

Short Communication

Association between 14bp Insertion/Deletion Polymorphism in Exon 8 of *HLA-G* gene and Oral Squamous Cell Carcinoma in Korean Population

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Abnormal *HLA-G* expression occurs in various diseases such as melanoma, renal cell carcinoma, asthma, and classic Hodgkin's lymphoma. The purpose of this study was to determine whether *HLA-G* gene is linked with oral squamous cell carcinoma (OSCC). To investigate the possible link with susceptibility to OSCC, 54 OSCC patients and 120 healthy controls were enrolled in this study. *HLA-G* 14bp insertion/deletion polymorphism is in 3'-untranslated region of *HLA-G* gene. *HLA-G* 14bp insertion/deletion polymorphism was analyzed using the polymerase chain reaction (PCR) method. For the analysis of genetic data, SPSS18.0 program was used. Logistic regression models were performed for odds ratio (OR), 95 percent confidence interval (CI), and *P* value. There was a significant difference in distribution allele between OSCC patients and control subjects (OR=0.018, 95% CI=0.002-0.131, $p<0.001$). Our results suggest that *HLA-G* 14bp insertion/deletion polymorphism may be linked with susceptibility to OSCC in the Korean population.

Key words: association study, oral squamous cell carcinoma, OSCC, *HLA-G* 14bp polymorphism

Introduction

The 1.4% of total cancer developed in Korea is oral cancer and oral squamous cell carcinoma (OSCC) consists of more than 90% of malignancy in oral cavity (National Cancer Information Center, Republic of Korea, <http://www.cancer.go.kr>).

Human leukocyte antigen-G (*HLA-G*) is a non-classical HLA class I molecule [1]. Many studies showed that *HLA-G* involved in the immuno suppressive response and long-term immune escape or tolerance [2]. Level of *HLA-G* is also significantly higher in patients with various tumors such as malignant melanoma, glioma, breast cancer, ovarian cancer, hepatocellular carcinoma and so on [3-6]. One of the characteristics of the *HLA-G* gene is that there is a limited genetic polymorphism. The most common polymorphism of *HLA-G* gene is a 14 bp (5'-ATTGTTTCATGCCT-3') insertion /deletion. It is located at the 3' UTR region (exon 8) of the *HLA-G* gene. Several studies suggested that it was associated with a risk factor for various disease including cancer, diabetics, and autoimmune diseases [7]. *HLA-G* polymorphism is evaluated with nasopharyngeal carcinoma risk [8], transitional cell carcinoma of the bladder in a Brazilian population [9], and it also could be a marker for genetic susceptibility to hepatocellular carcinoma in Chinese populations [10]. Although many studies have been conducted, the relationship between 14 bp (5'-ATTGTTTCATGCCT-3')

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insertion /deletion of *HLA-G* gene and oral cancer has not been studied yet.

In this study, we evaluated whether the 14 bp (5'-ATTT GTTCATGCCT-3') insertion /deletion of *HLA-G* gene was contributed to susceptibility of OSCC in Korean population.

Materials and Methods

Participants.

This study was approved by the Institutional Review Board of School of Dentistry, Kyung Hee University, Seoul, Republic of Korea (20040915). The OSCC subjects included 54 patients and 120 healthy controls were enrolled in this study. We reviewed the biopsy chart of department of oral pathology, Kyung Hee University and selected the OSCC cases. We found total 92 OSCC cases but some tissues were too small or some samples were not available to extract DNA. Clinical features of experimental subjects were shown in Table 1. Control subjects were selected through general health check-up program that they had no clinical evidence of OSCC or any other severe disorders.

DNA extraction and genotyping assays.

Genomic DNA from OSCC samples was prepared from

Table 1. Clinical features of experimental subjects diagnosed as OSCC.

Number		54
Sex	Male/female	43/11
Age	Average±SD	61.3±11.6
Location		
	anterior region of the maxilla	1
	posterior region of the maxilla	10
	anterior region of the mandible	3
	posterior region of the mandible	16
	lower lip	2
	mouth floor	5
	salivary gland	1
	hard palate	5
	soft palate	5
	sublingual	1
	tongue	5

SD, standard deviation

paraffin embedded block using a Qiagen® DNA Micro kit (Qiagen) and stored at -20°C before use. Genomic DNA from control subjects was prepared from peripheral blood using a genomic DNA isolation reagent kit (High Pure PCR template preparation kit, Roche, USA). Genotyping for 14bp insertion/deletion polymorphism in exon 8 (3'UTR) of the *HLA-G* gene was performed by polymerase chain reaction (PCR) method using the primers antisense 5'-GGAAGGAA TGCAGTTCAGCATGA-3' and sense 5'-GTGATGGGCTGTTT AAAGTGTCACC-3' (fluorescent labeled). PCR method was carried out basically as described by Tripathi et al. using the following cycling profile: 94°C for 5 minutes, and 39 cycles at 94°C for 30 seconds, 58°C for 40 seconds, and 72°C for 1 minutes[11]. The PCR products were analyzed by gene scan (3730xl DNA Analyzer, Applied Biosystems, USA).

