

Development of Nested PCR, Multiplex PCR, and Loop-Mediated Isothermal Amplification Assays for Rapid Detection of *Cylindrocladium scoparium* on Eucalyptus

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Eucalyptus dieback disease, caused by *Cylindrocladium scoparium*, has occurred in last few years in large Eucalyptus planting areas in China and other countries. Rapid, simple, and reliable diagnostic techniques are desired for the early detection of Eucalyptus dieback of *C. scoparium* prior to formulation of efficient control plan. For this purpose, three PCR-based methods of nested PCR, multiplex PCR, loop-mediated isothermal amplification (LAMP) were developed for detection of *C. scoparium* based on factor 1-alpha (tef1) and beta-tubulin gene in this study. All of the three methods showed highly specific to *C. scoparium*. The sensitivities of the nested PCR and LAMP were much higher than the multiplex PCR. The sensitivity of multiplex PCR was also higher than regular PCR. *C. scoparium* could be detected within 60 min from infected Eucalyptus plants by LAMP, while at least 2 h was needed by the rest two methods. Using different Eucalyptus tissues as samples for *C. scoparium* detection, all of the three PCR-based methods showed much better detection results than regular PCR. Base on the results from this study, we concluded that any of the three PCR-based methods could be used as diagnostic technology for the development of efficient strategies of Eucalyptus dieback disease control. Particularly, LAMP was the most practical method in field application because of its one-step and rapid reaction, simple operation, single-tube utilization, and simple visualiza-

tion of amplification products.

Keywords : *Cylindrocladium scoparium*, Eucalyptus, loop-mediated isothermal amplification, multiplex PCR, nested PCR

Eucalyptus was introduced into many countries worldwide including China due to its fast growth, high productivity and ever-increasing demand for paper, plywood, and fuel. Eucalyptus dieback disease is one of the most serious Eucalyptus diseases not only in China but also in many Eucalyptus plantation countries (Li et al., 2013). This disease is mainly caused by fungal pathogens of several species in *Cylindrocladium* (teleomorph: *Calonectria*) genus. For example, *C. clavatum* and *C. quinqueseptatum* were identified as the pathogens of Eucalyptus dieback in India (Sharma and Mohanan, 1982). *Cylindrocladium* and *Cylindrocladiella* spp. were found to be the pathogen of Eucalyptus dieback in Africa (Crous et al., 1993). According to a report by Crous and Wingfield (1994), *C. quinqueseptatum* caused Eucalyptus dieback in Australia, America and Brazil, and *C. scoparium* also caused this disease in Spain, Thailand, Australia, and Brazil. Previous studies indicated that *C. scoparium* is one of the most important pathogens of Eucalyptus dieback and caused disease on Eucalyptus frequently in most parts of Eucalyptus planting areas in China (Zhu et al., 2011a). It was estimated that the Eucalyptus planting area reached more than 10 million hm² in China, and 60% of which suffered Eucalyptus dieback infections in a certain degree. Eucalyptus dieback caused by *C. scoparium* has been defined as a key national plant quarantine disease in China (Pang et al., 2001). *C. scoparium* usually infects Eucalyptus seedlings and young Eucalyptus trees under 4 years old. In early stage of infection, this disease can easily lead to reddish-brown spots on leaves and shoots,

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leaves wilting, and branches dropping, which develop upwards from the base of young trees (Zhu et al., 2011b). In late stage of infection, it can cause large damage to the growth of Eucalyptus, result in branch wither, and lead to crown dieback or tree death depending on the severity of the disease (Li et al., 2013).

For a long time, the widest applied detection technique for Eucalyptus dieback disease is direct observation (Anonymus, 1984). However, this detection method by human judgment requires professional persons who have solid theoretical knowledge, and it is intensive time and labor consuming. Moreover, this method particularly has low levels of sensitivity and specificity, and it is hard to distinguish *C. scoparium* from other *Cylindrocladium* species. Therefore, it is not suitable for large scale diagnosis and surveillance. Although accurate detection of *C. scoparium* on Eucalyptus dieback can depend on pathogen isolation and classification by morphological, biological and genetic characteristics, it usually takes a long time to achieve a final result. To develop efficient strategies of fungal disease control as early as possible, a rapid and reliable detection technique is desired to accurately distinguish Eucalyptus dieback caused by *C. scoparium* from other *Cylindrocladium* species. However, such technique for detection of *C. scoparium* on Eucalyptus dieback has

not been established till now. In recent years, some rapid molecular detection techniques, such as nested PCR, multiplex PCR and loop-mediated isothermal amplification (LAMP) have been successfully developed for detection of fungal diseases of plants (Grote et al., 2002; Guglielmo et al., 2007; Hansen et al., 2016).

The objective of this study was to develop a rapid, simple, easy operating, sensitive and specific method for detection of *C. scoparium* in most Eucalyptus materials. Here, we reported for the first time that three PCR-based methods, including nested PCR, multiplex PCR, and LAMP, were developed for detection of *C. scoparium* based on factor 1-alpha (tefl) and beta-tubulin gene. In particular, the sensitivity, specificity and field application of these new methods for detection of Eucalyptus dieback disease caused by *C. scoparium* were assessed.

