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Evaluation of Boldine Activity against Intracellular Amastigotes of *Leishmania amazonensis*

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Abstract: Leishmaniasis is a neglected and endemic disease that affects poorest population mainly in developing countries. A lack of adequate and definitive chemotherapeutic agents to fight against this infection has led to the investigation of numerous compounds. The aim of this study was to investigate in vitro activity of boldine against *Leishmania amazonensis* murine cell infection. Boldine ((S)-2,9-dihydroxy-1,10-dimethoxy-aporphine) is an aporphine alkaloid found abundantly in the leaves/bark of boldo (*Peumus boldus* Molina), a widely distributed tree native to Chile. The in vitro system consisted of murine macrophage infection with amastigotes of *L. amazonensis* treated with different concentrations from 50 to 600 µg/ml of boldine for 24 hr. Intracellular parasite destruction was assessed by morphological examination and boldine cytotoxicity to macrophages was tested by the MTT viability assay. When cells were treated with 100 µg/ml of boldine the reduction of parasite infection was 81% compared with untreated cultures cells. Interestingly, boldine-treatment caused a concentration-dependent decrease of macrophage infection that culminated with 96% of reduction when cells were submitted to 600 µg/ml of boldine. Cell cultures exposed to 100 µg/ml of boldine and 300 µg/ml of Glucantime[®] during 24 hr showed a significant reduction of 50% in parasitized cells compared with cell cultures exposed just to Glucantime[®]. The study showed that treatment with boldine produces a better effect than treatment with the reference antimonial drug, glucantime, in *L. amazonensis* infected macrophage. Our results suggest that boldine is a potentially useful agent for the treatment of leishmaniasis.

Key words: Leishmania amazonensis, anti-parasitic activity, boldo, neglected disease

Leishmaniasis is an endemic zoonosis caused by several species of the genus *Leishmania*, a cell obligate parasite, transmitted through the bite of an infected sandfly that can cause a range of symptoms from cutaneous to visceral lesions in humans [1]. Approximately 0.2-0.4 million cases of visceral leishmaniasis, and about 0.7-1.2 million of cases of cutaneous leishmaniasis are registered annually, demonstrating the importance of this disease [2].

The control strategy is based on sinantropic vectors combat, effective diagnosis, and chemotherapy. In fact, the development of new and cost-effective alternative therapeutic strategies for leishmaniasis has become a high priority [3], mainly due to their high toxicity and lack of efficacy in some cases of leishmaniasis [4-6].

© 2017, Korean Society for Parasitology and Tropical Medicine This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. In recent years, considerable attention has been given to studying plants in an attempt to search for new antileishmanial drugs [7]. Identification of natural products and the wide variety of separation techniques available could increase the probability of finding novel products that in turn could lead to the development of new pharmaceutical compounds [8]. However, novel therapeutics for diseases endemic to the third world would seem out of reach. The costs associated with creating, maintaining, and screening large compound libraries are high. Thus, the development of drugs for leishmaniasis is hampered by the general lack of interest by the pharmaceutical industry in this class of disease; less than 1% of all new drugs target neglected tropical diseases [9].

Therefore, a strategy based on the testing of drugs already in human use and originally designed and evaluated for nonrelated diseases can be another alternative [6]. Recently, tamoxifen, an antineoplastic agent for the prophylactic treatment of malignancies was tested against cutaneous leishmaniasis with promising results [10,11].

Boldine [(S)-2,9-dihydroxy-1,10-dimethoxy-aporphine] is

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an aporphine alkaloid found abundantly in the leaves/bark of boldo (*Peumus boldus* Molina), a widely distributed tree native to Chile [12]. Boldine has been reported to have several pharmacological activities, such as anti-inflammatory, antipyretic, antidiabetic, antiatherogenic, antiplatelet, antitumor promoting and cytoprotective effects [13]. Recently, a study showed that boldine increased NO bioavailability into endothelial cells and restored endothelial functions [14]. It has been well established that NO-dependent mechanisms are critical for control of *Leishmania* infection [15]. Thus, based on Boldine proprieties and aiming to identify a new antileishmanial agent, the present work was carried out to investigate the effect of boldine using in vitro leishmaniasis model.