Statistical analysis.

We used SPSS 18.0 for analyzing genetic data between OSCC and control subjects. Logistic regression model was used for odd ratio (OR), 95% confidence interval (CI), and *p* value. The level of significance was set at 0.05.

Results

In the present study, we examined whether 14bp insertion/deletion polymorphism in exon 8 (3'UTR) of the *HLA-G* gene has a relation with susceptibility to OSCC. No deviation from Hardy-Weinberg equilibrium was found in the control group ($p>0.05$). Table 2 shows genotypic and allelic frequencies of *HLA-G* 14bp insertion/deletion polymorphism among OSCC and control subjects. We observed a significant difference in genotype and allelic frequencies between OSCC and control subjects (OR=0.017, 95% CI=0.002-0.129, $p<0.001$, OR=0.019, 95% CI=0.003-0.139, $p<0.001$, respectively). Genotypic frequencies 210bp/210bp, 210bp/224bp, and 224bp/224bp were 98.1%, 1.9% and 0.0% versus (*vs*) 43.3%, 47.5%, and 9.2% (OSCC subjects *vs* control subjects). The allelic frequencies of 210bp and 224bp were 99.1% and 0.9% *vs* 67.1% and 32.9%, respectively. Frequency of 224bp allele in the OSCC group was significantly decreased compared with the control group. There were significant differences between OSCC and control subjects ($p<0.05$).

Table 2. Genotypic and allelic frequencies of *HLA-G* 14bp insertion/deletion polymorphism between oral squamous cell carcinoma (OSCC) and control subjects.

Genotype/ Allele	Control	OSCC		OR	95 % CI	p
	n, (%)	n, (%)				
Genotype						
210bp/210bp	52 (43.3%)	53 (98.1%)	Codominant 1	0.017	0.002-0.129	0.000077
210bp/224bp	57 (47.5%)	1 (1.9%)	Codominant 2	NA	NA	NA
224bp/224bp	11 (9.2%)	0 (0.0%)	Dominant	0.014	0.002-0.108	0.000036
			Recessive	NA	NA	NA
Allele						
210bp	161 (67.1%)	107 (99.1%)				
224bp	79 (32.9%)	1 (0.9%)		0.019	0.003-0.139	0.000094

OR and 95% CI were from logistic regression analyses.

OR, odds ratio; CI, confidence interval; n, number of subjects; NA, not applicable.

Discussion

HLA-G is a non classical tolerogenic molecule and many previous reports showed its relationship with various immune reactions. *HLA-G* degrades the function of all immune cells, thus defects in HLA class I expression allow tumor cells to resist cytotoxic immune functions [12]. Immune cells that may be affected by *HLA-G* antigens include natural killer cells, CD8⁺ T cells, CD4⁺ T lymphocyte function, and dendritic cells, which may affect their cytotoxicity [13]. *HLA-G* is selectively expressed in the cytotrophoblast of the feto-maternal interface and plays an important role in protecting the fetus from maternal natural killer cells [14]. Therefore, there were reports that its polymorphism is related to abortion and failure of in vitro fertilization failure [15,16]. *HLA-G* also has a relationship with the infection. *HLA-G* polymorphism is associated with human papilloma virus infection and squamous intraepithelial lesions, which represents a profile of predisposition to cervical cancer in woman [17]. *HLA-G* polymorphism is known to be an estimated susceptibility factor for human cytomegalovirus infection in children [18] and human immunodeficiency virus vertical transmission [19,20]. As previously mentioned, *HLA-G* inhibits natural killer cell cytotoxicity in some tumors. Human melanoma cell secretes *HLA-G* [21] and this enable tumor cells to evade from immune surveillance of the host [12]. *HLA-G* interacts with *HLA-G*-recognizing killer-cell inhibitory receptors (KIRs) and inhibits host immune response in human breast cancer [22].

The 14 bp insertion/deletion polymorphism of *HLA-G* is a large, polymorphic promoter region, which appears to play an important role in the regulation of *HLA-G* expression [23]. The presence of the 14 bp insertion allele (+ 14 bp) destabilizes the mRNA and reduces *HLA-G* protein production. This suggests that differences in genotypes may cause differences in immune responses [13,24]. The 14 bp deletion polymorphism of the *HLA-G* gene also influences mRNA stability and plays an important role in the expression of soluble *HLA-G* in plasma [25].