Materials and Methods

Strains and culture conditions. The 22 fungal strains used in this study were listed in Table 1, including 10 *Cylindrocladium* isolates (6 isolates from *C. scoparium* and 4 isolates from other species in *Cylindrocladium* genus), and 12 none-*Cylindrocladium* isolates obtained from diseased leaves and stem tissues caused by Eucalyptus

Table 1. Fungal strains used in this study for comparisons in polymerase chain reaction tests

No.	Species	Host plant	Sources
1	<i>Cylindrocladium scoparium</i>	<i>Eucalyptus</i>	Sichuan Province
2	<i>C. scoparium</i>	<i>Eucalyptus</i>	Guangxi Province
3	<i>C. scoparium</i>	<i>Eucalyptus</i>	Fujian Province
4	<i>C. scoparium</i>	<i>Eucalyptus</i>	Guizhou Province
5	<i>C. scoparium</i>	<i>Eucalyptus</i>	Chongqing Province
6	<i>C. scoparium</i>	<i>Eucalyptus</i>	Guangdong Province
7	<i>C.colhounii</i>	<i>Vaccinium Vitis-Idaea</i>	From Forest Protection Institution of Chinese Academy
8	<i>C. gracile</i>	<i>Paeonia suffruticosa</i>	From Forest Protection Institution of Chinese Academy
9	<i>C. parasiticum</i>	<i>Castanea mollissima</i>	From Forest Protection Institution of Chinese Academy
10	<i>C. quinquesepatum</i>	<i>Hevea brasiliensis</i>	From Forest Protection Institution of Chinese Academy
11	<i>Colletotrichum gloeosporioides</i>	<i>Eucalyptus</i>	Key Laboratory of Forest Protection of Sichuan Province
12	<i>Septoria mortarlensis</i>	<i>Eucalyptus</i>	Key Laboratory of Forest Protection of Sichuan Province
13	<i>Botryosphaeria dothidea</i>	<i>Eucalyptus</i>	Key Laboratory of Forest Protection of Sichuan Province
14	<i>Pseudocercospora chiangmaiensis</i>	<i>Eucalyptus</i>	Key Laboratory of Forest Protection of Sichuan Province
15	<i>Phyllosticta lindericola</i>	<i>Eucalyptus</i>	Key Laboratory of Forest Protection of Sichuan Province
16	<i>Diaporthe</i> sp.	<i>Eucalyptus</i>	Isolated from Eucalyptus tissues
17	<i>Gibberella</i> sp.	<i>Eucalyptus</i>	Isolated from Eucalyptus tissues
18	<i>Hypocrea</i> sp.	<i>Eucalyptus</i>	Isolated from Eucalyptus tissues
19	<i>Alternaria</i> sp.	<i>Eucalyptus</i>	Isolated from Eucalyptus tissues
20	<i>Nigrospora</i> sp.	<i>Eucalyptus</i>	Isolated from Eucalyptus tissues
21	<i>Fusicoccum</i> sp.	<i>Eucalyptus</i>	Isolated from Eucalyptus tissues
22	<i>Biscogniauxia</i> sp.	<i>Eucalyptus</i>	Isolated from Eucalyptus tissues

tus dieback, or healthy plants. These isolates were either available from Forest Protection of Chinese Academy (Beijing, China) or isolated and identified by sequencing and BLAST analyses of 26s and internal transcribed spacer (ITS), and deposited in the culture collection of Key Laboratory of Forest Protection of Sichuan Province, Sichuan, China. All isolates were stored on potato dextrose agar (PDA) medium covered by plum-extract agar with liquid paraffin at 4°C in the dark. Fresh cultures of isolates were prepared by placing small agar plugs from stock cultures in deionized water and then subculturing on PDA grown at 25°C. After 7-day incubation in dark, the mycelium was collected for DNA extraction or immediately stored at -20°C until use.

DNA extraction. Fungal mycelium and plant materials were ground and homogenized in liquid nitrogen using a mortar and pestle. The DNA was extracted from fungal mycelium and plant samples using a Plant Genomic DNA Extraction Kit (Tiangen Biotech Co., Ltd., Beijing, China) according to the manufacturer's instructions. The

DNA concentrations and purities were determined by a NanoDrop 2000C Spectrophotometer (Thermo Fisher Scientific, Wilmington, DE, USA) and further verified by agarose gel electrophoresis.

