

Effect of *Bacillus aryabhatai* H26-2 and *B. siamensis* H30-3 on Growth Promotion and Alleviation of Heat and Drought Stresses in Chinese Cabbage

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Plants are exposed to biotic stresses caused by pathogen attack and complex abiotic stresses including heat and drought by dynamic climate changes. To alleviate these stresses, we investigated two bacterial strains, H26-2 and H30-3 in two cultivars ('Ryeokkwang' and 'Boram-3-ho') of Chinese cabbage in plastic pots in a greenhouse. We evaluated effects of bacterial strains on plant growth-promotion and mitigation of heat and drought stresses; the role of exopolysaccharides as one of bacterial determinants on alleviating stresses; biocontrol activity against soft rot caused by *Pectobacterium carotovorum* subsp. *carotovorum* PCC21. Strains H26-2 and H30-3 significantly increased fresh weights compared to a MgSO₄ solution; reduced leaf wilting and promoted recovery after re-watering under heat and drought stresses. Chinese cabbages treated with H26-2 and H30-3 increased leaf abscisic acid (ABA) content and reduced stomatal opening after stresses treatments, in addition, these strains stably colonized and maintained their populations in rhizosphere during heat and drought stresses. As well as tested bacte-

rial cells, exopolysaccharides (EPS) of H30-3 could be one of bacterial determinants for alleviation of tested stresses in Chinese cabbages, however, the effects were different to cultivars of Chinese cabbages. In addition to bacterial activity to abiotic stresses, H30-3 could suppress incidence (%) of soft rot in 'Boram-3-ho'. The tested strains were identified as *Bacillus aryabhatai* H26-2 and *B. siamensis* H30-3 based on 16S rRNA gene sequence analysis. Taken together, H26-2 and H30-3 could be candidates for both plant growth promotion and mitigation of heat and drought stresses in Chinese cabbage.

Keywords : abiotic stress, exopolysaccharide, soft rot

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Plants frequently face adverse climates and environmental factors including extreme temperatures (heat, cold, and freezing), drought and salinity; these are the major abiotic stresses that limit plant growth, development, and crop yield. In particular, heat and water deficit are the main environmental stress factors and these two different stresses can occur simultaneously in fields (Dobra et al., 2010). When plants are exposed to stressful conditions, they often respond to the stress by producing ethylene. Stress-induced ethylene can inhibit plant growth and root elongation as well as trigger plant senescence and chlorosis, ultimately leading to death (Barnawal et al., 2014). Agrochemicals, such as aminoethoxyvinylglycine (AVG), Co²⁺, and Ag⁺, are used as inhibitors of ethylene formation or action, increasing plant growth under abiotic stress (Belimov et al.,

2009). However, these ethylene inhibitors are high-priced, are toxic to humans, and impair soil health. Therefore, alternative approaches, including plant-rhizobacteria interactions to alleviate abiotic stresses in plants, have been studied (Belimov et al., 2009; Dimkpa et al., 2009).

Plant-associated bacteria have been used in sustainable agricultural system, especially, plant growth-promoting rhizobacteria (PGPR) have an effect on plant growth and development by production of (i) phytohormones such as indole-3-acetic acid (IAA), abscisic acid (ABA), cytokinins, and gibberellic acid; (ii) volatile compounds; and (iii) bacterial exopolysaccharides (EPS) (Sang et al., 2018; Timusk et al., 2014; Vurukonda et al., 2016); many PGPR are well known to suppress plant disease as biotic stresses by induced resistance (Yang et al., 2009). The beneficial effect of plant-associated bacteria including PGPR is not limited to induced tolerance to biotic stresses but extends to abiotic stresses (Kaushal and Wani, 2016; Yang et al., 2009). However, the agricultural uses of plant-associated bacteria against abiotic stresses remain largely uncommon compared to PGPR and biocontrol against biotic stresses.

