



Lignans with NADPH Oxidase 2 (NOX2)-inhibitory Activity from the Fruits of *Schisandra chinensis*

Jung-Min Park[†], Pisey Pel[†], Young-Won Chin*, and Moo-Yeol Lee*

College of Pharmacy, Dongguk University, Goyang, Gyeonggi-do 10326, Republic of Korea

Abstract – An isoform of NADPH oxidase (NOX), NOX2 is a superoxide-generating enzyme involved in diverse pathophysiological events. Although its potential as a therapeutic target has been validated, there is no clinically available inhibitor. Herein, NOX2-inhibitory activity was screened with the constituents isolated from *Schisandra chinensis*, which has been reported to have antioxidant and reactive oxygen species (ROS)-scavenging effects. Among the partitions prepared from crude methanolic extract, a chloroform-soluble partition showed the highest NOX2-inhibitory activity in PLB-985 cell-based NOX2 assay. A total of twenty nine compounds (**1** - **29**) were identified from the chloroform fraction, including two first isolated compounds; dimethyl-malate (**25**) and 2-(2-hydroxyacetyl) furan (**27**) from this plants. Of these constituents, two compounds (gomisin T, and pregomisin) exhibited an NOX2-inhibitory effect with the IC₅₀ of 9.4 ± 3.6, and 62.9 ± 11.3 μM, respectively. They are confirmed not to be nonspecific superoxide scavengers in a counter assay using a xanthine–xanthine oxidase system. These findings suggest the potential application of gomisin T (**6**) and other constituents of *S. chinensis* to inhibit NOX2.

Keywords – NADPH oxidase 2 (NOX2), NOX2 inhibitor, *Schisandra chinensis*, Gomisin T

Introduction

NADPH oxidase (NOX) is a membrane-bound enzyme complex that catalyzes the electron transfer from NADPH to molecular oxygen, as shown in the following reaction. NOX is a genuine reactive oxygen species (ROS) generator in the sense that ROS generation is the primary and sole function of NOX, unlike other sources of ROS, such as xanthine oxidase or cytochrome P450, which generate ROS as a byproduct of their reactions or after conversion to a dysfunctional state.¹



NOX was originally found in phagocytic leukocytes, such as neutrophils and macrophages. It participates in respiratory burst during immune responses by producing

superoxide. NOX in phagocytes is composed of membrane-bound catalytic subunits, such as gp91phox and p22phox, and regulatory cytosolic subunits, including p47phox, p67phox, p40phox, and Rac. NOX is latent under normal circumstances, but upon activation, its regulatory cytosolic subunits translocate to the membrane and associate with membrane-bound catalytic subunits. The assembled enzyme complex then produces superoxide via the one-electron reduction of the oxygen molecule by gp91phox using NADPH as the electron donor.² During the last two decades, NOX activity has been reported in non-phagocytic cells and additional isoforms of gp91phox have been discovered. To date, seven NOX isoforms have been identified in humans and named NOX1, NOX2, NOX3, NOX4, and NOX5 and dual oxidase (DUOX) 1 and DUOX2. The originally identified phagocytic NOX has now been named NOX2. NOX exhibits tissue-specific expression patterns and distinct subcellular localizations that are dependent on isoforms. Furthermore, each isoform works with different regulatory subunits and has a specific regulatory mechanism; thus the isoforms have quite different pathophysiological functions.³

As its etiological roles have been uncovered, NOX has come to be viewed as a therapeutic target. Among NOX isoforms, NOX2 has been proved to be a therapeutic

*Author for correspondence

Young-Won Chin (Ph.D.), College of Pharmacy, Dongguk University-Seoul, 32 Dongguk-ro, Ilsandong-gu, Goyang, Gyeonggi-do 10326, Republic of Korea.

Tel: +82-31-961-5218; E-mail: f2744@dongguk.edu

Moo-Yeol Lee (Ph.D.), College of Pharmacy, Dongguk University-Seoul, 32 Dongguk-ro, Ilsandong-gu, Goyang, Gyeonggi-do 10326, Republic of Korea.