Fungal verification and primer design. All isolates used in this study were verified by amplification and sequencing of fungal ITS region using universal primer pair of ITS1 and ITS4 (Table 2) (White et al., 1990). Conventional PCR reaction was carried out in 50 µl of total volume consisting of 10 mM Tris-HCl (pH 8.0), 50 mM KCl₂, 1.5 mM MgCl₂, 0.3 mM each dNTP, 0.3 µM each ITS1 primer and ITS4 primer, 1.5 U Taq DNA polymerase (New England Biolabs, Beverly, MA, USA), and 4 µl of DNA template. The amplification protocol included an initial denaturation at 95°C for 5 min, followed by 30 cycles at 95°C for 30 s (denaturing), 55°C for 45 s (annealing) and 72°C for 90 s (elongation), and a final elongation at 72°C for 5 min. The PCR reaction was run on a Mastercycler Personal PCR Thermocycler (Eppendorf, Hamburg, Germany). All the PCR amplifications in this study were the

Table 2. Primer sequence, genes utilized in design, expected product size and source

Primers	Sense	Gene	Product size (bp)	Sequence (5'-3')	References
ITS1	Forward	ITS	Around 500	TCCGTAGGTGAACCTGCGC	White et al., 1990
ITS4	Reverse			TCCTCCGCTTATTGATATGC	
BT-T1	Forward	Beta-tubulin gene	Around 500	AACATGCGTGAGATTGTAAGT	Hagemann and Rose, 1988
BT-CYLTUBIR-A	Reverse			AGTTGTCGGGACGGAAGAG	Hagemann and Rose, 1988
EF1-728F	Forward	Factor 1-alpha (tef1)	Around 500	CATCGAGAAGTTCGAGAAGG	
EF2	Reverse			GGA(G/A)GTACCAGT(G/C)-ATCATGTT	
BT-S-1	Forward	Beta-tubulin gene	148	GGCTCCAAGAACTATGTGA	This study
BT-A-1	Reverse			CCTAACCACGAATGTCAGT	This study
EF-S-4	Forward	Factor 1-alpha (tef1)	272	CAAGAGTCGGATGGAATCAA	
EF-A-4	Reverse			CACAGGAGGTCGTCAAAC	This study
BT-S-9	Forward	Beta-tubulin gene	157	TGCGTAAAGTGCTCATTCTG	
BT-A-9	Reverse			AACTGGAGGTCGGAGGTA	This study
F3	Forward outer	Beta-tubulin gene		CTCGACAGCAATGGTGTCT	
B3	Backward outer			GCTCAAGATCGACGAGGAC	
FIP	Forward inner			TGCGCGCTCTTTCGCTAC-ATACACAACGGTACCTCCGACC	
BIP	Backward inner			CGCTCACCTGCGAGAAAC-AGCGCGAGGAACATACTTGTT	
LF	Loop forward			GTAGACGTTTCATGCGCTCC	
LB	Loop backward			ATTATTTGTGAGTGTGATAG	

the LAMP products, including visualization under ultraviolet (UV) light, on 2% agarose gel electrophoresis, and by fluorescent detection stained with SYBR Green I (Sigma-Aldrich).

Specificity and sensitivity of PCR primers. To evaluate the specificity of the designed primers, 22 DNA samples, including 6 DNA samples of *C. scoparium*, 4 samples from the same genus, and 12 samples from other 12 fungal genera (Table 1), were tested for nested-PCR, multiplex PCR, and LAMP assays. The sensitivities of nested-PCR, multiplex PCR, and LAMP were comparatively studied in this research. Briefly, the fungal DNA from pure culture of *C. scoparium* was used to make 10-fold serial dilutions (from 50 ng/μl to 500 ag/μl) to evaluate the sensitivity of these three assays in comparison to standard PCR assay. Regular PCR primed by primers of BT-S-1/BT-A-1 were also carried out for comparative analysis. All of these three assays were conducted as described above on the serial dilution of DNA.

Detection of Eucalyptus samples. To evaluate the reliability of these three assays as a diagnosis system, we extracted the DNA samples from Eucalyptus plant in cultivation district, on June 2014. The collected samples included the fresh dead tissues, diseased tissues with typical symptoms, tissues infected at early stage, infected tissues, residues of dead plant, and healthy tissues. All these samples were tested by nested-PCR, multiplex PCR, and LAMP assay, respectively.

Results

Evaluation of DNA quality. The concentrations of extracted fungal DNA from 22 fungal isolates were from 532 ng/μl to 675 ng/μl, and the ratios of A260/A280 of all the DNA samples were from 1.78 to 1.89. The quality of the extracted fungal DNA was further evaluated by amplification of ITS region using 50 ng of each extracted DNA as template and fungal universal primers of ITS1 and ITS4. Five microliters of PCR products were used for agarose gel electrophoresis and the PCR products amplified from all the tested isolates displayed a single and bright band in each lane (Fig. 4), indicating the DNA quality was good enough for subsequent PCR reactions.

Specificity of designed primers. Using 50 ng of each extracted fungal DNA as template, an amplification product of the expected size of 148 bp was obtained with DNA from all the *C. scoparium* isolates by nested PCR (Fig. 5). Meanwhile, no amplification product was obtained from the negative control. Moreover, no amplification product

was obtained with DNA from the isolates of other *Cylindrocladium* species and other genera tested as template in this study. Sequencing analysis indicated that the PCR product was the partial sequence of the beta-tubulin gene from *C. scoparium*. These results indicated that the designed primers for nested PCR are highly specific to *C. scoparium*.