Chinese cabbage (*Brassica campestris* spp. *pekinensis*) is one of the most important vegetables in Asia, especially, Korea, China, and Japan. In Korea, commercial Chinese cabbage cultivars were separated by sowing seasons, such as 'Ryeokkwang' as a spring cultivar and 'Boram-3-ho' as a fall cultivar; largely mass consumption of Chinese cabbage occurred in fall-early winter season. Recently, heat and drought conditions by dynamic global climate changes occur at the same times in Korea, it frequently causes serious problems in Chinese cabbage fields during the growing season, causing the productivity of Chinese cabbage to fluctuate and be unstable (Kim et al., 2010).

Here, we investigated effect of bacterial strains on plant growth promotion and mitigation of concurrent stresses including heat and drought in Chinese cabbages; we demonstrated physiological changes in leaves of Chinese cabbages, and bacterial colonization of the rhizosphere during heat and drought stresses. In addition, we determined whether bacterial EPS can promote plant growth and alleviate heat and drought stresses; finally, we evaluated biocontrol activity against bacterial soft rot in two cultivars of Chinese cabbages; identified strains H26-2 and H30-3 based on 16S rRNA sequence analysis.

Bacterial strains were isolated from the rhizospheres of cucumber and Chinese cabbage in Korea. The sampled soils were suspended in 0.85% NaCl solution for 30 min at 28°C and incubated on Reasoner's 2A (R2A) agar (Lab M Ltd., UK) amended with 50 µg/ml of cycloheximide for 3 d at 28°C. A total 46 strains were collected, two strains

(H26-2 and H30-3) among these were selected based on alleviating effect on heat and drought stresses in our preliminary test (Supplementary Fig. 1). Bacterial strains were stored in vials containing tryptic soy broth (TSB, Difco, USA) amended with 20% glycerol and stored at -80°C before use. For the preparation of bacterial suspensions, two strains were grown on tryptic soy agar (TSA, Difco, USA) at 28°C for 48 h and then single colonies were cultured in 250-ml Erlenmeyer flasks containing 100 ml of TSB in a rotary shaking incubator (150 rpm) at 28°C for 48 h. Bacterial cells were harvested by centrifugation at 5,000 rpm for 10 min and suspended in 10 mM MgSO₄ solution. The bacterial suspensions were adjusted to 10⁷ cells/ml with a UV/VIS spectrophotometer (Infinite M200 PRO, TECAN, Austria) and used for further tests.

For *in vitro* assay for bacterial plant growth-promoting (PGP) traits, bacterial ACC deaminase activity was evaluated by the method of Penrose and Glick (2003) and Khandelwal and Sindhu (2013); phosphate solubilization by bacterial strains was measured onto PVK agar amended with bromophenol blue (Pikovskaya, 1948); bacterial IAA production was assayed according to methods of Bric et al. (1991) and Kumar et al. (2012). Bacterial EPS production was determined on a TSA plate by observation of a slimy colony morphology (Tallgren et al., 1999). In addition, siderophore production was detected on chrome azurol S (CAS) agar plate assay (Milagres et al., 1999); hydrogen cyanide (HCN) production was measured on TSA (Castric and Castric, 1983). These bacterial biological traits were measured as positive or negative reactions.

For assessment of growth promotion activity, two cultivars of Chinese cabbage ('Ryeokkwang', NongWoo Bio Ltd, Suwon, Korea and 'Boram-3-ho', Farm Hannong Co., Ltd, Gyeonggi-do, Korea) were used. Chinese cabbage seeds were sown into plastic pots (diameter 9 cm) containing commercial potting mixture (Bunong, Korea) and grown in a greenhouse. Seedlings (four-leaf stage) were drenched with bacterial suspensions of H26-2 and H30-3 (10⁷ cells/ml, 1 ml/g of potting mixture) or a 10 mM MgSO₄ solution as untreated control in a greenhouse. Two weeks after the drench treatment, the fresh weights of the Chinese cabbages (six~seven-leaf stages) were measured. For test of bacterial-mediated alleviation of abiotic stresses (heat and drought), bacterial suspensions and plant materials were prepared according to the methods described above. One week after bacterial drench treatment, plants were uniformly watered with 20 ml tap water and then heat and drought stressed by withholding watering for 5-7 d at 35°C in a growth chamber (16 h fluorescent light/d, 40% relative humidity). Pot positions were randomly changed

every day. When the control had wilted 60-80%, the wilting of plants was assessed using a leaf wilting score (LWS) based on relative water contents (RWC, %) (Barrs and Weatherley, 1962): score 0, 100%; 1, 80-100%; 2, 60-80%; 3, 40-60%; 4, 20-40%; and 5, 0-20% RWC. The stress treatment was terminated by drenching with 40 ml of sterile water (re-watering). Twenty-four h later, fresh weight (g) were evaluated. This experiment was conducted twice with ten replicates each.

