

Original Article

Journal of Acupuncture Research

Journal homepage: https://www.e-jar.org

Establishment and Selection of Indicator Materials for *Cervi Parvum Cornu* Pharmacopuncture



Kyu-Jin Yang ¹, Ki-Beom Lee ¹, No-Hyeon Kim ¹, Tae-Gyu Kim ¹, Yu-Mi Gi ¹, Hwan-Soo Joo ¹, Chang-Yong Suh ¹, In-Hee Lee ², Hwa-Jin Chung ², In-Hyuk Ha ², Jae-Woong Lee ^{2,*}

¹ Jaseng Hospital of Korean Medicine, Seoul, Korea
² Jaseng Medical Foundation, Seoul, Korea

ARTICLE INFO	ABSTRACT	
<i>Article history:</i> Submitted: July 10, 2017 Revised: August 3, 2017 Accepted: August 22, 2017	Background: Recently, <i>Cervi Parvum Cornu</i> pharmacopuncture has been widely used. But no studies on the indicator materials for <i>Cervi Parvum Cornu</i> pharmacopuncture have been conducted. The aim of this studies was to select indicator materials that would aid in the uniform preparation of standardized <i>Cervi Parvu Cornu</i> pharmacopuncture.	
<i>Keywords:</i> alanine, <i>Cervi Parvum Cornu</i> , chondroitin sulfate, leucine, pharmacopuncture, standardization	the same methods in <i>Cervi Parvum Cornu</i> pharmacopulcture were analysed. Each lot was prepared using the same methods and materials. Chondroitin sulfate, alanine, and leucine were selected as the indicator materials for <i>Cervi Parvum Cornu</i> . For standardization, chondroitin sulfate analysis was performed using the colorimetric method, while alanine and leucine were analyzed using liquid chromatography-mass spectrometry (LC–MS).	
	Results: Analysis of the three lots of <i>Cervi Parvum Cornu</i> pharmacopuncture found chondroitin sulfate levels of 108.9 ± 17.3 ug/ml, 118.8 ± 5.0 ug/ml and 112.3 ± 11.9 ug/ml. Alanine levels were 44.9 ± 2.8 ug/ml, 44.6 ± 0.3 ug/ml, and 43.9 ± 0.2 ug/ml. Leucine levels were 29.6 ± 0.7 ug/ml, 29.0 ± 0.1 ug/ml, and 29.4 ± 0.1 ug/ml. Conclusion: These results suggest that chondroitin sulfate, alanine, and leucine may be useful for the standardization of <i>Cervi Parvum Cornu</i> pharmacopuncture.	
	©2017 Korean Acupuncture & Moxibustion Medicine Society. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).	

Introduction

Cervi Parvum Cornu is the dried deer velvet from the unboned horns of young bucks. Depending on the section, the bone is classified as tip, upper section, midsection, or base. There are many amino acids, proteins, lipids, polyamines, and saccharides in the tip of *Cervi Parvum Cornu*. The midsection and the base are ossified, and the amount of the substances mentioned above is small.

In oriental medicine, the nature of *Cervi Parvum Cornu* is warm, the taste is sweet and salty, and it enters the kidney and liver. It has purported efficacy in nephrotic syndrome, hyperplasia, hypertrophy, and stiff muscles. Traditionally, it has been used for impotence, lassitude of spirit, fear of cold, dizziness, tinnitus, deafness, vaginal discharge, and lumbar vertebrae disease. Recent studies reported that *Cervi Parvum Cornu* seemed to have effects on accelerating body weight and neuromuscular development, reducing muscle fatigue, improving musculoskeletal function, promoting neural cell growth, increasing bone strength and weight, and improving arthritis symptoms, immune function, anticancer function, and wound healing [1–3].

In recent years, the use of *Cervi Parvum Cornu* pharmacopuncture based on acupuncture, meridians, and herbalism has increased among oriental medicine doctors. In *Cervi Parvum Cornu* pharmacopuncture, extracts of *Cervi Parvum Cornu* obtained by distillation are injected into tender points or acupuncture points for an expected result similar to that from acupuncture and drug treatment.