In this study, we evaluated whether 14bp insertion/deletion polymorphism was associated with OSCC in Korean population. We identified genotypic and allelic frequencies of *HLA-G* in OSCC and control subjects. In control subjects, genotypic frequencies 210bp/210bp, 210bp/224bp, and 224bp/224bp were 43.3%, 47.5% and 9.2% and allelic frequencies of 210bp and 224bp were 67.1% and 32.9%, respectively. This result was similar to previous study. In Chinese, genotypic frequencies 210bp/210bp, 210bp/224bp, and 224bp/224bp were 37.3%, 46.6%, and 16.1% and allelic frequencies of 210bp and 224bp were 60.6% and 39.4%, respectively [26]. There were few differences in genotypic and allelic frequencies between normal Chinese and Korean population. However, genotypic frequencies 210bp/210bp, 210bp/224bp, and 224bp/224bp and allelic frequencies of 210bp and 224bp were completely different between OSCC and control subjects. In OSCC subjects, only one 224bp was found, and even there was no 224bp/224bp in OSCC subjects. This result suggests that 14bp insertion/deletion polymorphism of *HLA-G* might be associated with the

development of OSCC.

In conclusion, we found a significant association between OSCC patients and control subjects. Our results suggest that 14bp insertion/deletion polymorphism of *HLA-G* may be contributed to the susceptibility to OSCC in Korean population.

Conflict of Interest

The authors indicate that no potential conflict of interest exist.