Tel: +82-31-961-5222; E-mail: mlee@dongguk.edu

[†]These authors contributed equally to this work.

target for inflammation, diabetic complications, cardiovascular disorders, and neurodegenerative diseases, such as Alzheimer's and Parkinson's diseases.⁴ Accordingly, considerable effort has been put into developing NOX2 inhibitors. Not a few chemicals have been reported to inhibit NOX2. However, most have limitations in specificity and *in vivo* or clinical applicability, including non-specific ROS-scavenging activity, off-target effects, and poor pharmacokinetic properties.⁵ Consequently, they have yet to reach the clinic and improved NOX2 inhibitors are currently under development.

Schisandra chinensis (Turcz.) Baill. (Schisandraceae) is a deciduous woody vine native to Far East Asia. Its berries have been used in traditional oriental medicine. Indeed, various recent studies have revealed their diverse biological effects on the central nervous, immune, endocrine, and cardiovascular systems.⁶ In particular, the fruits of *S. chinensis* have anti-oxidant and anti-inflammatory activities in *in vitro* and *in vivo* studies. In addition, its active constituents, such as gomisin A and schisandrin B, are capable of inhibiting lipopolysaccharide-induced NOX activation and ROS production.^{7,8} Based on these previous reports, the NOX2-inhibitory effect was explored with the fruits of *S. chinensis*.

Experimental

General experimental procedure – Nuclear magnetic resonance (NMR) spectra were determined on a Varian 400 (Varian, Palo Alto, CA, USA) - 400 MHz spectrometer at 400 MHz for ¹H-NMR and at 100 MHz for ¹³C-NMR. High-resolution mass spectra data were obtained from a Waters Xevo G2 Q-TOF mass spectrometer (Waters, Milford, MA, USA). Semi-preparative high performance liquid chromatography (HPLC) was performed on a Gilson 321 pump and Gilson 172 Diode Array Detector (Gilson, Middleton, WI, USA). The HPLC columns used were YMC-pack Ph, 250 × 20 mm (YMC, Kyoto, Japan) and Luna 5 μm C18 (20) 100A column 250 × 10 nm (Phenomenex, Seoul, Korea). Water was purified using a Milli-Q system (MilliporeSigma, Burlington, MA, USA). Column chromatography was performed on C-18 RP silica gel (Cosmosil, Kyoto, Japan) and Sephadex LH-20 (GE Healthcare, Uppsala, Sweden). TLC analysis was conducted on silica gel 60 F₂₅₄ plates (Merck, Darmstadt, Germany). The spots were visualized by spraying with 10% aqueous H₂SO₄.

Plant materials – the fruits of *S. chinensis* was obtained from an oriental market, and the raw material (CYWDU-KP0009) used in the present study was deposited

at the College of Pharmacy, Dongguk University-Seoul, Republic of Korea.

