

## *In vitro* antibiotic susceptibility of field isolates of *Mycoplasma hyopneumoniae* and *Mycoplasma hyorhinis* from Korea

Jisung Jang<sup>1</sup>, Kiju Kim<sup>1</sup>, Soyeon Park<sup>1</sup>, Bokyoung Park<sup>1</sup>, Hyungmin Um<sup>2</sup>, Marc Coulier<sup>3</sup>, Tae-Wook Hahn<sup>1,\*</sup>

<sup>1</sup>Department of Veterinary Medicine, College of Veterinary Medicine & Institute of Veterinary Science, Kangwon National University, Chuncheon 24341, Korea

<sup>2</sup>DongBang Co., Ltd., Seoul 06301, Korea

<sup>3</sup>ECO Animal Health, London N14 6WS, United Kingdom

(Received: February 4, 2016; Revised: April 8, 2016; Accepted: April 15, 2016)

**Abstract:** The present study was conducted to determine the antibiotic susceptibilities of local *Mycoplasma hyopneumoniae* (Mhp) and *Mycoplasma hyorhinis* (Mhr) field isolates. Minimum inhibitory concentrations (MICs) of Mhp and Mhr field isolates (twelve each) obtained from enzootic pneumonia-like lung lesions during 2009–2011 from Korea were determined using the broth microdilution method. Tylvalosin showed the highest activity against Mhp and Mhr field isolates, with MIC<sub>90</sub> values of 0.06 µg/mL and 0.12 µg/mL, respectively. Therefore, Korean Mhp and Mhr isolates are highly susceptible to tylvalosin.

**Keywords:** *Mycoplasma hyopneumoniae*, *Mycoplasma hyorhinis*, antimicrobials, minimum inhibitory concentration, porcine enzootic pneumonia

Swine mycoplasma *Mycoplasma hyopneumoniae* (Mhp) and *Mycoplasma hyorhinis* (Mhr) are respiratory pathogens in pigs. They are associated with enzootic pneumonia (EP) and porcine respiratory disease complex (PRDC) causing huge losses to the porcine industry. Currently commercial vaccines are available only against Mhp; however, their protection ability is not complete [7]. Therefore, use of antimicrobials often becomes necessary to limit the disease in the event of outbreak or as an additional measure to prevent mycoplasma disease. Although considerable data have been obtained on the *in vitro* antibiotic susceptibility of Mhp and Mhr around the world [6, 8–11], there are few reports about their susceptibility, and to our knowledge, there is no recent information on antibiotic susceptibility of these mycoplasmas from Korea. In the present study, minimum inhibitory concentrations (MICs) of Korea Mhp and Mhr isolates to tylvalosin and other commonly used antibiotics (tiamulin, lincomycin, tilmicosin and chlortetracycline) was determined.

The reference strains used in this study were Mhp (ATCC 25934) and Mhr (ATCC 27717) (American Type Culture Collection, USA). Field isolates (twelve each) of Mhp and Mhr which were obtained from EP-like lung lesions during 2009–2011 were used in this study [1, 2]. Each of these isolates was identified by multiplex polymerase chain reaction (mPCR) [3] and passaged 6 times before using in the anti-

microbial assay. Both *Mycoplasma* species were propagated in Friis broth [4] and used at final concentration of approximately  $1 \times 10^5$  CCU/mL. The antimicrobials chlortetracycline, tiamulin and tilmicosin were obtained from Sigma (USA). The lincomycin and tylvalosin were obtained from Dong Bang (Korea) and ECO Animal Health (UK), respectively. The antimicrobials were prepared by binary micro dilution method, in 96-well micro plate (ranging between 64 and 0.0038 µg/mL; SPL Life Sciences, Korea). MICs were determined using the broth microdilution method as described by Hannan *et al.* [5]. Readings were taken after incubation of 7 days at 37°C, and the lowest concentration of antimicrobial inhibiting color change from red to yellow/orange of the medium was defined as MIC of the drug.

The results of the *in vitro* antimicrobial testing for mycoplasma field isolates and type strains are presented in Table 1 as MIC ranges, MICs at which 50% and 90% of the isolates were inhibited (MIC<sub>50</sub> and MIC<sub>90</sub>). The values for three replicate testing of type strains (both Mhp and Mhr) were equal or differed from each other by only one doubling dilution, indicating good reproducibility of the test. For the Mhp field isolates, tylvalosin showed the lowest MICs values with MIC<sub>50</sub> and MIC<sub>90</sub> of 0.03 µg/mL and 0.06 µg/mL, respectively. Next to tylvalosin and tiamulin showed high activity and its MIC<sub>90</sub> value was 0.12 µg/mL. High potency was also

\*Corresponding author

Tel: +82-33-250-8671, Fax: +82-33-259-5625

E-mail: twahn@kangwon.ac.kr

**Table 1.** Minimal inhibitory concentrations (MIC) ranges for 5 antimicrobials against *Mycoplasma (M.) hyopneumoniae* and *M. hyorhinis* isolates in Korea

Mycoplasma species and antibiotic*	Field isolates			
	Type strain†	Range	MIC <sub>50</sub>	MIC <sub>90</sub>
<i>M. hyopneumoniae</i>				
Tylvalosin	≤ 0.03	0.0075–0.06	0.03	0.06
Lincomycin	0.5	0.12–0.5	0.25	0.5
Chlortetracycline	4	4–64	16	64
Tiamulin	0.12	0.06–0.25	0.12	0.12
Tilmicosin	≤ 4	0.5–4	1	4
<i>M. hyorhinis</i>				
Tylvalosin	0.06	0.06–0.12	0.06	0.12
Lincomycin	0.25	0.25–1	1	1
Chlortetracycline	2	2–64	64	64
Tiamulin	0.06	0.12–0.25	0.12	0.25
Tilmicosin	≤ 1	0.12–4	2	4