Optimization analysis of the annealing temperature of the multiplex method indicated that two clear bands with expected size (272 bp and 157 bp) were presented in the same lane on the electrophoresis gel at 58°C of annealing temperature (Fig. 6A). Good presentation of the two expected bands on the electrophoresis gel also indicated that there was no competition between the two primer pair for multiplex PCR amplification. Therefore, this temperature was used as annealing temperature of multiplex methods for later study and application. Specificity study of the designed primers for multiplex method showed that two amplification products were obtained with DNA from all the *C. scoparium* isolates as template (Fig. 6B). Like the nested PCR, no amplification product was obtained without DNA from the negative control, with DNA from the isolates of other *Cylindrocladium* species and other genera tested as template by multiplex PCR in this study. Sequencing analysis indicated that the two PCR products were the expected partial sequence of the beta-tubulin gene and *tef1* gene from *C. scoparium*. These results indicated that the designed primers (EF-S-4/EF-A-4 and BT-S-9/BT-A-9) for multiplex PCR are highly specific to *C. scoparium*.

Optimal reaction for the LAMP assay with the primers sets designed in this study was found to be at 65°C

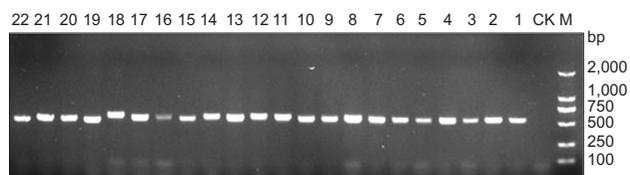


Fig. 4. Internal transcribed spacer (ITS) amplification results of 22 fungal isolates using universal primers of ITS1 and ITS4. Lane M: 2 kb DNA marker, Lane CK: negative control, Lanes 1–22: strains in the same order as listed in Table 1.



Fig. 5. Specificity test of the primers for nested PCR. Lane M: 2 kb DNA marker, Lane CK: negative control, Lanes 1–22: strains in the same order as listed in Table 1.

for 45 min and ended at 90°C for 5 min. Using this optimized LAMP procedure, specificity of the primers for the LAMP was evaluated. Positive results were observed in LAMP assays with DNA from all the *C. scoparium* isolates as template under UV light, on agarose gel electrophoresis and by SYBR Green I fluorescent stain (Fig. 7). However, the LAMP reactions showed negative results without DNA from the negative control, with DNA from the isolates of other *Cylindrocladium* species and other genera tested as template by all of the three visualization methods in this study. Sequencing analysis indicated that the DNA products from LAMP assay were the expected partial sequence of the beta-tubulin gene from *C. scoparium*. All of these evidences indicated that the designed primers (F3/B3, FIP/BIP, and LF/LB) for LAMP assay are highly specific to *C. scoparium*.

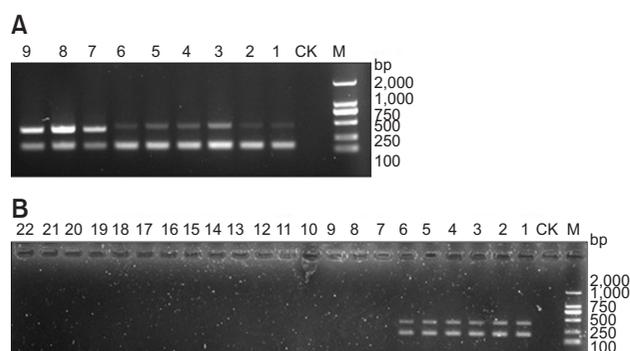


Fig. 6. Optimization results of annealing temperature of multiplex PCR and specificity test of the primers for multiplex PCR. (A) The results of multiplex PCR using primers EF-S-4/EF-A-4 and BT-S-9/BT-A-9 at a series of gradient annealing temperatures. Lane M: 2 kb DNA marker, Lane CK: negative control, Lanes 1–9: annealing temperature at 51°C, 52°C, 53°C, 54°C, 55°C, 56°C, 57°C, 58°C, and 59°C, respectively. (B) Specificity test of the primers for multiplex PCR. Lane M: 2 kb DNA marker, Lane CK: negative control, Lanes 1–22: strains in the same order as listed in Table 1.

Sensitivity of nested PCR, multiplex PCR, and LAMP.

Sensitivity of nested PCR, multiplex PCR, and LAMP were detected on a serial 10-fold dilution of pure fungal DNA of *C. scoparium* (Table 3, Fig. 8). The nested PCR and LAMP showed much higher sensitivity than the multiplex PCR developed in this study. The detection limit of these two PCR-based methods was approximately 5 fg/ μ l of pure fungal DNA (Fig. 8A, C), which was much higher than the regular PCR (5 pg/ μ l) (Fig. 8D) by 1,000 folds. Although the sensitivity of the multiplex PCR (50 fg/ μ l) (Fig. 8B) was lower than that of the nested PCR and LAMP, it was 10-fold higher than the regular PCR (Fig. 8D).