Extraction and Isolation: the dried fruit (5.8 kg) was kept at room temperature, extracted by MeOH three times, and evaporated *in vacuo*. MeOH extract (1.53 kg) was suspended in H₂O and partitioned with CHCl₃, EtOAc, and *n*-butanol successively to give the residue of 180 g of CHCl₃-soluble extract, 132 g of EtOAc-soluble extract, 434 g of butanol-soluble extract, and 759 g of water-soluble extract. The portion of CHCl₃-soluble extract (35 g, SCC) was chromatographed with silica column chromatography (5 × 90 cm, 600 g) using the gradient of increasing polarity with hexane–EtOAc (30:1–2:1) and CHCl₃–MeOH (20:1–1:1) as solvents. The resultant eluent was fractionated into 17 sub-fractions (SCC-1–SCC-17). The SCC-3 fraction (42.4 mg) was chromatographed with Sephadex LH-20 column chromatography (2 × 90 cm) using the gradient with CHCl₃–MeOH (1:1) as solvents and fractionated into three sub-fractions (SCC-3A and SCC-3C). The SCC-3C fraction was subjected to HPLC separation with MeCN–H₂O (80:20) at 3.0 ml/min by isocratic elution for 20 min and then 100% MeCN for 6 min to afford compound **15** (*t*_R 19.55 min, 17.2 mg). The SCC-5 fraction (162.3 mg) was subjected to HPLC separation with MeCN–H₂O (70:30) at 3.0 ml/min by isocratic elution for 34 min and then 100% MeCN for 6 min to afford compounds **28** (*t*_R 20.25 min, 4.2 mg), **16** (*t*_R 30.25 min, 6.5 mg), and **17** (*t*_R 33.10 min, 21.2 mg). The SCC-7 fraction (112.2 mg) was subjected to HPLC separation with MeCN–H₂O (80:20) at 7.0 ml/min by isocratic elution for 35 min and then 100% MeCN for 6 min to afford compounds **18** (*t*_R 21.0 min, 5.9 mg), **23** (*t*_R 28.25 min, 1.2 mg), and **24** (*t*_R 31.25 min, 27.1 mg). The SCC-8 fraction (71.1 mg) was purified by MPLC equipped with a reversed-phase (RP) column using a gradient mixture of MeOH–H₂O (30:70–90:10), giving eight sub-fractions (SCC-8A–SCC-8H). The SCC-8H fraction (14.8 mg) was subjected to HPLC separation with MeCN–H₂O (80:20) at 2.0 ml/min by isocratic elution for 20 min and then 100% MeCN for 6 min to afford compound **19** (*t*_R 17.75 min, 2.1 mg). The SCC-9 fraction (576.4 mg) was purified by MPLC equipped with an RP column using a gradient mixture of MeOH–H₂O (5:95–90:10), giving 12 sub-fractions (SCC-9A–SCC-9L). Compounds **27** (6.5 mg) and **20** (105.6 mg) were obtained from SCC-9B and SCC-9F, respectively. The SCC-9G fraction (148.4 mg) was chromatographed with silica column chromatography (2 × 35 cm, 75 g) using the gradient with hexane–EtOAc (50:1–10:1) as solvents and fractionated into five sub-fractions (SCC-9G1–SCC-9G5). The