Primary mouse macrophages were obtained from normal BALB/c mice by peritoneal lavage [16,17]. Leishmania amazonensis (MHOM/BR/73/M2269) amastigote forms were isolated from active skin lesions of BALB/c mice as described previously [16]. The parasites were suspended in RPMI 1640 medium and used immediately after isolation. Peritoneal macrophages were infected with L. amazonensis amastigotes (3:1 parasites/host cell) for 1 hr, as previously described [18]. After the interaction period, the cultures were washed to remove extracellular parasites, and fresh medium was added to the cell culture supplemented with 50 to 600 µg/ml of boldine or 300 µg/ml of Glucantime[®] (Sanofi-Aventis, Brazil), and incubated at 35°C, 5% of CO2 for 24 hr, as previously described [19]. Intracellular parasite destruction was assessed by morphological examinations. Briefly, for evaluating the percentage of infected macrophages and the number of amastigotes per macrophage, cells on coverslips were stained with Giemsa [17]. Intracellular amastigotes which were exclusively localized in parasitophorous vacuoles [20] were examined microscopically at a magnification of 1,000×. About 600 cells were counted per triplicate coverslip [21].

Boldine (Sigma-Aldrich, St. Louis, Missouri, USA) cytotoxicity to macrophages was tested by the MTT viability assay. Briefly, after incubation of peritoneal derived macrophages with 50 to 600 µg/ml of boldine for 24 hr, cells were also incubated with the highest concentration of DMSO used for boldine solubilization (0.01%). The formation of formazan was measured by adding 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT; Molecular Probes, Eugene, Oregon, USA) 0.5 mg/ml and incubation of the cultures at 37°C in the dark. After 4 hr, the medium was removed, 200 µl of DMSO was added per well, and the absorbance was measured using an ELISA reader at 540 nm (Labsystems Multiskan, Helsinki, Finland). All experiments were repeated at least 3 times, and the results are expressed as the mean \pm SD. Data obtained under different conditions were analyzed statistically by the Student's t-test, with levels of significance set at *P* ≤ 0.05.

Peritoneal macrophages were efficiently infected with *L. amazonensis* for 24 hr, showing a significantly high number of infected macrophages, 72%. Similar results were found with DMSO treated macrophages (Fig. 1A). In cell cultures subjected to 50 μ g/ml of boldine for 24 hr, there was a significant reduction of 50% of the infected macrophages. When cells were treated with 100 μ g/ml of boldine, the reduction was 81% compared with untreated cultures cells. Interestingly, boldine-treatment caused a concentration-dependent decrease of macrophage infection that culminated with 96% of reduction when cells were submitted to 600 μ g/ml of boldine. This performance was 4-fold higher than cells treated with 300 μ g/ml of Glucantime®, the first choice drug to treat leishmaniasis (Fig. 1A).

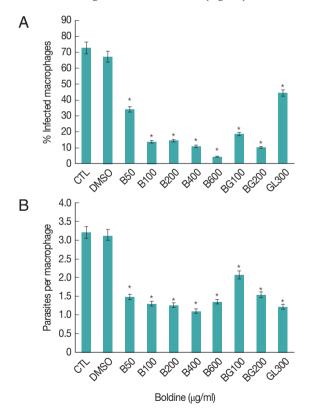


Fig. 1. Effects of boldine on *L. amazonensis* infected macrophages. Peritoneal macrophages were infected with *L. amazonensis* amastigotes and cultured at 37°C and 5% of CO₂ for 24 hr. The percentage of infected macrophages (A) and the nnumber of amastigotes per macrophage (B) were determined as described in Materials and Methods. The result represents the mean \pm SD of 3 experiments. The significance of the difference between cells cultures treatments is indicated: **P* ≤ 0.05.