For quantification of ABA in Chinese cabbage leaves, plant materials and bacterial suspension were prepared; heat and drought stresses were performed as described above. Leaves were sampled 24 h after stresses treatment, and then, ABA content was measured using the Phytodetek competitive ELISA kit (Agdia, Elkhart, IN, USA). At the same time, stomatal opening was observed using light microscopy (Zeiss Model, Germany) (Jung et al., 2000). At least 100 stomata per plant were counted. This experiment was conducted twice with four (for ABA contents), and eight (for stomatal opening) replicates each.

Bacterial colonization in rhizosphere soils of Chinese cabbage under heat and drought conditions was evaluated by using spontaneous rifampicin-resistant mutants of H26-2 (H26-2 Rif) and H30-3 (H30-3 Rif) (10^7 cells/ml, 1 ml/g) (Sang and Kim, 2012). Bacterial suspensions of H26-2 Rif and H30-3 Rif were drenched into Chinese cabbages (four-leaf stage), one week later, pots were transferred to a growth chamber for heat and drought stresses induced by withholding watering for 5 d at 35°C. The rhizosphere soil was collected 1, 3, and 5 d after stresses (DAS) treatment. Rhizosphere soil (1 g) was put into a 15-ml tube with 10 mM MgSO₄ solution and then incubated at 120 rpm at 28°C for 1 h. Soil suspensions were serially diluted on rifampicin (100 µg/ml)-supplemented TSA (RTSA), incubated at 28°C for 48 h, and colony forming units (cfu) was counted. At the same time, bacterial colonization under non-stress conditions was tested. This experiment was conducted twice with four replicates each.

To assess whether EPS of H26-2 and H30-3 were one of bacterial determinant for growth promotion and mitigation heat and drought stresses in Chinese cabbages, EPS were extracted from the strains (Santaella et al., 2008). For in plate assay, seeds of two Chinese cabbage cultivars were surface-sterilized with 2% NaOCl for 2 min and rinsed with sterile distilled water three times. Sterile seeds were transferred to plates (height 4 cm) containing MS media amended with 400 mM mannitol, as an artificial drought treatment, and EPS (10 µg/ml) of H26-2 and H30-3. They were then kept at 25°C and 16 h fluorescent light per d. The number of cotyledon with open and green leaves after

seed germination and number of lateral root was recorded at 3-4 and 17 d, respectively. This experiment was conducted twice with ten replicates each. *In planta*, seedlings (four-leaf stage) were drenched with an EPS solution (10 µg/ml) of H26-2 and H30-3 or with distilled water as the control. One week later, pots were transferred to heat and drought stresses conditions in a growth chamber as described above. Then, leaf wilting scores and fresh weights were measured; at the same time, a non-stress condition was tested. This experiment was conducted twice with eight replicates each.

For test of biocontrol activity of two bacterial strains against soft rot in Chinese cabbages, preparation of plants and bacterial suspensions were conducted as described above. Benzothiadiazole (BTH, 0.1 mM) was used as a chemical control. For plant inoculation, *P. carotovorum* subsp. *carotovorum* PCC21 was cultured in 50 ml of Luria-Bertani (LB) broth at 28°C, 160 rpm for 24 h, and then diluted to OD₆₀₀ = 0.1. One week after bacterial treatment, two cultivars of Chinese cabbages were drenched by a 4:1 mixture of PCC21 and sterile mineral oil (heavy white oil, sigma) (Lee and Cha, 2001), and disease incidence (%) was determined 10-15 d after inoculation. This experiment was conducted twice with three replicates with 15 seedlings each.

