Disinfection by Ozone Microbubbles Can Cause Morphological Change of Fusarium oxysporum f. sp. melonis Spores

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To investigate the difference in the disinfectant efficiency of ozone microbubbles (O3MB) and ozone millibubbles (O3MMB), the morphological change of the treated Fusarium oxysporum f. sp. melonis spores was observed with scanning and transmission electron microscopies (SEM and TEM). The disinfectant efficiency of O3MB on F. oxysporum f. sp. melonis spores was greater than that of O3MMB. On observation with SEM, it was revealed that morphological change of F. oxysporum f. sp. melonis spores was caused by O3MB and O3MMB, and damage to the spore surfaces by O3MB occurred sooner than that by O3MMB. On observation with TEM, it was furthermore confirmed that F. oxysporum f. sp. melonis spores treated with O3MB induced wavy deformation of cell membrane and the intracellular change different from that with O3MMB. Therefore, the greater disinfection efficiency of O3MB was suggested to be caused due to the function of the MB in addition to the oxidative power of O3.

Keywords: disinfection, Fusarium oxysporum f. sp. melonis spores, ozone microbubbles

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Hydroponic culture is a plant cultivation technique in which plants are grown in a nutrient solution without soil. It has the potential for high crop productivity in a small area. However, if plant pathogens enter the solution, they can spread rapidly throughout the hydroponic culture facility and cause catastrophic damage. For this reason, disinfection of the solution is essential, although direct administration of pesticides into the solution has been prohibited at law (Ministry of Agriculture, Forestry and Fisheries, 2010). Therefore, it is desired to establish safe and effective alternative disinfection methods, and the various disinfection treatments have been investigated, including UV light, heat, the titanium dioxide photocatalytic reaction, and ozone (O3) (Bando et al., 2008; Dannehl et al., 2016; Ehret et al., 2001; Igura et al., 2004; Koohakan et al., 2003; Ohtani et al., 2000; Runia, 1995). However, these treatments still are not put into any practical use in terms of the efficiency, treatment time, and running cost. Particularly, O3 gas is effective disinfectant due to its strong oxidation power, although it is difficult to use in hydroponic cultures because of its extremely low solubility in water (0.105 g 100 ml−1 (0°C)).

Recently, tiny bubbles less than 50 µm in diameter, called microbubble (MB), have been studied and used in many fields. They rise more slowly in water than millibubbles (MB), which have diameters in the mm to cm range (Takahashi et al., 2003; Takahashi, 2005), and possess additional properties such as the interface charge, long stagnation, slow buoyancy, the shrinkage and the generation of free radicals by their collapsing other than dissolving power (Li and Tsuge, 2006; Li et al., 2009; Zheng et al., 2015). Previously, we focused on long retention time in water and the dissolving power of MB in addition to the strong oxidative power of O3, and found that O3MB were more effective than O3MMB for the disinfection of Fusarium oxysporum f. sp. melonis spore and Pectobacterium
carotovorum subsp. carotovorum in nutrient solution for hydroponic culture (Kobayashi et al., 2011a, 2011b, 2011c, 2012). Furthermore, effective disinfection by O₃MB has been reported by other researchers (Chuajedton et al., 2015; Inatsu et al., 2011), although it is still not clear about the exact mechanism of disinfection by O₃MB. In this study, the morphological change of *F. oxysporum* f. sp. melonis spores by O₃MB and O₃MMB was therefore observed by using scanning and transmission electron microscopies (SEM and TEM), and the difference was discussed.

