakya R, Mahawar R. Genetic diversity and patterns of variation among Indian mustard (*Brassica juncea* (1.) czernj. & cosson) varieties. SABRAO J Breed Genet. 2014b;46:329-339.

Singh N, Kumar S, Singh P, Raj HG, Prasad AK, Parmar VS, Ghosh B. *Piper longum* Linn. Extract inhibits tnf-alphainduced expression of cell adhesion molecules by inhibiting nfkappab activation and microsomal lipid peroxidation. Phytomedicine. 2008;15:284-291.

Singh R, Arif T, Khan I, Sharma P. Phytochemicals in antidiabetic drug discovery. J Biomed Ther Sci. 2014a;1:1-33.

Sinha R, Jastreboff AM. Stress as a common risk factor for obesity and addiction. Biol Psychiatry. 2013;73:827-835.

Soni KB, Rajan A, Kuttan R. Reversal of aflatoxin induced liver damage by turmeric and curcumin. Cancer Lett. 1992;66:115-121.

Speciale A, Chirafisi J, Saija A, Cimino F. Nutritional antioxidants and adaptive cell responses: An update. Curr Mol Med. 2011;11:770-789.

Sri KS, Kumari DJ, Sivannarayana G. Effect of amla, an approach towards the control of diabetes mellitus. Int J Curr Microbiol Appl Sci. 2013;2:103-108.

Srikumar R, Parthasarathy NJ, Manikandan S, Narayanan GS, Sheeladevi R. Effect of triphala on oxidative stress and on cellmediated immune response against noise stress in rats. Mol Cell Biochem. 2006;28:67-74.

Steptoe A, Hackett RA, Lazzarino AI, Bostock S, La Marca R, Carvalho LA, Hamer M. Disruption of multisystem responses to stress in type 2 diabetes: Investigating the dynamics of allostatic load. Proc Natl Acad Sci U S A. 2014;111:15693-15698.

Sunila ES, Kuttan G. Immunomodulatory and antitumor activity of *Piper longum* Linn. And piperine. J Ethnopharmacol. 2004;90:339-346.

Suri A, Szallasi A. The emerging role of trpv1 in diabetes and obesity. Trends Pharmacol Sci. 2008;29:29-36.

Szallasi A. Piperine: Researchers discover new flavor in an ancient spice. Trends Pharmacol Sci. 2005;26:437-439.

Tarozzi A, Angeloni C, Malaguti M, Morroni F, Hrelia S, Hrelia P. Sulforaphane as a potential protective phytochemical against neurodegenerative diseases. Oxid Med Cell Longev. 2013;2013:415078.

Tatiraju DV, Bagade VB, Karambelkar PJ, Jadhav VM, Kadam V. Natural bioenhancers: An overview. J Pharmacogn Phytochem. 2013;2:55-60.

Thakur AK, Chatterjee SS, Kumar V. Antidepressant-like effects of *Brassica juncea* L. Leaves in diabetic rodents. Indian J Exp Biol. 2014a;52:613-622.

Thakur AK, Chatterjee SS, Kumar V. Anxiolytic-like activity of leaf extract of traditionally used Indian-mustard (*Brassica juncea*) in diabetic rats. TANG (Humanitas Medicine). 2013b;3:1-7.

Thakur AK, Chatterjee SS, Kumar V. Beneficial effects of *Brassica juncea* on cognitive functions in rats. Pharm Biol. 2013a;51:1304-1310.

Thakur AK, Shakya A, Husain GM, Emerald M, Kumar V. Gut-microbiota and mental health: Current and future perspectives. J Pharmacol Clin Toxicol. 2014b;2:1016.1-1016.15.

Thiyagarajan M, Sharma SS. Neuroprotective effect of curcumin in middle cerebral artery occlusion induced focal cerebral ischemia in rats. Life Sci. 2004;74:969-985.