SCC-9G3 fraction (24.3 mg) was chromatographed with silica column chromatography (1 × 10 cm, 7 g) using the gradient with hexane–EtOAc (10:1–2:1) as solvents and fractionated into six sub-fractions (SCC-9G3A–SCC-9G3F). Compound **21** (9.5 mg) was obtained from the precipitate from the SCC-9G3A fraction. SCC-10 (1.12 g) was chromatographed with silica column chromatography (1.5 × 45 cm, 90 g) using the gradient of increasing polarity with hexane–EtOAc (10:1–3:1) and CHCl₃–MeOH (100:0–10:1) as solvents and fractionated into seven sub-fractions (SCC-10A–SCC-10G). The SCC-10B fraction (15.2 mg) was subjected to HPLC separation with MeCN–H₂O (80:20) at 2.0 ml/min by isocratic elution for 11 min and then 100% MeCN for 6 min to afford compound **7** (*t_R* 10.25 min, 2.0 mg). The SCC-10D fraction (305.5 mg) was purified by MPLC equipped with an RP column using a gradient mixture of MeOH–H₂O (40:60–90:10), giving 11 sub-fractions (SCC-10D1–SCC-10D11). Compound **2** (25.0 mg) was obtained from SCC-10D10. The SCC-10D6 fraction (20.1 mg) was subjected to HPLC separation with MeCN–H₂O (70:30) at 7.0 ml/min by isocratic elution for 20 min and then 100% MeCN for 6 min to afford compound **1** (*t_R* 18.25 min, 3.3 mg). The SCC-11 fraction (1096.0 mg) was purified by an MPLC equipped with an RP column using a gradient mixture of MeOH–H₂O (5:90–90:10), giving 20 sub-fractions (SCC-11A–SCC-11T). Compound **25** (5141.1 mg) was obtained from SCC-11A. The SCC-11H to SCC-11K fractions (241.3 mg) were combined and subjected to HPLC separation with MeCN–H₂O (60:40) at 3.0 ml/min by isocratic elution for 30 min and then 100% MeCN for 6 min and afforded compounds **29** (*t_R* 12.75 min, 5.5 mg), **11** (*t_R* 17.25 min, 2.6 mg), **3** (*t_R* 18.75 min, 7.7 mg), and **4** (*t_R* 20.10 min, 9.7 mg). The SCC-12 fraction (2062.5 mg) was purified by an MPLC equipped with an RP column using a gradient mixture of MeOH–H₂O (5:90–90:10), giving 14 sub-fractions (SCC-12A–SCC-12N). The SCC-12J fraction (69.9 mg) was subjected to HPLC separation with MeCN–H₂O (50:50) at 3.0 ml/min by isocratic elution for 34 min and then 100% MeCN for 6 min and afforded compound **5** (*t_R* 34.75 min, 6.4 mg). The SCC-12K fraction (56.5 mg) was purified by MPLC equipped with an RP column using a gradient mixture of MeOH–H₂O (40:60–80:20), giving five sub-fractions (SCC-12K1–SCC-12K5). The SCC-12K4 fraction (36.3 mg) was subjected to HPLC separation with MeCN–H₂O (60:40) at 3.0 ml/min by isocratic elution for 20 min and then 100% MeCN for 6 min and afforded compounds **8** (*t_R* 10.55 min, 1.5 mg), **22** (*t_R* 11.45 min, 2.7 mg), **9** (*t_R* 14.25 min, 1.8 mg), and **10** (*t_R* 16.45 min, 12.5 mg). The SCC-

13 fraction (1495.6 mg) was purified by MPLC equipped with an RP column using a gradient mixture of MeOH–H₂O (5:90–90:10), giving 26 sub-fractions (SCC-13A–SCC-13Z). The SCC-13K to SCC-13M fractions (163.2 mg) were combined and then subjected to HPLC separation with MeCN–H₂O (50:50) at 7.0 ml/min by isocratic elution for 16 min and then 100% MeCN for 6 min and afforded compounds **6** (*t_R* 13.50 min, 15.8 mg), **12** (*t_R* 15.75 min, 5.4 mg), and **13** (*t_R* 17.55 min, 2.8 mg). SCC-13T was chromatographed with Sephadex LH-20 column chromatography (2 × 90 cm) using the gradient with hexane–EtOAc 100% MeOH as solvents and fractionated into two sub-fractions (SCC-13T1 and SCC-13T2). Compound **14** (32.0 mg) was obtained from the SCC-13T1 fraction. The SCC-15 fraction (1441.9 mg) was purified by MPLC equipped with an RP column using a gradient mixture of MeOH–H₂O (10:90–90:10), giving 19 sub-fractions (SCC-15A–SCC-15S). Compound **26** (18.2 mg) was obtained from SCC-15B.

Dimethyl-malate (25): Amorphous oil, ¹H-NMR (CDCl₃, 400 MHz): δ 4.53 (1H, t, *J* = 6.0, 4.4 Hz, H-2), 3.82 (3H, s, 1-OCH₃), 3.73 (3H, s, 4-OCH₃), 3.39 (1H, brs, 2-OH), 2.87 (1H, dd, *J* = 16.5, 6.2 Hz, H-3b), 2.80 (1H, dd, *J* = 16.5, 6.2 Hz, H-3a). ¹³C NMR (CDCl₃, 100 MHz): δ 173.7 (C-1), 171.0 (C-4), 67.2 (C-2), 52.8 (4-OCH₃), 52.0 (1-OCH₃), 38.4 (C-20).