Several studies of *Cervi Parvum Cornu* pharmacopuncture have reported it to be effective in reducing the symptom severity of arthritis [4–14] and osteoporosis [15–19], in aging [20,21] and growth disorders [22,23]. Other studies have shown that *Cervi*

*Corresponding author

https://doi.org/10.13045/jar.2017.02180 pISSN 2586-288X eISSN 2586-2898

Jaseng Medical Foundation, 858, Unjuro, Gangnam-gu, Seoul, Korea E-mail: darkpret@naver.com

^{©2017} Korean Acupuncture & Moxibustion Medicine Society, Published by E-Tree Publishing. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Parvum Cornu pharmacopuncture increased levels of antioxidant enzymes [20,21], promoted hormone production [24], helped to regulate the autonomic nervous system [25,26], had anticancer effects [27,28], and enhanced immune function [29].

The clinical application of *Cervi Parvum Cornu* pharmacopuncture is also increasing and related research is actively underway. However, there is a lack of consideration of *Cervi Parvum Cornu* pharmacopuncture formulation and preparation. There is also risk that the active components decompose or are removed during preparation and purification. There is insufficient standardization for the process of *Cervi Parvum Cornu* pharmacopuncture. It is necessary to set the indicator materials for the quality control and standardization of *Cervi Parvum Cornu* pharmacopuncture. Therefore, chondroitin sulfate, alanine and leucine—which are easy to analyze without precipitation during the preparation process of *Cervi Parvum Cornu* pharmacopuncture. This study was conducted to establish the method of analyzing the indices for standardization.

Materials and Methods

Instruments and reagents

Glucuronolactone, alanine and leucine were purchased from Sigma-Aldrich Corp. (St. Louis, MO, USA); their purity were 99%, 98% and 98.5%, respectively. Carbazole used in the pretreatment was purchased from Daejung Chemical and Metal Co. Ltd. (Shiheung City, Gyeonggi-do, South Korea). Sodium borate and trifluoroacetic acid were purchased from Junsei Chemical Co. Ltd. (Tokyo, Japan). Anhydrous ethanol and sulfuric acid were purchased from J.T. Baker (Phillipsburg, NJ, USA), as were the water and acetonitrile (ACN; HPLC grade) used for quantitative analysis. Liquid chromatography-mass spectrometry (LC–MS) was used for component analysis (LCMS-2020; Shimadzu Corp., Kyoto, Japan). Sunrise RC from TECAN (Männedorf, Switzerland) was used for absorbance measurement.

How to prepare Cervi Parvum Cornu pharmacopuncture

The Cervi Parvum Cornu pharmacopuncture used in the analysis was from Namyangju outpatient department of Jaseng Oriental Medicine Hospital, Korea, which had been prepared according to Korean Traditional Medicine Standardization Project guidelines on pharmacopuncture. Cervi Parvum Cornu (400 g) was put through an extraction process using purified water at 105°C for 2 hours, for a total of seven times. Thereafter, it was concentrated under reduced pressure at 38°C. The concentrated extract was stirred with 90% ethyl alcohol seven times. The solution was filtered to remove the fibrous substance. The filtrate was adjusted to have an alcohol concentration of 70% using ethyl alcohol and filtered again. The filtrate was concentrated under reduced pressure at 38°C, and the extract was diluted with purified water, then sterilizing filtrated and lyophilized. The lyophilized powder was diluted to a concentration of 2 mg/ml in a clean room. The pH of the diluted solution was adjusted to 7.0-7.4, and the salinity to 0.9%. After this process, the pharmacopuncture was sterilized at 121°C for 25 minutes. It was then ready to be used in the clinic (Fig. 1).

Setting the indicator materials of Cervi Parvum Cornu pharmacopuncture

In order to establish suitable indicator materials for Cervi Par-



Fig.1. Manufacturing process of Cervi Parvum Cornu pharmacopuncture.



