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Genetic Parameters of Reproductive and Meat Quality Traits in Korean Berkshire Pigs

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ABSTRACT: Genetic parameters of Berkshire pigs for reproduction, carcass and meat quality traits were estimated using the records from a breeding farm in Korea. For reproduction traits, 2,457 records of the total number of piglets born (TNB) and the number of piglets born alive (NBA) from 781 sows and 53 sires were used. For two carcass traits which are carcass weight (CW) and backfat thickness (BF) and for 10 meat quality traits which are pH value after 45 minutes (pH45m), pH value after 24 hours (pH24h), lightness in meat color (LMC), redness in meat color (RMC), yellowness in meat color (YMC), moisture holding capacity (MHC), drip loss (DL), cooking loss (CL), fat content (FC), and shear force value (SH), 1,942 pig records were used to estimate genetic parameters. The genetic parameters for each trait were estimated using VCE program with animal model. Heritability estimates for reproduction traits TNB and NBA were 0.07 and 0.06, respectively, for carcass traits CW and BF were 0.37 and 0.57, respectively and for meat traits pH45m, pH24h, LMC, RMC, YMC, MHC, DL, CL, FC, and SH were 0.48, 0.15, 0.19, 0.36, 0.28, 0.21, 0.33, 0.45, 0.43, and 0.39, respectively. The estimate for genetic correlation coefficient between CW and BF was 0.27. The Genetic correlation between pH24h and meat color traits were in the range of -0.51 to -0.33 and between pH24h and DL and SH were -0.41 and -0.32, respectively. The estimates for genetic correlation coefficients between reproductive and meat quality traits were very low or zero. However, the estimates for genetic correlation coefficients between reproductive traits and drip and cooking loss were in the range of 0.12 to 0.17 and -0.14 to -0.12, respectively. As the estimated heritability of meat quality traits showed medium to high heritability, these traits may be applicable for the genetic improvement by continuous measurement. However, since some of the meat quality traits showed negative genetic correlations with carcass traits, an appropriate breeding scheme is required that carefully considers the complexity of genetic parameters and applicability of data. (Key Words: Berkshire, Genetic Parameter, Heritability, Repeatability, Genetic Correlation)

INTRODUCTION

Meat productivity traits such as carcass weight (CW) and backfat thickness and reproductive traits such as total litter size and the number of piglets born alive are very important economic traits in swine. Therefore, until now selection of pigs has been focused on these traits. However,

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due to the improved eating habits of consumers, breeding objectives have also put weight on meat quality (van Wijk et al., 2005). As customers' demands for pork depend upon the meat quality and its physical and biochemical components (Bonneau and Lebret, 2010), it is time that genetic improvement is required not only for productive and reproductive traits but also meat quality traits. Meat quality traits are highly heritable, causing an efficient selection response (Newcom et al., 2002). Therefore, meat quality is a trait applicable for the genetic improvement and should be one of the major selection benchmarks for breeding process in the swine industry (Luo et al., 2012).

It has been reported that there is a negative or very low genetic correlation between reproductive traits and meat productivity and quality traits (Ducos and Bidanel, 1996; Clutter and Brascamp, 1998; Chen et al. 2003b; Holm et al.,

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2004). When traits have a negative genetic correlation, it is difficult to select simultaneously for both traits. The establishment of the breeding improvement direction should be accurate when evaluating the genetic performance of sires and the most important preceding condition for the establishment of breeding improvement direction is the precise estimation of genetic parameters.

Recently, production of Berkshire pigs has increased in Korea because of their excellent meat qualities such as marbling, juiciness, tenderness and flavor. For the genetic improvement of Korean Berkshire pigs, selection should simultaneously consider productive, reproductive and meat quality traits, which requires the precise estimation of genetic parameters.

This experiment was conducted to estimate genetic correlations among major economic traits such as reproductive, carcass and meat quality traits in the Berkshire pig population and to establish selection and improvement direction thereafter.

MATERIALS AND METHODS

Reproduction traits

A total of 2,511 records on the reproductive traits of Berkshire sows farrowed from 2007 to 2013 were collected from Dasan Pig Breeding Farm (Namwon, Korea). Data that lacked litter size, abortion during pregnancy and farrowing difficulty were excluded from the study. Therefore, for the final analysis only 2,457 records composed of the total number of piglets born (TNB) and the numbers of piglets born alive (NBA) from 781 sows were used, while the total number of sires were 53 heads. The mean and maximum for the parity of sows were 3.27 and 10, respectively and the pedigree was constructed with 7 generations.