2-(2-hydroxyacetyl)furan (27): amorphous powder, ¹H-NMR (CDCl₃, 400 MHz): δ 7.64 (1H, d, *J* = 1.7 Hz, H-5), 7.32 (1H, d, *J* = 3.7 Hz, H-3), 6.60 (1H, dd, *J* = 3.7, 1.7 Hz, H-4), 4.75 (2H, s, H-7). ¹³C NMR (CDCl₃, 100 MHz): δ 187.6 (C-6), 150.1 (C-2), 147.0 (C-5), 117.8 (C-3), 112.6 (C-4), 65.0 (C-7).

NOX2 activity assay – NOX2 activity was assessed in PLB-985 cells using an 10-acetyl-3,7-dihydroxyphenoxazine (Amplex Red; Molecular Probes, Eugene, OR, USA) fluorescence ROS indicator. PLB-985 cells were kindly provided by Dr. M. Dinauer (Washington University, St. Louis, MO, USA). Cells were cultured in Roswell Park Memorial Institute (RPMI) 1640 medium containing 10% fetal bovine serum, 100 U/ml penicillin and 100 μg/ml streptomycin, and were maintained in a humidified chamber at 37 °C with a 5% CO₂ environment. For assay, cells were washed and resuspended with Hanks' balanced salt solution (HBSS; 130 mM NaCl, 5 mM KCl, 1.5 mM CaCl₂, 1 mM MgCl₂; pH 7.4) and 1 × 10⁵ cells were aliquoted into a well of 96-well plate. Cells were pretreated with testing material for 60 min and NOX2 was stimulated with 200 ng/ml of phorbol 12-myristate 13-acetate (PMA; Santa Cruz Biotechnology, Dallas, TX, USA). ROS generation was detected with 10 μM of

Amplex Red combined with 0.1 U/ml of horseradish peroxidase (HRP; Sigma-Aldrich, St. Louis, MO, USA). The fluorescence intensity was measured at 560/585-nm excitation/emission wavelengths for 60 min with 5-min interval using the SpectraMax M3 microplate reader (Molecular Devices, Sunnyvale, CA, USA). Increase in fluorescence intensity was confirmed to be linear for more than 60 min.

Superoxide scavenging assay – Superoxide scavenging activity was tested in cell-free, xanthine/xanthine oxidase superoxide generating system. Superoxide was produced by 200 μ M of xanthine and 0.01 U/ml of xanthine oxidase in HBSS with or without testing compound. Superoxide was quantified with 500 μ M 2-(4-iodophenyl)-3-(4-nitrophenyl)-5-(2,4-disulfophenyl)-2H-tetrazolium (WST-1; Seoul Clinical Genomics, Seoul, Korea) by measuring the absorbance at 450 nm for 5 min using the SpectraMax M3 microplate reader (Molecular Devices).

HRP activity assay – Peroxidase activity was determined by a colorimetric method. The reaction mixture was composed of 0.003% H_2O_2 , 0.5% *o*-dianisidine, and testing compound in 10 mM sodium phosphate buffer (pH 6.0). The reaction was initiated by the addition of 10 μ l of 0.1 U/ml HRP to 90 μ l of reaction mixture. The absorbance at 460 nm was measured at 15-s intervals for 5 min with the SpectraMax M3 microplate reader (Molecular Devices). Boiled HRP was prepared by heat-inactivation at 99 $^{\circ}$ C for 15 min and used as a negative control. Relative HRP activity was calculated from the slope of the graph of increasing absorbance.

Statistical analyses – Means and standard error of means were calculated for all experimental groups. Data were subjected to one-way analysis of variance followed by Dunn's test to determinewhether the differences relative to the controls were significant. Statistical analysis was performed with SigmaPlot software ver. 13 (Systat Software, San Jose, CA, USA). *P* values of < 0.05 were considered statistically significant.