Fig. 2. Structural formula of the index components of *Cervi Parvum Cornu* pharmacopuncture: (A) chondroitin sulfate; (B) alanine; (C) leucine.

vum Cornu pharmacopuncture, we looked at the literature. Various components such as glucosamine sulfate, chondroitin sulfate, prostaglandins, collagen, proteoglycans, amino acids and linoleic acid were identified [30,31]. We looked for components that would be easy to analyze without sedimentation in the preparation process. Chondroitin sulfate, alanine and leucine were chosen as indicator materials (Fig. 2). Chondroitin sulfate is a polysaccharide composed of N-acetylgalactosamine, uronic acid and sulfuric acid, which are the main components of cartilage. Chondroitin sulfate is also found in various connective tissues such as skin and umbilical cord, and is related to the osteolysis of cartilage. Alanine can be easily metabolized with lactic acid and is involved in producing glucose from protein through the alanine circuit. Leucine is an amino acid that makes up muscles. It is involved in blood sugar regulation, bone and muscle tissue growth, hormone production,

healing of damaged tissue, and energy generation.

Analysis of chondroitin sulfate levels

The level of chondroitin sulfate in Cervi Parvum Cornu pharmacopuncture was analyzed by the colorimetric method which is used in the Health Functional Food Test[32]. The analytical sample was obtained by filtering the Cervi Parvum Cornu pharmacopuncture with a 0.2 µm polyvinylidene difluoride syringe filter. There are three groups : blank (water, reference solution), standard solution, test solution. The standard solution was glucuronolactone at a concentration of 20 µg/ml. After adding 5 ml of sodium borate sulfate solution to the colorless tube and cooling thoroughly with ice water, 1 ml of the test solution and 1 ml of the standard solution were carefully added into the tube separately. It was then heated for 10 minutes and immediately cooled with ice water. Then, 0.2 ml of carbazole solution was added and mixed separately; the mixture was heated for 15 minutes and then cooled with ice water. 1 ml of water was used as a blank in the same manner as above. Absorbance was measured at a wavelength of 530 nm and the level of chondroitin sulfate was determined according to the following equation.



2.593 = Molecular weight of chondroitin sulfate / Molecular weight of glucuronic acid

1.1023 = Molecular weight of glucuronic acid / Molecular weight of glucuronolactone

Conditions for analysis of alanine and leucine

Analysis of alanine and leucine was based on a previously described method [33]. We developed a simultaneous method that could detect both indicator materials at the same time using LC– MS. The column used for the analysis was Shim-pack XR-ODS

Table 1. Composition of the Mobile Phase Employed in the Gradient LC System

Time (min)	Composition of mobile phase(%)		
	0.1% TFA in Water	0.1% TFA in Acetonitrile	
0	95	5	
19	36	64	
20	95	5	
30	95	5	

Analytes	Condition
Interface	ESI(+)
Nebulizer gas flow	1.5
DL temp (°C)	250
Heat block temp ($^{\circ}C$)	400
Dry gas flow (L/min)	15
selected ion monitoring(SIM)mode(m/z)	Alanine : 90, Leucine : 132



Fig. 3. Chondroitin sulfate levels in the three lots of *Cervi Parvum Cornu* pharmacopuncture.

(3.5 µm, 4.6 I.D. × 150 mm; Shimadzu Corp.). Temperature was maintained at 35°C, the injection volume was 1 µl, and the flow rate was 0.4 ml/min. (A) 0.1% trifluoroacetic acid in water and (B) 0.1% trifluoroacetic acid in acetonitrile were degassed using ultrasound and analyzed by gradient system (Table 1). Electrospray ionization–mass spectrometry was used for the detector, and the detection conditions are shown in Table 2.