Carcass and meat quality traits

For carcass and meat quality traits, data were collected from 1,953 pigs which were butchered at an abattoir in Namwon, but 11 incomplete records were deleted and only records from 1,942 pigs were used for the analysis and a total of 12 traits including (1) CW, (2) backfat thickness (BF), (3) pH value after 45 minutes (pH45m), (4) pH value after 24 hours (pH24h), (5) lightness in meat color (LMC), (6) redness in meat color (RMC), (7) yellowness in meat color (YMC), (8) moisture holding capacity (MHC), (9) drip loss (DL), (10) cooking loss (CL), (11) fat content (FC), and (12) shear force value (SH) were measured.

The CW (kg) and BF (mm) were measured at the parts of loins (*Longissimus dorsi*, LD) on the left side of the cold carcasses and used to determine meat quality parameters. The values for pH45m and pH24h were measured from the time of slaughter after inserting a pH meter (Orion 2 Star, Thermo Scientific, Waltham, MA, USA) directly into the muscle. The meat color values were measured using a colorimeter (CR-410, Minolta Co., Osaka, Japan) which was calibrated against a white reference tile plate (L^* = 89.2, $a^* = 0.921$, $b^* = 0.783$). The color values for L* (lightness, LMC), a* (redness, RMC), and b* (yellowness, YMC) were obtained from the inside cuts of loins after 30 minutes blooming at room temperature. Centrifugation was used to measure MHC. Minced meat samples (5 g) were placed into a centrifuge tube with a filter paper (No. 4, Whatman International Ltd., Maidston, UK), and centrifuged at $3,000 \times g$ for 10 minutes. The MHC was calculated as the remaining moisture in the meat sample on the basis of the moisture content of the original meat sample. DL was measured as the percentage weight loss of a standardized $(3 \times 3 \times 3 \text{ cm})$ meat sample placed in a sealed petri-dish at 4°C after 48 hours of storage. For the measurement of CL, the percentage weight loss of a standardized $(3 \times 3 \times 3 \text{ cm})$ meat sample was measured after cooking in an electric grill with double pans (Nova EMG-533, 1,400 W, Evergreen Enterprise, Yongin, Korea) for 90 seconds, until the internal temperature of the meat sample reached 72°C. The parts of loins $(3 \times 3 \times 2 \text{ cm})$ were cut and after installing a texture analyzer (TA-XT2, Stable Micro System Ltd., Godalming, UK) with a Warner-Bratzler blade, SH was measured with the blade cutting the muscle fibres at a right angle. Test and pre-test speeds were set at 2.0 mm/s and post-test speeds were set at 5.0 mm/s. The crude FCs were extracted according to the method described by Folch et al. (1957).

Statistical method

The analytical animal models (1 and 2) used for estimating the genetic parameters and expected breeding values (EBVs) were as follows:

$$y_{ijklm} = \mu + f_i + l_j + b_k + a_l + p_l + e_{ijklm}$$
⁽¹⁾

Where, y_{ijklm} = reproductive traits, μ = overall mean, f_i = fixed effect of the ith farrowed year-month-week (i = 1, 2, ..., 276), l_j = fixed effect of the jth parity (j = 1, 2, ..., 8 more than), b_k = random effect of the kth sire (k = 1, 2, ..., 53), a_l = random additive genetic effect of the lth animal (l = 1, 2, ..., 994), p_l = permanent environmental effect of individual animals (l = 1, 2, ..., 781) and e_{ijklm} = residual random effect.

$$y_{ijkl} = \mu + s_i + m_j + a_k + e_{ijkl} \tag{2}$$

Where, y_{ijkl} = carcass and meat quality traits, μ = overall mean, s_i = fixed effect of the ith sex (i = 1, 2), m_j = fixed effect of the jth year-month of birth (j = 1, 2, ..., 37), a_k = random additive genetic effect of the kth animal (k = 1, 2, ...,





Figure 1. Distributions of pH value after 45 minutes and cooking loss.