Result and Discussion

The NOX2-inhibitory effect was tested with crude methanolic extract and its organic solvent partitions, including chloroform-soluble, ethyl acetate-soluble, butanol-soluble, and water-soluble extracts. PLB-985 is a human diploid myeloid leukemia cell line that expresses NOX2 exclusively,^{9,10} which was also confirmed in this study by a real-time polymerase chain reaction (data not shown). PLB-985 was pretreated with 100 μ g/ml of each extract for 60 min, and NOX2 was stimulated with phorbol 12-

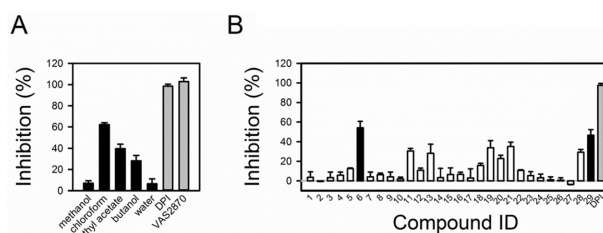


Fig. 1. Inhibitory effects of *S. chinensis* extract and its constituents on NOX2. PLB-985 cells were pretreated with (A) methanolic extract of *S. chinensis* and its partitions or (B) 29 constituents isolated from chloroform partition for 60 min. NOX2 was activated with PMA, and the resultant ROS was measured with Amplex Red for 60 min. Acknowledged NOX inhibitors DPI and VAS2870 were used as a positive control. All values are mean \pm standard error ($n = 5$ for A and 3 for B).

myristate 13-acetate (PMA). Among five extracts, the chloroform-soluble partition exhibited the highest inhibitory potential, which was followed by the ethyl acetate partition and butanolic partition, whereas the aqueous partition and methanolic extract showed minimal activity (Fig. 1A). The acknowledged NOX inhibitors diphenyleneiodonium (DPI) or VAS2870 were confirmed to work in this assay system at 10 mM (Fig. 1A, gray bars).

To determine the active constituents in the chloroform-soluble partition, repeated chromatographic separations were conducted. The twenty nine compounds were isolated and identified (Fig. 2). All structures of **1**–**29** were identified by the comparison of their spectroscopic data with those of the literature as the following: gomisin G (**1**),¹¹ angeloylgomisin Q (**2**),¹² tigloylgomisin P (**3**),¹³ schisantherin A (**4**),¹⁴ benzoylgomisin Q (**5**),¹⁵ (+)-gomisin T (**6**),¹⁶ schisandrol B (**7**),¹⁷ schisandrol A (**8**),^{16,18} tigloylgomisin H (**9**),¹⁹ angeloylgomisin H (**10**),¹⁹ schinlignan D (**11**),²⁰ (+)-gomisin H (**12**),²¹ schinlignan F (**13**),²² benzoylgomisin H (**14**),¹⁹ schisandrin C (**15**),^{22,23} g-schizandrin (**16**),²⁴ schisandrin B (**17**),²³ gomisin L₂ (**18**),²⁵ (-)-gomisin L₁ (**19**),²³ gomisin J (**20**),²³ (-)-gomisin K₁ (**21**),²⁶ gomisin D (**22**),¹² dehydroschisandro A (**23**),²⁷ schisandrin A (**24**),²³ dimethyl-malate (**25**),²⁸ 2-hydroxy-5-methyl ester (**26**),²⁹ 2-(2-hydroxyacetyl)furan (**27**),³⁰ anwulignan (**28**),³¹ and pregomisin (**29**)³² (Fig. 2).

These twenty nine compounds were screened for their NOX2-inhibitory activity. PLB-985 cells were pretreated with 10 μ M of testing compounds, and PMA-stimulated ROS generation was detected with Amplex Red. Of these, lignan-type structured compounds **6**, and **29** were identified as active inhibitors, which exhibited more than 40% inhibition (Fig. 1B). To calculate the half-maximal inhibitory concentration (IC₅₀), the concentration dependency for NOX2 inhibition was tested and plotted (Fig. 3). The