Results

Chondroitin sulfate levels in Cervi Parvum Cornu pharmacopuncture

When the three lots of *Cervi Parvum Cornu* pharmacopuncture prepared in the Namyangju outpatient department of Jaseng Oriental Hospital were analyzed, chondroitin sulfate levels were 108.9 \pm 17.3 µg/ml, 118.8 \pm 5.0 µg/ml, and 112.3 \pm 11.9 µg/ml, respectively (Fig. 3).

Analysis of alanine and leucine levels in Cervi Parvum Cornu pharmacopuncture

The coefficient of correlation between the calibration curve and linearity was calculated by analyzing the indicator materials, alanine and leucine, at five concentrations each (5, 10, 50, 100, 500 µg/ml). For alanine, a calibration curve Y = 2241X + 152,447was obtained, while for leucine, a calibration curve Y = 66,583X+ 155,241 was obtained. Both were in the range of 5–500 µg/ml, and linearity was obtained with a correlation coefficient of 0.999 or more.

After the three lots of *Cervi Parvum Cornu* pharmacopuncture prepared in the Namyangju outpatient department of Jaseng Oriental Hospital were analyzed, the results were added to the calibration curves. Alanine levels were found to be $44.9 \pm 2.8 \mu g/ml$, $44.6 \pm 0.3 \mu g/ml$, and $43.9 \pm 0.2 \mu g/ml$, while leucine levels were found to be $29.6 \pm 0.7 \mu g/ml$, $29.0 \pm 0.1 \mu g/ml$, and $29.4 \pm 0.1 \mu g/ml$ (Figs. 4 & 5).

Discussion

Cervi Parvum Cornu is the cut and dried horn of young bucks, including Cervus nippon Temminck, Cervus elaphus Linné, and Cervus canadensis Erxleben. The horns of the young bucks are not



Fig.4. Liquid chromatography-mass spectrometry chromatogram of *Cervi Parvum Cornu* pharmacopuncture and standard compounds: (A) alanine standard (SIM mode: m/z 90 [M+H]+); (B) leucine standard (SIM mode: m/z 132 [M+H]+); (C) *Cervi Parvum Cornu* pharmacopuncture (SIM mode: m/z 90, 132).



Fig. 5. Alanine and leucine levels in the three lots of *Cervi Parvum Cornu* pharmacopuncture.

ossified or slightly osseous, and are hairy. Because it is in a state of growth, there are various growth factors present. For example, it contains many components such as amino acids, proteins, lipids, polyamines, and sugars [34].

Donguibogam is a book that describes the effects of *Cervi Parvum Cornu* and prescriptions containing it. *Cervi Parvum Cornu* was believed to be efficacious in harmonizing the thoroughfare and conception vessels, so it was used in the treatment of dizziness, tinnitus, deafness, vaginal discharge, spermatorrhea, profuse menstruation, uterine bleeding, and hemophilia. It was believed to have a kidney yang-tonifying effect for the treatment of symptoms such as impotence, seminal emission, enuresis, lassitude of spirit, fear of cold, lumbar vertebrae disease, and consumptive disease. It was even used to treat delayed development of pediatric age or weak bone and muscle because of its apparent action on the musculoskeletal system. It also appeared to be effective in neutralizing sores and ulcers, so it was used to promote ulcer and wound healing, as well as to strengthen immune function.

Studies on components that incorporate *Cervi Parvum Cornu* have found that they contain various cell growth factors like vascular endothelial growth factor, epidermal growth factor, neuron growth factor, fibroblast growth factor, and insulin-like growth factor (IGF-1, IGF-2) [35,36]. Minerals, amino acids, polypeptides, proteins [37], fatty acids [38], and various polysaccharides are also present.

To utilize these substances within *Cervi Parvum Cornu* more efficiently and conveniently, oriental medicine doctors make use of *Cervi Parvum Cornu* pharmacopuncture. It has been reported that *Cervi Parvum Cornu* pharmacopuncture can be applied to arthritis [4–14], osteoporosis [15–19], growth disorder [22,23], and the symptoms of aging [20,21]. In addition, some studies have shown that *Cervi Parvum Cornu* pharmacopuncture has the effects of antioxidant enzymes [20,21], can promote hormone production [24], regulate the autonomic nervous system [25,26], and has anticancer activity [27,28].