2,487) and e_{ijkl} = residual random effect. Var(a) = A $\otimes \sigma_{a}^{2}$, Var(pe) = I $\otimes \sigma_{pe}^{2}$, and Var(e) = I $\otimes \sigma_{e}^{2}$, where A = numerator relationship matrix and I = identity matrix. Basic statistical analysis was performed using SAS ver. 9.2 (SAS Institute Inc., Cary, NC, USA) and genetic parameters and EBVs for traits were estimated using VCE ver. 6.0 (Institute of Farm Animal Genetics, Friedrich Loeffler Institute, Neustadt, Germany) and PEST program (Institute of Animal Husbandry and Animal Behaviour, Federal Agricultural Research Centre, Neustadt, Germany). The genetic correlations between reproductive and meat quality traits were estimated using the EBVs of sows with progeny records of carcass and meat quality.

RESULTS AND DISCUSSION

Data description

Basic data for the major economic traits of Berkshire pigs including reproductive, carcass and meat quality traits are shown in Table 1. The means for TNB and NBA were 8.65 ± 2.95 and 7.65 ± 2.58 , respectively. The means for CW and BF were 86.62 ± 5.67 kg and 25.11 ± 5.23 mm, respectively and means for pH45m and pH24h, which are criteria for cadaveric metabolic rate, were 6.06 ± 0.29 and 5.78 ± 0.22 , respectively. The means for LMC, RMC, and YMC, which are the evaluation indicators for abnormal meat, were 49.05 ± 3.18 , 8.47 ± 4.34 , and 3.59 ± 1.48 , respectively. The means for MHC, DL, CL, and FC were $57.90\pm11.10\%$, $4.14\pm2.07\%$, $23.80\pm6.00\%$, and $2.64\pm1.06\%$, respectively and SH which is the indicator of tenderness

Table 1. Means, standard deviations (SD) and skewness for reproductive (n = 2,511), carcass and meat quality traits (n = 1,942)

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Traits	Mean±SD	Skewness
Total number of piglets born (head)	8.65±2.95	-0.07
Number of piglets born alive (head)	7.65±2.58	-0.33
Carcass weight (kg)	86.62±5.67	0.20
Backfat thickness (mm)	25.11±5.23	0.19
pH value after 45 minutes	6.06±0.29	-0.03
pH value after 24 hours	5.78±0.22	0.92
Lightness in meat color	49.05±3.18	-0.01
Redness in meat color	8.47±4.34	1.17
Yellowness in meat color	3.59±1.48	0.23
Moisture holding ability (%)	57.90±11.10	0.17
Drip loss (%)	4.14±2.07	1.38
Cooking loss (%)	23.80±6.00	-0.50
Fat (%)	2.64±1.06	1.26
Shear force value (kg)	2.82±0.76	0.96

was 2.82 ± 0.76 kg. Distributions of pH45m and CL which were considered as the most important traits for the genetic improvement in Dasan Pig Breeding Farm are shown in Figure 1. The pH24h showed a normal distribution, however, the CL was not normally distributed (p<0.01) by Shapiro Wilk test (SAS).

Heritability estimates for TNB and NBA in Berkshire were 0.07 and 0.06, respectively (Table 2), which were similar to those of Su et al. (2007) who reported the heritability of TNB in Landrace (0.08) and Yorkshire (0.05), and those of Chen et al. (2003a) in Landrace (0.08),

Table 2. Additive genetic (σ_a^2), permanent environmental (σ_{pe}^2), boar (σ_b^2) and error variance (σ_e^2) components, heritabilities (h^2) and standard errors (SE), repeatabilities and phenotypic and genetic correlations for reproductive traits

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Traits	σ^2_a	σ^2_{pe}	σ^2_b	σ_e^2	h ² ±SE	r	(1) TNB	(2) NBA
(1) TNB	0.61	0.75	0.05	7.11	0.07 ± 0.03	0.16		0.916 ^a
(2) NBA	0.44	0.70	0.04	5.31	0.06 ± 0.03	0.18	0.988^{b}	

TBN, total number of piglets born; NBA, the number of piglets born alive.

^a Phenotypic. ^b Genetic correlation.

Yorkshire (0.10), and Duroc (0.09). Generally, heritability for litter size seems to be low although it depends on the genetic makeup in a population (Arango et al., 2005). Repeatability estimates for TNB and NBA were 0.16 and 0.18, respectively and the sire effect was estimated as 1% of total variation, which was lower than that (5%) of Woodward et al. (1993), while genetic correlation between TNB and NBA was very high (0.988).

Heritabilities for carcass and meat quality traits are shown in Table 3. Heritability estimates for CW and BF were 0.37 and 0.57, respectively, which were similar to results by Tomiyama et al. (2011) who reported 0.54 for BF at finish and 0.32 for CW in a Japanese Berkshire population (n = 4,773). For other breeds, it was reported that the heritability for BF ranged from 0.47 to 0.56 (Bryner et al., 1992; Li and Kennedy, 1994; te Pas and Soumillion, 1999; Chen et al. 2002; Johnson and Nugent, 2003), which was similar to our results.