*F. oxysporum* f. sp. *melonis* NBRC6385 suspension (approximately 1.0 × 10⁷ spores ml⁻¹) was prepared in a manner similar to a previous report (Kobayashi et al., 2011a). For each experiment, 15 l of tap water were collected in a plastic cylindrical container (28 cm dia. × 48 cm height) and kept for 24 h at room temperature to remove chlorine. Water quality test paper (Nissan aquacheck 3; Nissan Chemical Industries, Ltd., Tokyo, Japan) was used to confirm that no residual chlorine remained. O₃ was generated by using an O₃ generator (ED-OG-A10, Ecode- sign Co. Ltd., Saitama, Japan) at a flow rate of 2.5 l min⁻¹. O₃MB and O₃MMB were generated by using a decompression type MB generator (20NEDO4S, Shigen-Kaihatsu Co., Ltd., Kanagawa, Japan) and a commercial air pump, respectively. The concentration of dissolved O₃ in both O₃MB and O₃MMB waters was set to 1.5 ppm at 15°C. The pH of O₃MB and O₃MMB waters was the same at 6.8 and was not changed before and after O₃MB and O₃MMB generation. *F. oxysporum* f. sp. *melonis* spores were added to 100 ml of the O₃MB and O₃MMB waters with final concentrations of 1.0 × 10²-1.0 × 10⁴ cfu ml⁻¹. Aliquots of the treated waters were collected after 0, 15, 30, 45, 60 and 120 s. Aliquots of 0.1 ml of the collected waters were plated on potato dextrose agar (Difco, Becton Dickinson, Flanklin Lakes, NJ, USA) plates, and the plates were incubated at 30°C for 48 h. After incubation, the numbers of surviving spores were measured by counting the colonies formed on the plates. The detection limit was 10 cfu ml⁻¹. Each experiment was performed in duplicate.

*F. oxysporum* f. sp. *melonis* spores were collected from 5 ml of the suspension by filtration with a cartridge filter (Anotop 10, GE Healthcare UK Ltd., Buckinghamshire, UK). The sample on the cartridge filter was pre-fixed with 2.5% glutaraldehyde solution diluted with a phosphate buffer solution (PBS, pH 7.0). A filter ejected by decomposing the cartridge filter was washed with PBS (pH 7.0), post-fixed with 2% OsO₄ solution for 1 h, and then serially dehydrated for 20 min each in 50%, 70%, 80%, 90%, 95%, 99.5% and dehydrated ethanol. SEM observations were performed as follows: The dehydrated sample was immersed in a mixture of t-butyl alcohol and dehydrated ethanol (1:1) for about 10 min, transferred to 100% t-butyl alcohol, freeze-dried with a freeze drier (ES-2030, Hitachi High Technologies Co., Tokyo, Japan), and OsO₄-coated with a OsO₄ coater (HPC-1SW, Vacuum device Inc., Mito, Japan) (the thickness of the coating was adjusted to 3 nm). Then the sample was observed with an SEM (JSM-6700F, JEOL Ltd., Akishima, Japan) operated at 3.0 kV. Nine *F. oxysporum* f. sp. *melonis* spores in the SEM photographs were selected at random and the widths was measured with a scale. Significant differences were evaluated by the ANOVA and Fisher’s LSD using the Ekuseru-Toukei 2012 for Window statistical software (Social Survey Research Information Co., Ltd., Tokyo, Japan) (*P < 0.05*). TEM observations were performed as follows: The dehydrated samples were serially immersed for 2 h each in 1:1, 2:1, and 3:1 mixtures of Quetol-651 (Cosmo Bio Co., Ltd., Tokyo, Japan) and ethylene glycol diglycidyl ether, and then embedded in 100% Quetol-651 at 60°C. Ultra-thin sections (thickness 70-100 nm) were made from the embedded samples with an ultramicrotome (ULTRA CUT UCT, Leica Microsystems, Wetzlar, Germany). The ultra-thin sections were doubly electron-strained using 4% uranyl acetate for 12 min and lead nitrate for 5 min, and then observed with a TEM (JEM-2010, JEOL Ltd.) operated at 140 kV.