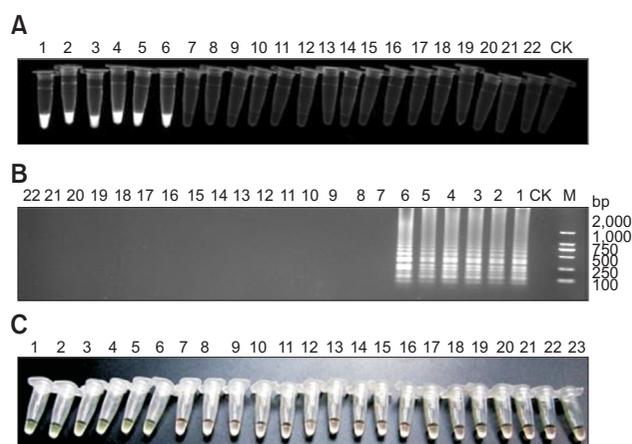


Fig. 7. Specificity test of the primers for loop-mediated isothermal amplification (LAMP) assay. (A) Reaction products were visualized under ultraviolet light. Lane CK: negative control, Lanes 1–22: strains in the same order as listed in Table 1. (B) Reaction products were visualized on agarose gel electrophoresis. Lane M: 2 kb DNA marker, Lane CK: negative control, Lanes 1–22: strains in the same order as listed in Table 1. (C) Reaction products were visualized by fluorescent detection stained with SYBR Green I and determined by color. Lanes 1–22: strains in the same order as listed in Table 1, Lane 23: negative control. Green or light green of the reaction solution indicates positive and orange or yellow indicates negative.

Table 3. Detection sensitivity of nested PCR, multiplex PCR, LAMP, and simple PCR on a serial dilution of pure fungal DNA of *C. scoparium* by 10 fold as templates

Method	Concentration of template DNA								
	50 ng/ μ l	5 ng/ μ l	500 pg/ μ l	50 pg/ μ l	5 pg/ μ l	500 fg/ μ l	50 fg/ μ l	5 fg/ μ l	500 ag/ μ l
Nested PCR	+	+	+	+	+	+	+	+	–
Multiplex PCR	+	+	+	+	+	+	–	–	–
LAMP	+	+	+	+	+	+	+	+	–
Regular PCR	+	+	+	+	+	–	–	–	–

LAMP, loop-mediated isothermal amplification; +, positive; –, negative.

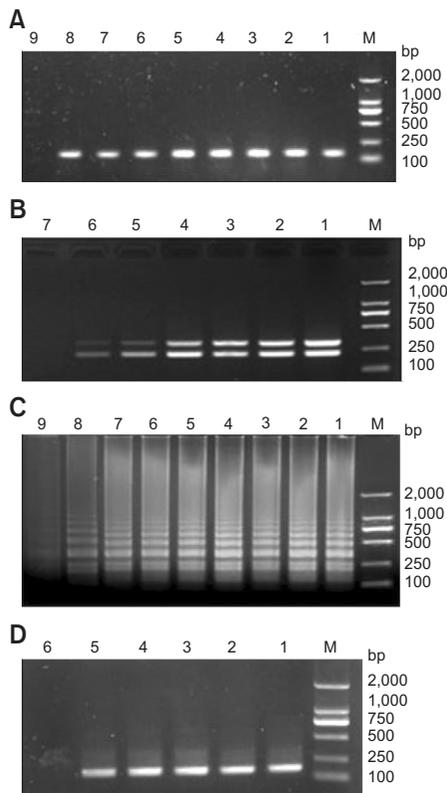


Fig. 8. Sensitivity of nested PCR, multiplex PCR, and loop-mediated isothermal amplification (LAMP) assays. (A) Sensitivity of nested PCR. Lane M: 2 kb DNA marker, Lanes 1–9: PCR products with a serial 10-fold dilution of fungal DNA of *Cylindrocladium scoparium* (from 50 ng/ μ l to 500 μ g/ μ l) as template. (B) Sensitivity of multiplex PCR. Lane M: 2 kb DNA marker, Lanes 1–7: PCR products with a serial 10-fold dilution of fungal DNA of *C. scoparium* (from 50 ng/ μ l to 5 fg/ μ l) as template. (C) Sensitivity of LAMP. Lane M: 2 kb DNA marker, Lanes 1–9: PCR products with a serial 10-fold dilution of fungal DNA of *C. scoparium* (from 50 ng/ μ l to 50 fg/ μ l) as template. (D) Sensitivity of regular PCR. Lane M: 2 kb DNA marker, Lanes 1–6: PCR products with a serial 10-fold dilution of fungal DNA of *C. scoparium* (from 50 ng/ μ l to 500 fg/ μ l) as template.