Heritability estimates for pH45m and pH24h, which are criteria for cadaveric metabolic rate, were 0.48 and 0.15, respectively. The heritability decreased sharply by the measurement time probably due to environmental factors such as storage temperature and management. The heritability estimates for pH24h by Sellier (1998), Hermesch et al. (2000), Suzuki et al. (2005), and Gaya et al. (2006) were 0.21, 0.07, 0.14, and 0.37, respectively. Since the pH45m and pH24h are the standards for muscle quality evaluation, they will be very important traits for meat quality improvement if we can obtain precise measurements. The heritability estimates for LMC was 0.19, which was lower than that (0.28) of Sellier (1998) and that (0.29) of Gava et al. (2006). The heritability estimates for RMC and YMC were 0.36 and 0.28, respectively, which were higher than those (0.25 and 0.16) of Gaya et al. (2006). The heritability estimates for MHC, DL, and CL were 0.21, 0.33,

Table 3. Additive genetic (σ_a^2) and error variance (σ_e^2) components, and heritabilities (h^2) and standard errors (SE) for carcass and meat quality traits

Traits	σ_a^2	σ_{e}^{2}	h ² ±SE
Carcass weight	11.20	18.76	0.37 ± 0.07
Backfat thickness	15.94	12.09	0.57 ± 0.06
pH value after 45 minutes	0.03	0.04	0.48 ± 0.06
pH value after 24 hours	0.01	0.05	0.15 ± 0.05
Lightness in meat color	2.05	8.49	$0.19{\pm}0.05$
Redness in meat color	0.41	0.71	0.36 ± 0.06
Yellowness in meat color	0.29	0.73	0.28 ± 0.06
Moisture holding ability	23.10	87.75	0.21±0.09
Drip loss	1.10	2.22	0.33 ± 0.06
Cooking loss	6.69	8.29	0.45 ± 0.06
Fat content	0.45	0.61	0.43 ± 0.06
Shear force value	0.19	0.30	0.39 ± 0.07

and 0.45, respectively. It was reported that the heritability estimate for DL was 0.23 (Hermesch et al., 2000) or 0.25 (Gaya et al., 2006) and the heritability estimates for MHC and CL were in the range of 0 to 0.63 (Hovenier et al., 1993) and 0 to 0.51 (Sellier, 1998). The heritability estimate for FC was 0.43, which was similar to estimates obtained by Knapp et al. (1997; 0.42), Larzul et al. (1997; 0.44), and Hermesch et al. (2000; 0.35). The heritability estimates for SH was 0.39, which was lower than that by Suzuki et al. (2005; 0.45), but higher than that by Gaya et al. (2006; 0.22). These differences of estimated genetic parameters by other researches can be cause by the model, measurement and size of data used in the analysis.

Genetic correlation

The estimate for genetic correlation coefficient between CW and BF was 0.27, which was lower than that (0.42) obtained by Tomiyama et al. (2011) and the estimates for

Table 4. Genetic and phenotypic correlations among carcass and meat quality traits

Traits	(1)	(2)	(3)	(4)	(5)	(6)	(7)	(8)	(9)	(10)	(11)	(12)
(1) CW (1)		0.27	0.29	-0.05	-0.24	-0.08	-0.19	-0.12	0.00	-0.36	0.04	-0.04
(2) BF (2)	0.26		0.16	0.07	-0.30	0.01	-0.19	0.18	-0.14	-0.28	0.27	-0.30
(3) pH45m	-0.02	-0.02		0.06	-0.45	0.23	0.01	-0.32	-0.05	-0.23	0.14	0.04
(4) pH24h	0.00	0.07	0.10		-0.33	-0.51	-0.51	0.20	-0.41	0.05	0.13	-0.32
(5) LMC	-0.05	-0.04	-0.17	-0.13		0.03	0.75	-0.10	0.53	0.68	0.16	0.14
(6) RMC	0.04	0.10	-0.03	-0.12	-0.07		0.41	-0.01	0.34	0.11	0.03	-0.02
(7) YMC	-0.06	-0.02	-0.05	-0.32	0.57	0.40		-0.25	0.43	0.62	-0.02	0.19
(8) MHC	0.02	0.05	-0.10	0.02	-0.01	-0.03	-0.05		0.04	0.06	-0.08	-0.06
(9) DL	0.03	0.00	-0.27	-0.27	0.35	0.20	0.34	0.01		0.20	-0.22	0.05
(10) CL	-0.08	-0.16	-0.15	-0.19	0.33	0.12	0.40	-0.03	0.28		0.09	0.44
(11) FC	0.06	0.33	0.11	-0.01	0.14	0.18	0.26	-0.02	-0.04	-0.02		-0.17
(12) SH	-0.03	-0.19	-0.14	-0.15	0.04	0.04	0.09	-0.02	0.23	0.32	-0.16	