As clinical use of *Cervi Parvum Cornu* pharmacopuncture increases and studies of it are actively proceeding, it has become clear that research on its formulation is insufficient and a standardization process is lacking. There is also a risk that components may be degraded or removed during the preparation and purification of *Cervi Parvum Cornu* pharmacopuncture. Therefore, it was considered necessary to select indicator materials that would aid in the uniform preparation of standardized *Cervi Parvum Cornu* pharmacopuncture and quality control. Chondroitin sulfate, alanine and leucine, which are easy to analyze without precipitation during the preparation of *Cervi Parvum Cornu* pharmacopuncture, were found to be suitable indicator materials.

Chondroitin sulfate is a suitable indicator as it has been acknowledged to be present at high levels and demonstrate good activity in *Cervi Parvum Cornu* according to the Health Functional Food Test method of the Korea Food and Drug Administration.

Though *Cervi Parvum Cornu* contains many kinds of amino acids, alanine and leucine were selected because they can be analyzed simultaneously, which means that it is easy to compare the patterns. The linearity showed a correlation coefficient of over 0.999 in the concentration range of 5–500 μ g/ml. Therefore, it was confirmed that both alanine and leucine were analyzed within the normal range. Using molecular weight, the specific substance is analyzed, so experimental error due to interference from other peaks can be minimized. Indeed, it was confirmed that there was no interference with other peaks.

Conclusion

Chondroitin sulfate, alanine, and leucine are three substances that are found in high concentrations in *Cervi Parvum Cornu* pharmacopuncture and that are easy to analyze, making them suitable indicator materials. The results of this study suggest that they are useful for the standardization of *Cervi Parvum Cornu* pharmacopuncture so that the reliability of this treatment may be improved.

Conflicts of Interest

The authors have no conflicts of interest to declare.

Acknowledgments

This study was supported by the Korea Herbal Medicine Foundation Service Project (Standardization Project of Herbal Medicine for Pharmacopuncture–Development of Manufacturing Process of Herbal Medicine and Quality of Controlling the Herbal Medicine for Pharmacopuncture).