\*All data are given in µg/mL. †The type strains used in this study were *M. hyopneumoniae* ATCC 25934 and *M. hyorhinis* ATCC 27717. Values indicate results of three replicate testing of type strains.

observed for antimicrobials lincomycin and tilmicosin with MIC<sub>90</sub> of 0.5 µg/mL and 4 µg/mL, respectively. The antimicrobial chlortetracycline, with MIC range from 4 to 64 µg/mL and MIC<sub>90</sub> of 64 µg/mL, displayed low activity against the Mhp field isolates.

Similar findings were observed for the Mhr field isolates. Tylvalosin had an MIC<sub>90</sub> of 0.12 µg/mL and was the most active antimicrobial against all Mhr isolates. It was followed by tiamulin, lincomycin, and tilmicosin with MIC<sub>90</sub> values of 0.25 µg/mL, 1 µg/mL and 4 µg/mL, respectively. As with the Mhp isolates, chlortetracycline was also less active against the Mhr field isolates with MIC range from 4 to 64 µg/mL and MIC<sub>90</sub> of 64 µg/mL.

For the Mhp field isolates, MIC<sub>90</sub> values of tylvalosin, tiamulin and chlortetracycline were consistent with the previous studies [8-10]. Lincomycin had an MIC<sub>90</sub> value almost same as the isolates from Thailand [9], but it was four times of the corresponding MIC<sub>90</sub> values for the isolates from Spain [8] and Belgium [10]. Likewise, tilmicosin had MIC<sub>90</sub> value four and eight times of the values for isolates from Spain [8] and Belgium [10], respectively. In case of Mhr isolates, the MIC<sub>90</sub>s of lincomycin, tilmicosin and tiamulin are in agreement with the previously published reports [6, 11]. Tylvalosin, which had an *in vitro* activity up to thousand times higher compared to other antibiotics, showed highest efficacy against both mycoplasma species.

To conclude, our findings of *in vitro* susceptibility must be taken into consideration while treating these swine mycoplasma in Korea and will help for a choice among several antibiotics. The *in vitro* sensitivity testing of field isolates of Mhp and Mhr from Korea shows that tylvalosin was the most effective antimicrobial against these *mycoplasma* followed by

tiamulin, lincomycin, and tilmicosin.

## Acknowledgments

This research was supported by the Technology Development Program for Agriculture and Forestry, Ministry for Food, Agriculture, Forestry and Fisheries, and was technically supported by the Institute of Veterinary Science, Kangwon National University, Korea.

## References

1. Barate AK, Jang G, Cho SB, Hahn TW. RAPD and SDS-PAGE analysis of *Mycoplasma hyorhinis* isolates of South Korea. Indian J Anim Sci 2014, **84**, 26-28.
2. Barate AK, Jang GS, Cho S, Hahn TW. Proteomic and genetic diversity of *Mycoplasma hyopneumoniae* isolates from South Korea. J Pure Appl Microbiol 2013, **7**, 3125-3129.
3. Barate AK, Lee HY, Jeong HW, Truong LQ, Joo HG, Hahn TW. An improved multiplex PCR for diagnosis and differentiation of *Mycoplasma hyopneumoniae* and *Mycoplasma hyorhinis*. Korean J Vet Res 2012, **52**, 39-43.
4. Friis NF. Some recommendations concerning primary isolation of *Mycoplasma suis pneumoniae* and *Mycoplasma flocculare* a survey. Nord Vet Med 1975, **27**, 337-339.
5. Hannan PCT. Guidelines and recommendations for antimicrobial minimum inhibitory concentration (MIC) testing against veterinary mycoplasma species. Vet Res 2000, **31**, 373-395.
6. Hannan PCT, Windsor GD, de Jong A, Schmeer N, Stegemann M. Comparative susceptibilities of various animal-pathogenic mycoplasmas to fluoroquinolones. Antimicrob Agents Chemother 1997, **41**, 2037-2040.
7. Maes D, Segales J, Meyns T, Sibila M, Pieters M,

- Haesebrouck F.** Control of *Mycoplasma hyopneumoniae* infections in pigs. *Vet Microbiol* 2008, **126**, 297-309.
8. **Tavío MM, Poveda C, Assunção P, Ramírez AS, Poveda JB.** In vitro activity of tylvalosin against Spanish field strains of *Mycoplasma hyopneumoniae*. *Vet Rec* 2014, **175**, 539.
9. **Thongkamkoon P, Narongsak W, Kobayashi H, Pathanasophon P, Kishima M, Yamamoto K.** *In vitro* susceptibility of *Mycoplasma hyopneumoniae* field isolates and occurrence of fluoroquinolone, macrolides and lincomycin resistance. *J Vet Med Sci* 2013, **75**, 1067-1070.
10. **Vicca J, Stakenborg T, Maes D, Butaye P, Peeters J, de Kruif A, Haesebrouck F.** In vitro susceptibilities of *Mycoplasma hyopneumoniae* field isolates. *Antimicrob Agents Chemother* 2004, **48**, 4470-4472.
11. **Wu CC, Shryock TR, Lin TL, Faderan M, Veenhuizen MF.** Antimicrobial susceptibility of *Mycoplasma hyorhinis*. *Vet Microbiol* 2000, **76**, 25-30.