Detection of Eucalyptus samples. Using regular PCR with primers of BT-S-1/BT-A-1, *C. scoparium* was detected in fresh dead Eucalyptus tissues and diseased tissues with typical symptoms (Fig. 9A). In comparison, *C. scoparium* was detected not only in the two tissues mentioned above but also in tissues infected at early stage, infected tissues, and residues of dead plant by nested PCR and LAMP (Fig. 9A, C). Similar to nested PCR, *C. scoparium* was also detected in most tissues mentioned above except for some samples from infected tissues by multiplex PCR (Fig. 9B). Considering LAMP assay's characteristics of one-step and rapid reaction, simple operation, single-tube utilization, and simple visualization of

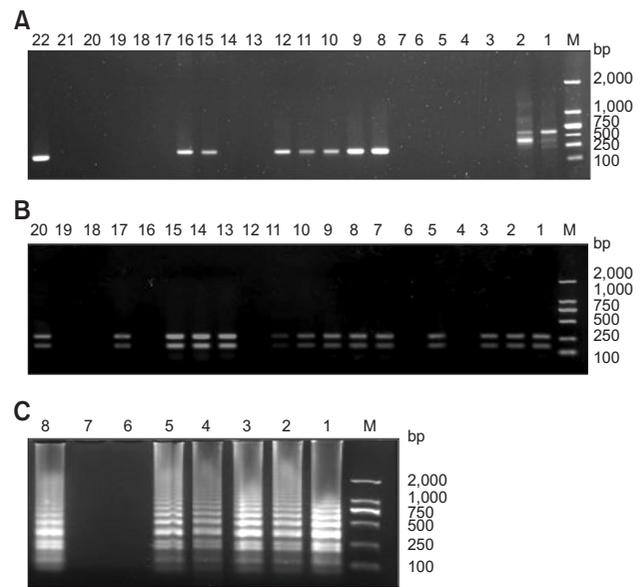


Fig. 9. Detection of *Cylindrocladium scoparium* on Eucalyptus samples by nested PCR, multiplex PCR, and loop-mediated isothermal amplification (LAMP) assays. (A) Detection of *C. scoparium* on Eucalyptus samples by nested PCR. Lane M: 2 kb DNA marker, Lanes 1–7: PCR products with primers of BT-T1/BT-CYLTUBIR-A for the first round amplification, Lanes 8–14: nested PCR products with primers of BT-S-1/BT-A-1 for the second round amplification, Lanes 15–21: PCR products with primers of BT-S-1/BT-A-1, Lane 22: positive control. Lanes 1, 8, 15: fresh dead tissues, Lanes 2, 9, 16: diseased tissues with typical symptoms, Lanes 3, 10, 17: tissues infected at early stage, Lanes 4, 11, 18: infected tissues, Lanes 5, 12, 19: residues of dead plant, Lanes 6, 13, 20: healthy tissues, Lanes 7, 14, 21: negative control. (B) Detection of *C. scoparium* on Eucalyptus samples by multiplex PCR. Lane M: 2 kb DNA marker, Lanes 1, 7, 13: fresh dead tissues, Lanes 2, 8, 14: diseased tissues with typical symptoms, Lanes 3, 9, 15: tissues infected at early stage, Lanes 4, 10, 16: infected tissues, Lanes 5, 11, 17: residues of dead plant, Lanes 6, 12, 18: healthy tissues, Lane 19: negative control, Lane 20: positive control. (C) Detection of *C. scoparium* on Eucalyptus samples by LAMP. Lane M: 2 kb DNA marker, Lane 1: fresh dead tissues, Lane 2: diseased tissues with typical symptoms, Lane 3: tissues infected at early stage, Lane 4: infected tissues, Lane 5: residues of dead plant, Lane 6: healthy tissues, Lane 7: negative control, Lane 8: positive control.

amplification products, it was the most practical method for detection of *C. scoparium*.

Discussion

Eucalyptus dieback can be caused by several rather than one species in *Cylindrocladium* genus (Crous et al., 1993). It is very important to accurately detect the pathogen of dieback for later efficient control plan-making. Therefore, specificity of the detection method should be

considered first in disease diagnosis (Rees-George et al., 2010). To well distinguish *C. scoparium* from other species in the same *Cylindrocladium* genus, representative isolates from four other species (*C. colhounii*, *C. gracile*, *C. parasiticum*, and *C. quinqueseptatum*) in *Cylindrocladium* genus were used as test strains in this study to evaluate whether the new developed nested PCR, multiplex PCR and LAMP are specific to *C. scoparium*. In addition, strains from 12 fungal genera usually isolated from Eucalyptus were included in this study. Some of them are pathogens to Eucalyptus, causing anthrax, peliosis, canker, grey speck, or round spot symptom, which undoubtedly will influence diagnostic results by direct observation. Therefore, using as many as possible strains of different species in the same genus and different genera from Eucalyptus as controls in the study could make the diagnosis of *C. scoparium* more accurate. To eliminate the influence of geographic variations on diagnosis by nested PCR, multiplex PCR and LAMP, we used six *C. scoparium* strains isolated from Sichuan, Guangxi, Fujian, Guizhou, Chongqing, and Guangdong provinces in China as representative strains.