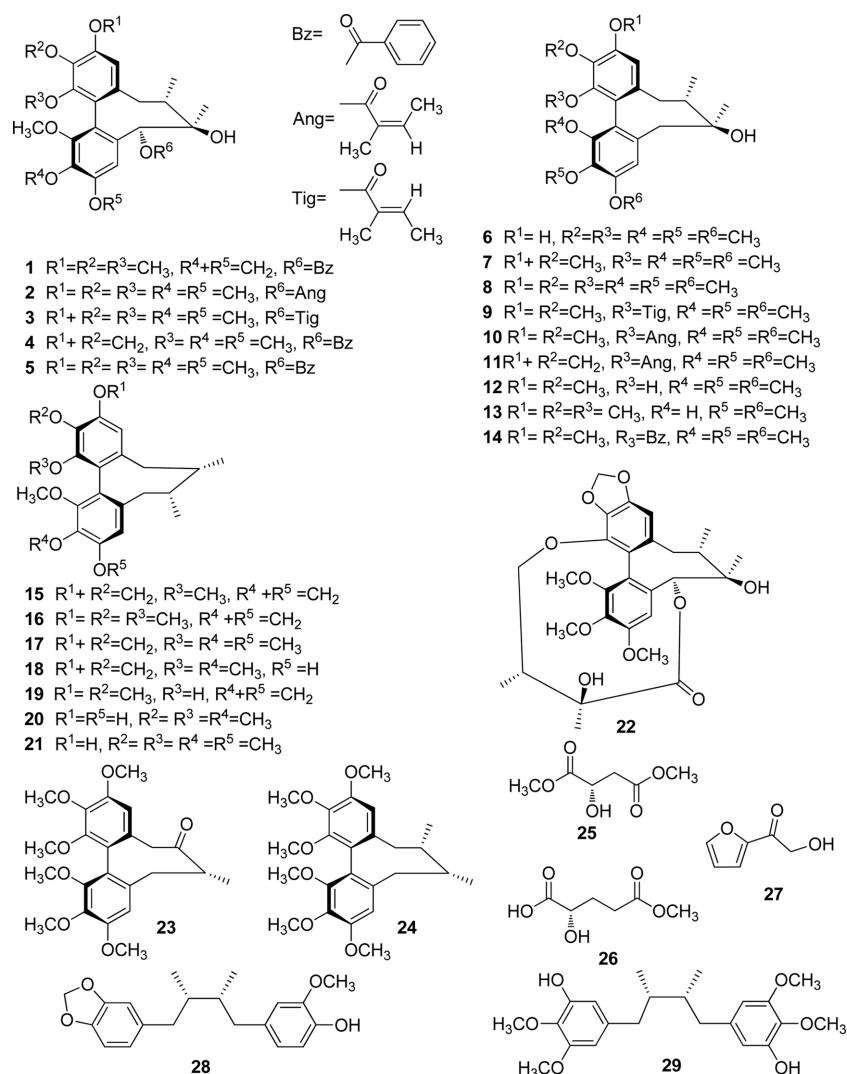


Fig. 2. Structures of compounds isolated from the chloroform-soluble extract of *S. chinensis*.

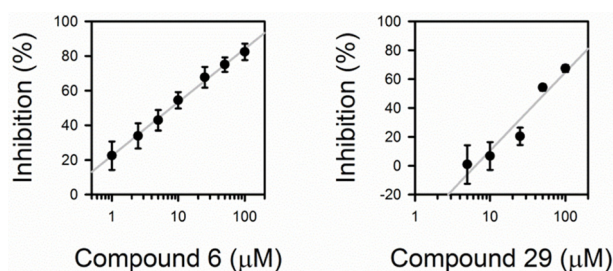


Fig. 3. Concentration-dependent NOX2-inhibitory activity of compounds **6**, and **29**. PLB-985 cells were pretreated with testing compounds, and PMA-stimulated NOX2 activity was assessed with Amplex Red. Values are mean \pm standard error ($n = 3 - 4$).

IC_{50} for compound **6** was approximately $9.4 \pm 3.6 \mu M$, and that for compound **29** was approximately $62.9 \pm 11.3 \mu M$, respectively.