References

- Moreau M, Dupuis J, Bonneau NH, Lécuyer M. Clinical evaluation of a powder of quality elk velvet antler for the treatment of osteoarthrosis in dogs. Can Vet J 2004;45:133–139.
- [2] Mikler JR, Theoret CL, High JC. Effects of topical elk velvet antler on cutaneous wound healing in streptozotocin induced diabetic rats. J Altern Complement Med 2004;10:835–840.
- [3] You L, Zhao M, Regenstein JM, Ren J. In vitro antioxidant activity and in vivo anti-fatigue effect of loach (Misgurnusanguillicaudatus) peptides prepared by papain digestion. Food Chem 2011;124:188–194.
- [4] Lee HJ, Cho HS, Hwang MS, Jung CY, Lee DG, Kim EJ, et al. Effect of Cervi Pantotrichum Cornu pharmacopuncture on suppressing the expression of iNOS and production of NO in type-II collagen induced arthritis mice. The Acupuncture 2008;25:105–116.
- [5] Chung YR, Lee SD, Byun H, Park IS, Jung CY, Lee CH, et al. The effect of deer antler herbal acupuncture control to hyper-inflammatory responses on synovial membrane by LPS-induced arthritis. The Acupuncture 2007;24:167–181.
- [6] Kim EJ, Kim GY, Chung HW. Effect of Achyrantis Radix administration and Cervi Cornu Parvum acupuncture in experimental osteoarthritis rats. Korean J Oriental Physiol Pathol 2007;21:1194–1199.
- [7] Ahn HJ, Kim KS. Inhibitory effect of deer antler aqua-acupuncture (DAA) on cathepsin S activity and rheumatoid arthritis in rats. The Acupuncture 2003;20:104–116.
- [8] Kim JG, Kim KS. Inhibitory effects of Cervi Pantotrichum Cornu herbal acupuncture on type-II collagen-induced arthritis. The Acupuncture 2002;19:155–170.
- [9] Choi YH, Choi WS, Song IK, Park JS, Lee SD, Kim KS. Protective and anti-arthritic effects of Cervi Pantotrichum Cornu herbal acupuncture, inhibiting dihydroorotate dehydrogenase, on phosphate-ion-mediated chondrocyte apoptosis and rat collagen-induced arthritis. The Acupuncture 2002;19:10–27.
- [10] Choi BJ, Kim MJ, Park SD, Lee AR, Jang JH, Kim KH. Modulation of bone mass, strength and turnover by a Cervi Pantotrichum Cornu herbal acupuncture in adjuvant-induced arthritic rats. The Acupuncture 2002;19:219–233.
- [11] Park SD, Kim MJ, Lee AR, Jang JH, Kim KH. Effect of Cervi Pantotrichum Cornu herbal acupuncture on protease activities, antioxidant in rheumatoid arthritis rats. The Acupuncture 2002;19:51–64.
- [12] Hwang JS, Hwang JH, Lee HJ, Lee DG, Kang MJ, Back SO, et al. The ability of Cervus Elaphus Sibiricus herbal acupuncture to inhibit the generation of inflammatory enzymes on collagen induced arthritis mice. The Acupuncture 2007;24:1–14.
- [13] Kim JK, Lee SD, Jeong YR, Kim KS. Protective action of cartilage and bone destruction by deer antler herbal-acupuncture solution, the pilose antler of Cervus Korean TEMMINCK Var. Mantchuricus Swinhoe, on Type-II collagen-induced arthritis in mice. The Acupuncture 2006;23:73–90.
- [14] Kim WY, Lee SD, Kim KH, Baek ST, Kim KS. The efficiency of deer antler herbal acupuncture on modulation and prevention of IL-1 mediated activation in rat chondrocytes at a receptor level. The Acupuncture 2006;23:113–123.
- [15] Kim JK, Kim KS. Effects of the pilose antler of Cervus Korean TEMMINCK var. mantchuricus Swinhoe (DAS), herbal acupuncture solution on suppression of collagenolysis and bone resorption in mouse calvarial

osteoblasts. The Acupuncture 2004;21:225-236.