References

1. Kamishikiryō, J, Maenaka K. HLA-G molecule. *Curr Pharm Des.* 2009;15:3318-3324.
2. Larsen, MH, Hviid TV. Human leukocyte antigen-G polymorphism in relation to expression, function, and disease. *Hum Immunol.* 2009;70:1026-1034. DOI: 10.1016/j.humimm.2009.07.015
3. Rebmann, V, Regel J, Stolke D, Grosse-Wilde H. Secretion of sHLA-G molecules in malignancies. *Semin Cancer Biol.* 2003;13:371-377.
4. Bezuhly, M, Howlett A, Colp P, Conrad DM, Walsh N, Rowden G, Morris SF, Langley RG. Quantitative HLA-G expression in metastasising and non-metastasising primary thin cutaneous melanomas. *Dermatology.* 2008;217:281-283. DOI: 10.1159/000150602
5. Kren, L, Muckova K, Lzicarova E, Sova M, Vybihal V, Svoboda T, Fadrus P, Smrcka M, Slaby O, Lakomy R, Vanhara P, Krenova Z, Michalek J. Production of immune-modulatory nonclassical molecules HLA-G and HLA-E by tumor infiltrating ameboid microglia/macrophages in glioblastomas: a role in innate immunity. *J Neuroimmunol.* 2010;220:131-135. DOI: 10.1016/j.jneuroim.2010.01.014
6. Elliott, RL, Jiang XP, Phillips JT, Barnett BG, Head JF. Human leukocyte antigen G expression in breast cancer: role in immunosuppression. *Cancer Biother Radiopharm.* 2011;26:153-157. DOI: 10.1089/cbr.2010.0924
7. Alvarez, M, Piedade J, Balseiro S, Ribas G, Regateiro F. HLA-G 3'-UTR SNP and 14-bp deletion polymorphisms in Portuguese and Guinea-Bissau populations. *Int J Immunogenet.* 2009;36:361-366. DOI: 10.1111/j.1744-313X.2009.00875.x
8. Ghandri, N, Gabbouj S, Farhat K, Bouaouina N, Abdelaziz H, Nouri A, Chouchane L, Hassen E. Association of HLA-G polymorphisms with nasopharyngeal carcinoma risk and clinical outcome. *Hum Immunol.* 2011;72:150-158. DOI: 10.1016/j.humimm.2010.10.006
9. Castelli, EC, Mendes-Junior CT, Viana de Camargo JL, Donadi EA. HLA-G polymorphism and transitional cell carcinoma of the bladder in a Brazilian population. *Tissue Antigens.* 2008;72:149-157. DOI: 10.1111/j.1399-0039.2008.01091.x
10. Jiang, Y, Chen S, Jia S, Zhu Z, Gao X, Dong D, Gao Y. Association of HLA-G 3' UTR 14-bp insertion/deletion polymorphism with hepatocellular carcinoma susceptibility in a Chinese population. *DNA Cell Biol.* 2011;30:1027-1032. DOI: 10.1089/dna.2011.1238
11. Tripathi, P, Abbas A, Naik S, Agrawal S. Role of 14-bp deletion in the HLA-G gene in the maintenance of pregnancy. *Tissue Antigens.* 2004;64:706-710. DOI: 10.1111/j.1399-0039.2004.00308.x
12. Rebmann, V, Wagner S, Grosse-Wilde H. HLA-G expression in malignant melanoma. *Semin Cancer Biol.* 2007;17:422-429. DOI: 10.1016/j.semcancer.2007.06.010
13. Baricordi, OR, Stignani M, Melchiorri L, Rizzo R. HLA-G and inflammatory diseases. *Inflamm Allergy Drug Targets.* 2008;7:67-74.
14. Rouas-Freiss, N, Goncalves RM, Menier C, Dausset J, Carosella ED. Direct evidence to support the role of HLA-G in protecting the fetus from maternal uterine natural killer cytotoxicity. *Proc Natl Acad Sci USA.* 1997;94:11520-11525.
15. Berger, DS, Hogge WA, Barmada MM, Ferrell RE. Comprehensive analysis of HLA-G: implications for recurrent spontaneous abortion. *Reprod Sci.* 2010;17:331-338. DOI: 10.1177/1933719109356802
16. Kano, T, Mori T, Furudono M, Ishikawa H, Watanabe H, Kikkawa E, Warita T, Onizuka M, Takahashi M, Maeda Y, Naruse T, Inoko H, Kimura A. Human leukocyte antigen may predict outcome of primary recurrent spontaneous abortion treated with paternal lymphocyte alloimmunization therapy. *Am J Reprod Immunol.* 2007;58:383-387. DOI: 10.1111/j.1600-0897.2007.00517.x
17. Simoes, RT, Goncalves MA, Castelli EC, Junior CM, Bettini JS, Discorde ML, Duarte G, Quintana SM, Simoes AL, Moreau P, Carosella ED, Soares EG, Donadi EA. HLA-G polymorphisms in women with squamous intraepithelial lesions harboring human papillomavirus. *Mod Pathol.* 2009;22:1075-1082. DOI: 10.1038/modpathol.2009.67
18. Zheng, XQ, Zhu F, Shi WW, Lin A, Yan WH. The HLA-G 14 bp insertion/deletion polymorphism is a putative susceptible factor for active human cytomegalovirus infection in children. *Tissue Antigens.* 2009;74:317-321. DOI: 10.1111/j.1399-0039.2009.01312.x
19. Segat, L, Catamo E, Fabris A, Padovan L, Morgutti M, Crovella S. HLA-G 3' UTR haplotypes and HIV vertical transmission. *AIDS.* 2009;23:1916-1918.
20. Fabris, A, Catamo E, Segat L, Morgutti M, Arraes LC, de Lima-Filho JL, Crovella S. Association between HLA-G 3'UTR 14-bp polymorphism and HIV vertical transmission in Brazilian children. *AIDS.* 2009;23:177-182.
21. Lesport, E, Baudhuin J, LeMaout J, Sousa S, Doliger C, Carosella ED, Favier B. Human melanoma cell secreting human leukocyte antigen-G5 inhibit natural killer cell cytotoxicity by impairing lytic granules polarization toward target cell. *Hum Immunol.* 2009;70:1000-1005. DOI: 10.1016/j.humimm.2009.07.019
22. Urosevic, M, Trojan A, Dummer R. HLA-G and its KIR ligands in cancer--another enigma yet to be solved. *J Pathol.*

- 2002;196:252-253. DOI: 10.1002/path.1057
23. Cordero, EA, Veit TD, da Silva MA, Jacques SM, Silla LM, Chies JA. HLA-G polymorphism influences the susceptibility to HCV infection in sickle cell disease patients. *Tissue Antigens*. 2009;74:308-313.
 24. Piancatelli, D, Maccarone D, Liberatore G, Parzanese I, Clemente K, Azzarone R, Pisani F, Famulari A, Papola F. HLA-G 14-bp insertion/deletion polymorphism in kidney transplant patients with metabolic complications. *Transplant Proc*. 2009;41:1187-1188. DOI: 10.1016/j.transproceed.2009.03.028
 25. Chen, XY, Yan WH, Lin A, Xu HH, Zhang JG, Wang XX. The 14 bp deletion polymorphisms in HLA-G gene play an important role in the expression of soluble HLA-G in plasma. *Tissue Antigens*. 2008;72:335-341. DOI: 10.1111/j.1399-0039.2008.01107.x
 26. Wu, FX, Wu LJ, Luo XY, Tang Z, Yang MH, Xie CM, Liu NT, Zhou JG, Guan JL, Yuan GH. Lack of association between HLA-G 14-bp polymorphism and systemic lupus erythematosus in a Han Chinese population. *Lupus*. 2009;18:1259-1266. DOI: 10.1177/0961203309345756