Thomas M, Sujatha KS, George S. Protective effect of *Piper longum* Linn. On monosodium glutamate induced oxidative stress in rats. Indian J Exp Biol. 2009;47:186-192.

Tiwari AK, Rao JM. Diabetes mellitus and multiple therapeutic approaches of phytochemicals: Present status and future prospects. Curr Sci. 2002;83:30-38.

Tiwari AK. Invigorated barley in diabetes. Curr Sci. 2008;95:25-29.

Tomas-Menor L, Barrajon-Catalan E, Segura-Carretero A, Marti N, Saura D, Menendez JA, Joven J, Micol V. The promiscuous and synergic molecular interaction of polyphenols in bactericidal activity: An opportunity to improve the performance of antibiotics? Phytother Res. 2015;29:466-473.

Triggle DJ. Nous sommes tous des bacteries: Implications for medicine, pharmacology and public health. Biochem Pharmacol. 2012;84:1543-1550.

Tripathi MK, Mishra AS. Glucosinolates in animal nutrition: a review. Anim Feed Sci Tech. 2007;132:1-27.

Tripathi S, Raghuram N, Ramarao NH. Validation of an integrated ayurveda-yoga module for residential treatment of patients with type 2 diabetes mellitus - a compilation from traditional literature. Int J Ayurvedic Herbal Med. 2012;2:921-934.

Trowbridge A. The evolutionary ecology of chemically mediated plant-insect interactions. In Ecology and the

environment. Monson RK ed. (New York, USA: Springer), pp. 1-29, 2014.

Tschop MH, DiMarchi RD. Outstanding scientific achievement award lecture 2011: Defeating diabesity: The case for personalized combinatorial therapies. Diabetes. 2012;61:1309-1314.

Ulbricht C, Basch E, Barrette E-P, Boon H, Chao W, Costa D, Higdon ERB, Isaac R, Lynch M, Papaliodis G, Grimes Serrano JM, Varghese M, Vora M, Windsor R, Woods J. Turmeric (*Curcuma longa*): An evidence-based systematic review by the natural standard research collaboration. Altern Complement Ther. 2011;17:225-236.

Vaghasiya Y, Nair R, Chanda S. Investigation of piper species for antibacterial and anti-inflammatory property. Int J Pharmacol. 2007;3:400–405.

Vaibhavi B, Satyam T, Sanjibkumar P, Raghuram N, Ramarao NH. Effect of holistic module of yoga and ayurvedic panchakarma in type 2 diabetes mellitus-a pilot study. Open J Endocr Metab Dis. 2013;3:90-98.

Vaidya AD. Metabolic management: The role of nutraceuticals, nutritionals and naturals. J Obes Metab Res. 2014;1:79-82.

Vaiserman AM. Aging-modulating treatments: From reductionism to a system-oriented perspective. Front Genet. 2014;5:446.

van Duynhoven J, Vaughan EE, Jacobs DM, Kemperman RA, van Velzen EJ, Gross G, Roger LC, Possemiers S, Smilde AK, Dore J, Westerhuis JA, Van de Wiele T. Metabolic fate of polyphenols in the human superorganism. Proc Natl Acad Sci USA. 2011;108 :4531-4538.

van Ommen B, Stierum R. Nutrigenomics: Exploiting systems biology in the nutrition and health arena. Curr Opin Biotechnol. 2002;13:517-521.

Vedhanayaki G, Shastri GV, Kuruvilla A. Analgesic activity of *Piper longum* Linn. Root. Indian J Exp Biol. 2003;41:649-651.

Velusami CC, Boddapati SR, Hongasandra Srinivasa S, Richard EJ, Joseph JA, Balasubramanian M, Agarwal A. Safety evaluation of turmeric polysaccharide extract: Assessment of mutagenicity and acute oral toxicity. Biomed Res Int. 2013;2013:158348.