- [16] Kim MJ, Lee SD, Kim KH, Byun H, Kim KS. Effects of deer antler water extract (pilose antler of Cervus Korean TEMMINCK var. mantchuricus Sinhoe) on chondrocytes. The Acupuncture 2006;23:103–111.
- [17] Han SW, Lee YH, Kim CH. A study on effects of the Cervi Pantotriculum Cornu herb-acupuncture on the osteoporosis induced by ovariectomy in rats. J Pharmacopuncture 2000;3:177–191.
- [18] Yook TH, Lee CH, Lee HI. A study on the effects of the Carthamisemen, Cervi Pantotrichum Cornu, Hominis placenta aquacupuncture on the osteoporosis in rats. The Acupuncture 2001;18:61–75.
- [19] Han SW, Choi JY, Lee YH. Healing of bony defects by Cervi Pantotriculum Cornu herbal acupuncture. The Acupuncture 2001;18:135–146.
- [20] Lee KB, Park SK. Effects of Cornu Cervi Parvum pharmacopuncture on the blood picture and antioxidative activity in rats. Korean J Acupunct 2010;27:25–34.
- [21] Lee JH, Lee KM, Kim JS. Anti-wrinkle effects of Cervi Pantotrichum Cornu pharmacopuncture solution. The Acupuncture 2010;27:1–8.
- [22] Lee JM, Kim YT, Lee HI, Son YS, Jin SH, Lee HS, et al. The effects of Cervus elaphus aquacupuncture and Gingseng radix aquacupuncture on the growth of animals. J Pharmacopuncture 2000;3:131–152.
- [23] Kim YT, Son YS, Jin SH, Han SW, Shim IS, Lim SBN, et al. The effects of Cervus elaphus on the growth and the intellectual development of animals. The Acupuncture 2001;18:122–134.
- [24] Kwak DG, Yang CH. A regulatory effect of Cervi Cornu Parvum aquaacupuncture on serum estradiol level after ovariectomy. The Acupuncture 1998;15:29–41.
- [25] Lee HY, Lee JB, Cho YH, Song BY, Yook TH. The effects of Cervi Pantotrichum Cornu pharmacopuncture and Bovis Calculus•FelUrsi pharmacopuncture on the heart rate variability. The Acupuncture 2010;27:65–74.
- [26] Kim HJ, Song BY, Yook TH. The effects of distilled Cervi Pantotrichum Cornu pharmacopuncture and Zzizyphispinosi semen pharmacopuncture on the heart rate variability. J Pharmacopuncture 2009;12:31–40.
- [27] Ryu SH, Lee KM, Lee BH, Lim SC, Jun TY, Seo JC. Oligonucleotide chip analysis of Cervi Parvum Cornu herbal-acupuncture solution (CPC-HAS) on SNU484 carcinoma cells. Korean J Acupunct 2006;23:137–148.
- [28] Ryu SH, Lee KM, Lee BH, Lim SC, Jung TY, Seo JC. DNA and proteomic expression of Cervi Parvum Cornu herbal-acupuncture solution (CPC-HAS) in HepG2 carcinoma cells. J Pharmacopuncture 2006;9:5–16.
- [29] Kim YW, Moon JY, Lim JK, Park WH, Park SD, Nam KS. Effects of CCP (Cervi Cornu Parvum) aqua-acupuncture on glucocorticoid-Induced immunosuppression in murine lymphocyte. The Acupuncture 1997;14:245– 252.
- [30] Jo SJ, Kim JH, Kim JW, Choi HO, Lee SH, Kim MK, et al. Comparative studies on velvet deer antler and ossified deer antler on the contents of bioactive components and on the bone mineral density improving activity for oophorectomized rat. Nat Prod Sci 2013;19:303–310.
- [31] Lee BY, Lee OH, Choi HS. Analysis of food components of Korean deer antler parts. Korean J Food Sci Technol 2003;35:52–56.
- [32] Korea Food and Drug Administration. Health Functional Food Act. 2008;3:6–10.
- [33] Park SW, Lee JW, Lee JH, Ha IH, Byun JH, Jung BH, et al. Identification of standard compound of Ja-ha-guh pharmacopuncture and validation of analytic methods. JORM 2016;26:33–40.
- [34] Lu L, Wang K, Li L, Xuan Z, Gong X. Effect of velvet antler polypeptide on peripheral nerve regeneration. Zhongguo Xiu Fu Chong Jian Wai Ke Za Zhi 2008;22:1458–1461. [In Chinese]
- [35] Clark DE, Lord EA, Suttie JM. Expression of VEGF and pleiotrophin in deer antler. Anat Rec A Discov Mol Cell Evol Biol 2006;288:1281–1293.
- [36] Barling PM, Lai AK, Nicholson LF. Distribution of EGF and its receptor in growing red deer antler. Cell Biol Int 2005;29:229–236.
- [37] Je JY, Park PJ, Lim DH, Jeon BT, Kho KH, Ahn CB. Antioxidant, antiacetylcholinesterase and composition of biochemical components of Russian deer velvet antler extracts. Korean J Food Sci Anim Resour 2011;31:349–355.
- [38] Ivankina NF, Isay SV, Busarova NG, Mischenko TYa. Prostaglandin-like activity, fatty-acid and phospholipid composition of sika deer (Cervus nippon) antlers at different growth stages. Comp Biochem Physiol B 1993;106:159-162.