We addressed the question of whether the combination of Boldine-Glucantime[®] treatment reduces the number of infected murine peritoneal macrophages. Cell cultures exposed to 100 μ g/ml of boldine and 300 μ g/ml of Glucantime[®] during 24 hr showed a significant reduction of 50% in parasitized cells compared with cell cultures exposed just to Glucantime[®] (Fig. 1A). These data indicated that boldine treatment increases the resistance of macrophages to *L. amazonensis* infection and corroborates with Glucantime[®] to reduce the number of infected cells.

We also analyzed the parasite per cell, as part of parasite burden investigation. As shown in Fig. 1B, boldine treatment induced a significant reduction in the number of parasites inside the cells (decrease of 50%) at initial experimental concentration (50 μ g/ml). Interestingly, the reduction in parasite per cell observed in macrophages under lower concentration was observed in cells cultured with higher concentration of boldine (600 μ g/ml). However, these data were similar to the Glucantime[®] treated cell (Fig. 1B).

Viability was determined in cells treated with boldine and/ or Glucantime[®]; there was no significant difference in the OD values obtained by the MTT assay in infected peritoneal macrophages submitted to all experimental concentration of boldine. In addition, higher concentration of boldine (600 µg/ ml) showed significantly high OD values by the MTT assay than Glucantime[®] treated cells (Fig. 2). The results also indicated no toxicity of boldine treatment in non-infected murine macrophage.

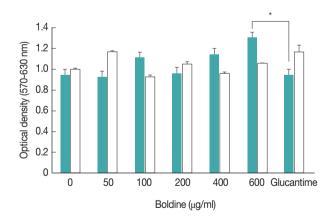


Fig. 2. Effects of boldine on peripheral macrophages. The MTT production by macrophages was determined in cell cultures treated with several concentrations of boldine in infected murine macrophage (\blacksquare) or in non-infected murine macrophage as control (\square). Results represent the mean ± SD of 1 representative experiment out of 3 independently performed experiments with similar results. The significance of the difference between cells cultures treatments is indicated: * $P \le 0.05$.

Boldine, an aporphine alkaloid found in the leaves/bark of boldo (*P. boldus*), a widely distributed tree native to Chile has been shown to have potential novel activities against viruses [22] and cytotoxic and chemotherapeutic properties in breast cancer [23]. Hung et al. [24] studied, for the first time, the leishmanicidal properties of the boldine in the promastigotes of Leishmania mexicana. Mollatagui et al. [25] also reported boldine activity against L. donovani infected macrophages. However, there are no reports available regarding the activity of Boldine against New World *Leishmania* species.

This research was designed to study the in vitro activity of boldine on peritoneal murine macrophages infected with *L. amazonensis* amastigotes. Treatment was initiated 1 hr after infection, an interval of time that allowed for the establishment of the amastigotes phagocytosis, and ensured a homogeneous cell culture infection. Results revealed marked improvement of macrophage capacity in decrease of the parasite burden. Similarly, boldine treatment showed significant reductions of the parasite cell burden and trypanocidal and anti-viral activity using in vitro model [22,26]. The biological relevance of boldine treatment appears to be correlated with its immunological modulatory effect by oxidative stress modulation [27].

Boldine is extensively prescribed as a potent "natural" antioxidant and possesses several health-promoting properties like antiinflammatory, antitumor promoting, antidiabetic, and cytoprotective. It is a relatively safe drug judging from the many animal studies and its widespread use as over-the-counter drug supplement. A relatively high boldine concentration is needed to induce lethality in laboratory animal tests. Kreitmair [28] demonstrated that an oral administration of 500-1,000 mg/kg of boldine was required to cause death in mice and guinea pigs, respectively. In the present study, boldine was evaluated for its cytotoxic effect in macrophage cells using the MTT cell viability assay. The macrophages, whether infected or not with *Leishmania*, and treated with boldine showed similar viability to non-treated cells even when treated with high boldine concentration.

In conclusion, this study proved, for the first time, boldine's in vitro protective activity against *L. amazonensis* macrophage infection, thus, setting the stage for boldine as a suitable lead compound for the synthesis of more effective and less toxic antileishmanial derivatives.

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CONFLICT OF INTEREST

We have no conflict of interest related to this work.

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