Venkateshwarlu G, Venkata NCH, Shantha TR, Kishore KR, Prathapa RM, Raghavendra HL. A comparative physicochemical and pharmacognostical evaluation of nishamalaki- an ayurvedic antidiabetic formulation. Sci Tech Arts Res J. 2013;2:69-78.

Verma S, Chatterjee SS, Kumar V. Comparative adaptogenic activity of bioavailable extracts of *Curcuma longa* and pure curcumin in rodents (NEU-22). Indian J Pharmacol. 2014;46:S93-S94.

Verma S, Chatterjee SS, Kumar V. Metformin like stress response modulating effects of turmeric curcuminoids in mice. SAJ Neurol. 2015;1:102.

Wakade AS, Shah AS, Kulkarni MP, Juvekar AR. Protective effect of *Piper longum* L. On oxidative stress induced injury

and cellular abnormality in adriamycin induced cardiotoxicity in rats. Indian J Exp Biol. 2008;46:528-533.

Walia H, Arora S. *Terminalia chebula-* a pharmacognistic account. J Med Plants Res. 2013;7:1351-1361.

Wang HH, Hsieh HL, Wu CY, Sun CC, Yang CM. Oxidized low-density lipoprotein induces matrix metalloproteinase-9 expression via a p42/p44 and jnk-dependent ap-1 pathway in brain astrocytes. Glia. 2009;57:24-38.

Wang MS, Boddapati S, Emadi S, Sierks MR. Curcumin reduces alpha-synuclein induced cytotoxicity in parkinson's disease cell model. BMC Neurosci. 2010;11:57.

Wang S, Moustaid-Moussa N, Chen L, Mo H, Shastri A, Su R, Bapat P, Kwun I, Shen CL. Novel insights of dietary polyphenols and obesity. J Nutr Biochem. 2014;25:1-18.

Wang Y, Tang H. Roles of herbal medicine in modulating gut microbiota associated with health and diseases. In Metabonomics and gut microbiota in nutrition and disease. Kochhar S, Martin F-P eds. (London, EK; Springer), pp. 185-197, 2015.

Wendlinger C, Hammann S, Vetter W. Various concentrations of erucic acid in mustard oil and mustard. Food Chem. 2014;153:393-397.

Winston D, Maimes S. Adaptogens: Herbs for strength, stamina, and stress relief. (Vermont, USA; Healing Arts Press), 2007.

Wu Y, Min X, Zhuang C, Li J, Yu Z, Dong G, Yao J, Wang S, Liu Y, Wu S, Zhu S, Sheng C, Wei Y, Zhang H, Zhang W, Miao Z. Design, synthesis and biological activity of piperlongumine derivatives as selective anticancer agents. Eur J Med Chem. 2014:82:545-551.

Xia X, Cheng G, Pan Y, Xia ZH, Kong LD. Behavioral, neurochemical and neuroendocrine effects of the ethanolic extract from *Curcuma longa* L. In the mouse forced swimming test. J Ethnopharmacol. 2007;110:356-363.

Xia X, Pan Y, Ou-Yang Z, Wang J, Pan L-L, Zhu Q, Huang J-J, Kong L-D. Pharmacokinetic-pharmacodynamic modeling of monoamine oxidase a inhibitory activity and behavior improvement by curcumin in the mouse forced swimming test. Chin J Nat Med. 2011;9:293-304.

Xia X, Pan Y, Zhang WY, Cheng G, Kong LD. Ethanolic extracts from *Curcuma longa* attenuates behavioral, immune, and neuroendocrine alterations in a rat chronic mild stress model. Biol Pharm Bull. 2006;29:938-944.

Xiao JB, Hogger P. Dietary polyphenols and type 2 diabetes: Current insights and future perspectives. Curr Med Chem. 2015;22:23-38.

Xu Y, Ku BS, Yao HY, Lin YH, Ma X, Zhang YH, Li XJ. Antidepressant effects of curcumin in the forced swim test and olfactory bulbectomy models of depression in rats. Pharmacol Biochem Behav. 2005;82:200-206.