A strongly conservative and species-specific sequence is the key factor to design primers successfully (Bonants et al., 1997). Beta-tubulin gene (also called β -tubulin gene), was widely used in the classification, identification, and phylogenetic analysis of fungi, bacteria and actinomycetes etc., due to its well specificity (Tan et al., 2009). Similarly, factor 1-alpha gene, existed both in prokaryotes and eukaryotes, was also proved to be specific to species (Huang et al., 2007). In this paper, we designed the nested PCR and LAMP primers based on the sequence of beta-tubulin gene, and multiplex PCR primer based on the sequences of both beta-tubulin gene and factor 1-alpha gene. Specificity studies established that all the primers used for nested PCR, multiplex PCR and LAMP were well species-specific to *C. scoparium*. In general, multiplex PCR is used to diagnose various species of pathogens. However, we developed this method for detection of *C. scoparium* in order to improve the diagnostic specificity in this study. Based on this foundation, a more complicated multiplex PCR for diagnosis of main species causing Eucalyptus dieback should be developed in further study.

The optimized reaction system was an essential prerequisite for the quantity of amplification products (Li et al., 2008). Previous study proved that an appropriate annealing temperature was as important as an optimized reaction system (Sipos et al., 2007). The annealing temperature of these three detection methods was optimized, respectively, in this study. The best temperature for nested PCR and multiplex PCR was at 58°C, while for LAMP it

was 65°C.

PCR-based techniques, such as regular PCR, nested PCR, multiplex PCR, and LAMP, have been widely used for diagnosis and monitoring of plant diseases caused by microorganisms (Feng et al., 2012; Grote et al., 2002; Henson and French, 1993; Wang et al., 2010). Compared with traditional or conventional method (such as direct observation, pathogen isolation and identification, etc.) for detection of *C. scoparium*, the PCR-based methods possess some advantage characteristics of rapid, simple, reliable detection. However, there was no report of PCR-based diagnostic techniques covering the nested PCR, multiplex PCR, and LAMP on detection of *C. scoparium*. In this study, we developed these three methods for detection of one kind of pathogen at the same time, and compared them to regular PCR and within themselves using the same research materials and laboratory conditions. The comparative results based on data from such conditions are more reliable than other reported results based on data from different research groups and geographic regions. In this study, we found that the sensitivities of the nested PCR and LAMP were similar and higher than multiplex PCR, and the latter was also higher than regular PCR. Using different Eucalyptus tissues as samples for *C. scoparium* detection, all of the three PCR-based methods showed much better detection results than regular PCR. Considering the good sensitivity, the nested PCR and LAMP developed in this study are the priority selection for the specific detection of in laboratory diagnosis. Moreover, LAMP is the most practical method in field application due to its one-step and rapid reaction, simple operation, single-tube utilization, and simple visualization of amplification products. In order to achieve this goal of *C. scoparium* detection in field, a Rapid Fungi Genomic DNA Isolation Kit (Sangon Biotech Co., Ltd.), which lyse cells by heat, can be used for fungal DNA extraction, and a portable electric generator and a metal-bath heater need to be equipped.

In conclusion, any of the new developed nested PCR, multiplex PCR, and LAMP is a rapid, sensitive, and reliable diagnostic method for detection of *C. scoparium* on Eucalyptus dieback disease. All of these three PCR-based methods can be used as valuable tools in the indoor diagnosis of Eucalyptus dieback disease caused by *C. scoparium*, and LAMP can also be used as valuable tool in outdoor diagnosis. This study can contribute to the efficient prevention and control of Eucalyptus dieback disease caused by *C. scoparium*.