The survival rates of *F. oxysporum* f. sp. *melonis* spores in water treated with O₃MB and O₃MMB are shown in Fig. 1. The disinfectant efficiency of O₃MB on *F. oxysporum* f. sp. *melonis* spores was greater than that of O₃MMB, because the numbers of surviving spores after treatment with O₃MB and O₃MMB reached the detection limit at 45 s and 60 s, respectively. The result agreed with our previous study (Kobayashi et al., 2011a). Amount of hydroxyl

![Fig. 1. Disinfection of *F. oxysporum* f. sp. *melonis* spores by O₃MB and O₃MMB. The data presented was the mean of duplicate.](image-url)
The Change of Fungal Spores by O₃MB

Radicals generated from O₃ is not enough to have a disinfectant effect (Cho et al., 2003). However, O₃MB may generate more hydroxyl radicals than O₃MMB, because the oxidation-reduction potential and iodine liberation is higher with O₃MB than with O₃MMB (Chuajedton et al., 2015). Furthermore, the use of MB enhances the formation of hydroxyl radicals due to more rapid O₃ decomposition (Tsuge et al., 2009), and the hydroxyl radicals generated from O₃MB accelerate the oxidative power (Chu et al., 2008). The high oxidative power of O₃MB contributes to the oxidative power of O₃, the reactivity of the hydroxyl radicals, the substantivity, ζ surface potential, mass-transfer coefficient, and use efficiency (Zheng et al., 2015). Therefore, the greater disinfectant efficiency of O₃MB than O₃MMB is likely be due to these synergistic effects.

The SEM images of *F. oxysporum* f. sp. *melonis* spores treated with O₃MMB and O₃MB are shown in Fig. 2. The spores treated with O₃MMB for 30 s showed no obvious surface injury, although spores treated with O₃MMB for 180 s were deformed. On the other hand, spores treated with O₃MB showed obvious surface injury after 30 s and the spores were completely destroyed after 180 s. The widths of *F. oxysporum* f. sp. *melonis* spores treated with O₃MMB for 30 s were lower than those of non-treated spores, and then the spores swelled after 180 s (Fig. 3). However, the widths of spores treated with O₃MB for 30

**Fig. 2.** SEM images of *F. oxysporum* f. sp. *melonis* spores. (A) After treatment with O₃MMB for 30 s, (B) After treatment with O₃MMB for 60 s, (C) After treatment with O₃MB for 30 s, (D) After treatment with O₃MB for 60 s, (E) Non-treated.
s were the same as those of non-treated spores and diminished in size by 180 s. Furthermore, the TEM images indicated the appearance of liquid foam in the mid-regions of the *F. oxysporum* f. sp. *melonis* spores treated with O₃MB for 180 s (Fig. 4). Then, the spores had swelled by O₃MB for 180 s, as water entered the cells through the damaged cell wall. Cho et al. (2010) reported that disinfection of bacterial cells by O₃ was due to injury of the cell wall. Zhang et al. (2011) showed that disinfection of *Pseudomonas aeruginosa* by O₃ was due to increase in cell membrane permeability and coagulation of the intracellular substrate. Thanomsub et al. (2002) concluded that disinfection by O₃ was caused due to the destruction of the cell wall and leakage of the intracellular substrate, followed by

![Graphical representation of the widths of *F. oxysporum* f. sp. *melonis* spores in SEM images.](image)

**Fig. 3.** The widths of *F. oxysporum* f. sp. *melonis* spores in SEM images. NT: Non-treated. The results indicate the means with standard deviation of 9 spores.

![TEM images of *F. oxysporum* f. sp. *melonis* spores.](image)

**Fig. 4.** TEM images of *F. oxysporum* f. sp. *melonis* spores. Top: Non-treated, Middle: After treatment with O₃MB for 180 s, Bottom: After treatment with O₃MB for 180 s.
cell lysis. Therefore, it is possible that the *F. oxysporum* f. sp. *melonis* spores treated with O₃MB for 30 s initially shrank due to leakage of the intracellular substrate through the damaged cell wall. On the other hand, the TEM images of the *F. oxysporum* f. sp. *melonis* spores treated with O₃MB for 180 s showed the wavelike deformation of cell membrane and appeared to have a space between the cell membrane and/or wall and the cytoplasm. Diao et al. (2004) confirmed that hydroxyl radicals generated by the Fenton reaction induced the injury of *E. coli* cell membranes. Cho, M., Kim, J., Kim, J. Y., Yoon, J. and Kim, J. H. 2010. Electron microscopic investigation of the bactericidal action of electrochemical disinfection in comparison with chlorination, ozonation and Fenton reaction. Process Biochem. 39:1421-1426.


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