Upper triangle, genetic correlation; lower triangle, phenotypic correlation.

CW, arcass weight; BF, Back fat thickness; pH45m, pH value after 45 minutes; pH24h, pH value after 24 hours; LMC, lightness in meat color; RMC, redness in meat color; YMC, yellowness in meat color; MHC, moisture holding ability; DL, drip loss; CL, cooking loss; FC, fat content; SH, shear force value.

genetic correlation coefficients between CW and meat quality traits were in the range of -0.36 to 0.29 (Table 4). The genetic correlation coefficient between BF and FC was positive (0.27), while the genetic correlation coefficients between BF and CL and between BF and SH were negative (-0.28 and -0.30). The genetic correlation coefficients between pH24h and meat color traits were in the rage of -0.51 to -0.33, which were similar to those of Suzuki et al. (2005). The genetic correlation coefficients of pH24h for DL, CL, and SH were -0.41, 0.05, and -0.32, respectively.

The genetic correlation coefficients between DL and meat color traits were in the range of 0.34 to 0.53, and those between CL and meat color traits were in the range of 0.11 to 0.68, showing positive relationship.

Genetic correlation coefficient between DL and CL was 0.20. A higher value for genetic correlation (0.66) was reported by Sellier (1998), but Suzuki et al. (2005) reported a low genetic correlation coefficient (0.04) between DL and CL when measured by the same method used in this experiment. Genetic correlation coefficients between SH and FC was -0.17, which was similar to that (-0.20) by Suzuki (2005). Genetic correlation coefficients between SH and DL and between SH and CL were 0.05 and 0.44, respectively, and positively correlated. The estimates for genetic correlation coefficients between reproductive and meat quality traits were very low or zero (Table 5), which were similar to previous reports (Ducos and Bidanel, 1996; Clutter and Brascamp, 1998; Chen et al., 2003b; Holm et al., 2004). However, the estimates for genetic correlation coefficients between reproductive traits and DL and CL were in the range of 0.12 to 0.17, and -0.14 to -0.12, respectively, which were quite different from those of other meat quality traits, suggesting that DL and CL may have different water retention mechanisms.

Table 5. Genetic correlations between carcass and meat qualitytraits and reproductive traits (pairs = 308)

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Traits	TNB	NBA
Carcass weight	-0.09^{NS}	-0.05^{NS}
Back fat thickness	-0.11^{NS}	-0.12*
pH value after 45 minutes	-0.04^{NS}	-0.03^{NS}
pH value after 24 hours	0.12*	$0.07^{ m NS}$
Lightness in meat color	-0.06^{NS}	-0.03^{NS}
Redness in meat color	-0.01^{NS}	0.01^{NS}
Yellowness in meat color	-0.14*	-0.10^{NS}
Moisture holding ability	0.00^{NS}	0.03 ^{NS}
Drip loss	0.12*	0.17**
Cooking loss	-0.12*	-0.14*
Shear force value	-0.02^{NS}	-0.04^{NS}
Fat	-0.07^{NS}	-0.07^{NS}

TBN, total number of piglets born; NBA, the number of piglets born alive; NS, non-significant.

* p<0.05, ** p<0.01.

CONCLUSION

Since carcass and meat quality traits have medium to high heritability, these valuable traits will respond to selection more quickly than reproductive traits since the heritability estimates for these traits are below 10%. Furthermore, the evaluation of meat quality, is very difficult to apply on the spot since various physical and chemical factors should be evaluated. Therefore, it seems to be practical to put emphasis on measuring pH value and backfat thickness, which are measurable on the spot, and then run parallel with selection for litter size. To overcome weaknesses of this study, the use of genomic selection would be desirable after establishing reference population for meat quality traits.

CONFLICT OF INTEREST

We certify that there is no conflict of interest with any financial organization regarding the material discussed in the manuscript.

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