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References

- Anonymus. 1984. Nursery diseases of Eucalyptus in Kerala and their possible control measures. *Kerala For. Res. Inst. Inf. Bull.* 6:16.
- Bonants, P., Hagenaar-de Weerd, M., van Gent-Pelzer, M., Lacourt, I., Cooke, D. and Duncan, J. 1997. Detection and identification of *Phytophthora fragariae* Hickman by the polymerase chain reaction. *Eur. J. Plant Pathol.* 103:345-355.
- Crous, P. W., Phillips, A. J. L. and Wingfield, M. J. 1993. New records of *Cylindrocladium* and *Cylindrocladiella* spp. in South Africa. *Plant Pathol.* 42:302-305.
- Crous, P. W. and Wingfield, M. J. 1994. A monograph of *Cylindrocladium*, including anamorphs of *Calonectria*. *Mycotaxon* 51:341-435.
- Feng, X., Zhang, X. and Yan, D. 2012. Detection to pathogens of poplar cankers by the multiplex PCR technique. *Scientia Silvae Sinicae* 48:72-77 (in Chinese).
- Glass, N. L. and Donaldson, G. C. 1995. Development of primer sets designed for use with the PCR to amplify conserved genes from filamentous ascomycetes. *Appl. Environ. Microbiol.* 61:1323-1330.
- Grote, D., Olmos, A., Kofoet, A., Tuset, J. J., Bertolini, E. and Cambra, M. 2002. Specific and sensitive detection of *Phytophthora nicotianaeanae* by simple and nested-PCR. *Eur. J. Plant Pathol.* 108:197-207.
- Guglielmo, F., Bergemann, S. E., Gonthier, P., Nicolotti, G. and Garbelotto, M. 2007. A multiplex PCR-based method for the detection and early identification of wood rotting fungi in standing trees. *J. Appl. Microbiol.* 103:1490-1507.
- Hagemann, G. D. and Rose, P. D. 1988. Leaf spot and blight on *Acacia longifolia* caused by *Cylindrocladium scoparium*. *Phytophylactica* 20:311-316.
- Hansen, Z. R., Knaus, B. J., Tabima, J. F., Press, C. M., Judelson, H. S., Grünwald, N. J. and Smart, C. D. 2016. Loop-mediated isothermal amplification for detection of the tomato and potato late blight pathogen, *Phytophthora infestans*. *J. Appl. Microbiol.* 120:1010-1020.
- Henson, J. M. and French, R. 1993. The polymerase chain reaction and plant disease diagnosis. *Annu. Rev. Phytopathol.* 31:81-109.
- Huang, J., Song, D., Flores, A., Zhao, Q., Mooney, S. M., Shaw, L. M. and Lee, F. S. 2007. IOP1, a novel hydrogenase-like protein that modulates hypoxia-inducible factor-1 α activity. *Biochem. J.* 401:341-352.
- Li, G., Wu, Z., Xie, Y., Li, T., Chang, R., Wang, Y. and Zhou, X. 2013. Research progress of Eucalyptus dieback in China and abroad. *Eucalypt Sci. Technol.* 1:45-50 (in Chinese).
- Li, W., Li, D., Twieg, E., Hartung, J. S. and Levy, L. 2008. Optimized quantification of unculturable *Candidatus liberibacter* spp. in host plants using real-time PCR. *Plant Dis.* 92:854-861.
- Mori, Y. and Notomi, T. 2009. Loop-mediated isothermal amplification (LAMP): a rapid, accurate, and cost-effective diagnostic method for infectious diseases. *J. Infect. Chemother.* 15:62-69.
- O'Donnell, K. and Cigelnik, E. 1997. Two divergent intragenomic rDNA ITS2 types within a monophyletic lineage of the fungus *Fusarium* are nonorthologous. *Mol. Phylogenet. Evol.* 7:103-116.
- Pang, L. D., Pang, W. W., Zeng, W. Q., Liu, Z. C., Wei, Z. Z. and Tan, G. B. 2001. Chemical control of Eucalyptus dieback. *Plant Quar.* 15:273-275 (in Chinese).
- Rees-George, J., Vanneste, J. L., Cornish, D. A., Pushparajah, I. P. S., Yu, J., Templeton, M. D. and Everett, K. R. 2010. Detection of *Pseudomonas syringae* pv. *actinidiae* using polymerase chain reaction (PCR) primers based on the 16S-23S rDNA intertranscribed spacer region and comparison with PCR primers based on other gene regions. *Plant Pathol.* 59:453-464.
- Sharma, J. K. and Mohanan, C. 1982. *Cylindrocladium* spp. associated with various diseases of Eucalyptus in Kerala. *Eur. J. For. Pathol.* 12:129-136.
- Sipos, R., Székely, A. J., Palatinszky, M., Révész, S., Márialigeti, K. and Nikolauz, M. 2007. Effect of primer mismatch, annealing temperature and PCR cycle number on 16S rRNA gene-targeting bacterial community analysis. *FEMS Microbiol. Ecol.* 60:341-350.
- Tan, Z., Tong, X., Fang, C., Jiang, C. and Chen, C. 2009. Prokaryotic expression of α -tubulin gene of *Camellia sinensis* and preparation of α -tubulin polyclonal antibody. *J. Tea Sci.* 29:336-340 (in Chinese).
- Tomlinson, J. A., Dickinson, M. J. and Boonham, N. 2010. Detection of *Botrytis cinerea* by loop-mediated isothermal amplification. *Lett. Appl. Microbiol.* 51:650-657.
- Wang, Z., Yang, M., Yang, Y. Q., Peng, Z. K. and Zhou, E. X. 2010. Morphological and molecular identification of citrus anthracnose pathogen from Guangdong province. *Mycosystema* 29:488-493 (in Chinese).
- White, T. J., Bruns, T., Lee, S. and Taylor, J. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: *PCR protocols: a guide to methods and applications*, eds. by M. A. Innis, D. H. Gelfand, J. J. Sninsky and T. J. White, pp. 315-322. Academic Press, New York, NY, USA.
- Zhu, J. H., Chen, H. M., Wu, J. Q., Chen, Q. Z., Feng, L. Z., Meng, X. M. and Guo, W. S. 2011a. Loss estimation of eucalyptus growth caused by of eucalyptus dieback. *For. Pest Dis.* 30:6-10 (in Chinese).
- Zhu, J. H., Chen, H. M., Wu, J. Q., Chen, Q. Z., Feng, L. Z., Meng, X. M. and Guo, W. S. 2011b. Study on the control technology of *Cylindrocladium quinqueseptatum*. *J. Fujian For. Sci. Technol.* 38:41-47 (in Chinese).