In addition to NOX2 inhibition, the nonspecific ROS-

scavenging effect may lead to a false positive result in NOX2 activity assay. Hence, ROS-scavenging activity was tested in a xanthine-xanthine oxidase superoxide-generating system. Superoxide produced by xanthine and xanthine oxidase was quantified with WST-1 in the presence or absence of compound **6** or **29**. However, neither compound **6** nor **29** showed superoxide-scavenging activity at $50 \mu M$, while the superoxide dismutase mimetic TEMPOL abolished superoxide nearly completely (Fig. 4). Therefore, compounds **6** and **29** are not nonspecific superoxide scavengers, but NOX2 inhibitors.

Since Amplex Red requires peroxidase to react ROS and to produce the red-fluorescent oxidation product, HRP was used in this assay. Therefore, the effect of compounds **6** and **29** on HRP activity was examined to test whether NOX2 inhibition observed in Fig. 1 and Fig.

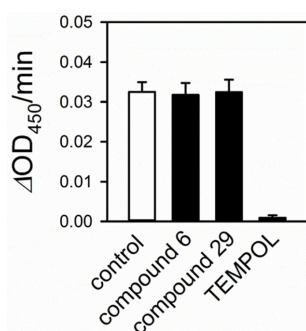


Fig. 4. Assessment of superoxide-scavenging activity of compounds **6** and **29**. Superoxide was produced by xanthine and xanthine oxidase in the presence or absence of testing compound and was quantified with WST-1. Values are mean \pm standard error (n = 8).

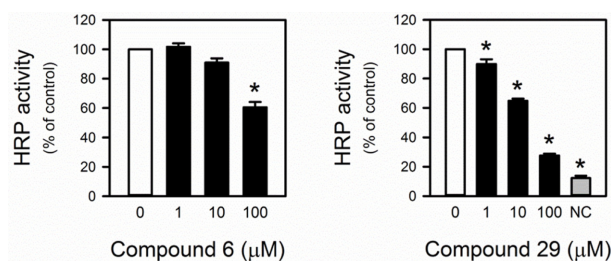


Fig. 5. Effect of compounds **6** and **29** on HRP activity. HRP was treated with the indicated concentrations of compound **6** or **29**, and its activity was determined by a colorimetric method using a peroxidase substrate *o*-dianisidine. The activity of heat-inactivated HRP was measured as a negative control. Values are mean \pm standard error (n = 3).

3 is ascribed to artifact caused by HRP inhibition rather than NOX2 inhibition. Compound **6** inhibited HRP marginally; its inhibitory activity was minimal at 10 μ M, although the extent of inhibition was approximately 39% at 100 μ M (Fig. 5, left panel). Considering less than 10 μ M of IC₅₀, the attenuation of ROS generation shown in Fig. 1 and Fig. 3 should be mainly attributed to NOX2 inhibition. However, compound **29** was capable of inhibiting HRP substantially (Fig. 5, right panel). Although it is unlikely that HRP inhibition is directly proportional to NOX2 inhibition, HRP inhibitory activity may be involved in NOX2 inhibition observed in Fig. 1 and Fig. 3. According to these results, the efficacy on NOX2 might be lower than those calculated based on the data from Fig. 3, especially for compound **29**. Further study will be needed to characterize the non-specific effect of compounds **6** and **29**.

In conclusion, potential inhibitors for NOX2 were found from the fruit extract of *S. chinensis*. One of them, gomisin T (**6**), exhibited the highest efficacy with an IC₅₀ of less than 10 μ M. NOX2 inhibitory activity was also

observed with pregomisin. Although several lines of chemical structure have been reported to inhibit NOX,⁴ lignan-type structures (**6**, **29**) are proposed as NOX2 inhibitors for the first time. Gomisin T (**6**) and other constituents of *S. chinensis* were identified as promising NOX2 inhibitors. Further characteristics regarding structure–activity relationships, isoform specificity, the molecular mechanism of action, pharmacokinetic properties, and *in vivo* efficacy will be investigated in future studies.

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