Yadav SP, Vats V, Ammini AC, Grover JK. *Brassica juncea* (rai) significantly prevented the development of insulin resistance in rats fed fructose-enriched diet. J Ethnopharmacol. 2004;93:113-116.

Yadav V, Kumar, V. Advances in modern knowledge of dravyaguna of Piper longum during last five years (REV-15). Indian J Pharmacol. 2014;4:S109.

Yang C, Zhang X, Fan H, Liu Y. Curcumin upregulates transcription factor nrf2, ho-1 expression and protects rat brains against focal ischemia. Brain Res. 2009;1282:133-141.

Yang F, Lim GP, Begum AN, Ubeda OJ, Simmons MR, Ambegaokar SS, Chen PP, Kayed R, Glabe CG, Frautschy SA, Cole GM. Curcumin inhibits formation of amyloid beta oligomers and fibrils, binds plaques, and reduces amyloid *in vivo*. J Biol Chem. 2005;280:5892-5901.

Yang J, Su Y, Luo J-F, Gu W, Niu H-M, Li Y, Wang Y-H, Long C-L. New amide alkaloids from *Piper longum* fruits. Nat Prod Bioprospect. 2013;3:277-281.

Yang T, Sun S, Wang T, Tong X, Bi J, Wang Y, Sun Z. Piperlonguminine is neuroprotective in experimental rat stroke. Int Immunopharmacol. 2014;23:447-451.

Yokozawa T, Kim HY, Cho EJ, Choi JS, Chung HY. Antioxidant effects of isorhamnetin 3,7-di-o-beta-dglucopyranoside isolated from mustard leaf (*Brassica juncea*) in rats with streptozotocin-induced diabetes. J Agric Food Chem. 2002;50:5490-5495.

Yokozawa T, Kim HY, Cho EJ, Yamabi N, Choi JS. Protective effects of mustard leaf (*Brassica juncea*) against diabetic oxidative stress. J Nutr Sci Vitaminol (Tokyo). 2003;49:87-93.

Yoon BH, Jung JW, Lee JJ, Cho YW, Jang CG, Jin C, Oh TH, Ryu JH. Anxiolytic-like effects of sinapic acid in mice. Life Sci. 2007;81:234-240.

Yu JC, Jiang ZT, Li R, Chan SM. Chemical composition of the essential oils of *Brassica juncea* (L.) coss. Grown in different regions, hebei, shaanxi and shandong, of china. J Food Drug Anal. 2003;11:22-26.

Yu SY, Gao R, Zhang L, Luo J, Jiang H, Wang S. Curcumin ameliorates ethanol-induced memory deficits and enhanced brain nitric oxide synthase activity in mice. Prog Neuropsychopharmacol Biol Psychiatry. 2013;44:210-216.

Yu ZF, Kong LD, Chen Y. Antidepressant activity of aqueous extracts of *Curcuma longa* in mice. J Ethnopharmacol. 2002;83:161-165.

Zaki M, Begum W, Bhat TA, Kausar H. Amla (*Emblica officinalis* gaertn) the wonderful unani drug: A review. World J Pharm Pharm Sci. 2014;3:1369-1381.

Zaveri M, Khandhar A, Patel S, A. P. Chemistry and pharmacology of *Piper longum* L. Int J Pharm Sci Rev Res. 2010;5:67-76.

Zhang DW, Fu M, Gao SH, Liu JL. Curcumin and diabetes: A systematic review. Evid Based Complement Alternat Med. 2013:636053.

Zhang YJ, Abe T, Tanaka T, Yang CR, Kouno I. Phyllanemblinins a-f, new ellagitannins from *Phyllanthus emblica*. J Nat Prod. 2001;64:1527-1532.

Zhao L, Ackerman SL. Endoplasmic reticulum stress in health and disease. Curr Opin Cell Biol. 2006;18:444-452.