Bacterial identification was conducted by 16S rRNA gene sequencing using the universal primers fD1 (5'-AGAGTTTGATCCTGGCTCAG-3') and rP2 (5'-ACGGCTACCTTGTACGACTT-3') (Weisburg et al., 1991). Polymerase chain reaction (PCR), DNA sequencing, and phylogenetic analyses were conducted as described by Kim et al. (2009). The 16S rRNA gene sequence of H26-2 (1,511 base pairs, bp) and H30-3 (1,381 bp) were analyzed and compared with related type species within the *Bacillus* genus using EzBioCloud (Yoon et al., 2017).

All experiments were performed as completely randomized designs. Statistical analysis of the data was conducted using Statistical Analysis Systems 9.02 software (SAS Institute, Cary, NC). Data from repeated experiments were pooled after confirming the homogeneity of variances with Levene (1960) test and were further statistically analyzed. The microbial cfu data were analyzed after logarithmic transformation. Analysis of variance was determined using the general linear model procedure. Means were separated using the least significant difference test at $P < 0.05$.

In our present study, we evaluated the effects of bacterial strains on growth promotion and mitigation of heat and drought stresses in Chinese cabbage plants. Strains H26-2 and H30-3 were selected from 46 strains, whereas *B. vallismortis* BS07M (Bacillo V3, NAPROBIOTEC Inc.,

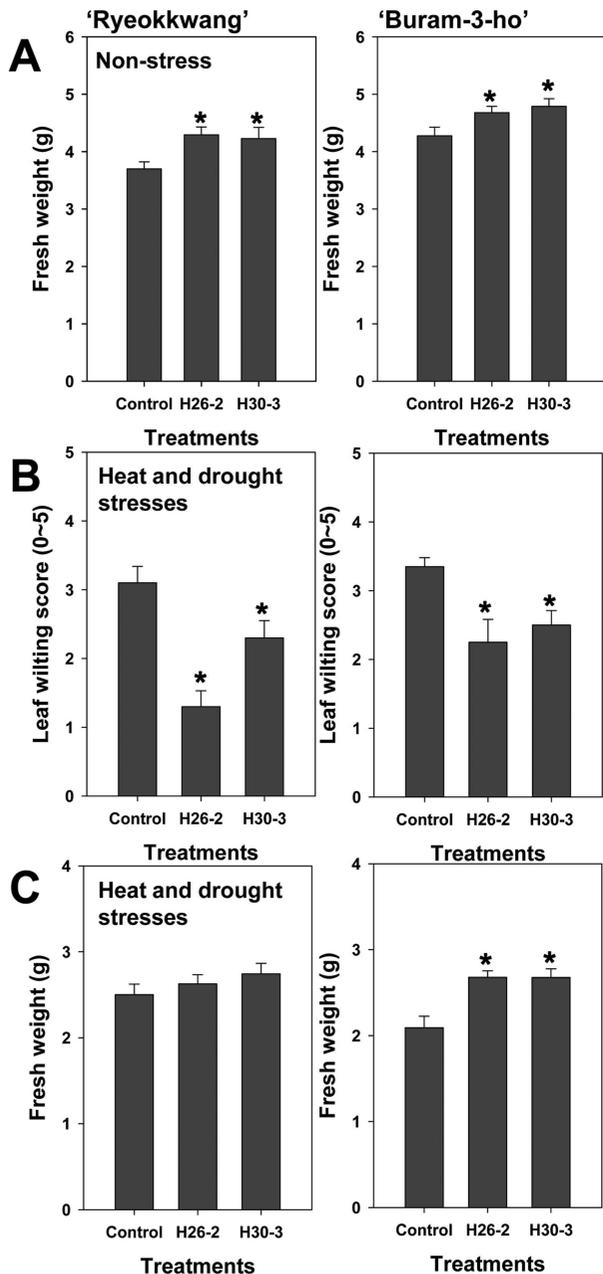


Fig. 1. (A) Growth promotion by *Bacillus aryabhattai* H26-2 and *Bacillus siamensis* H30-3 compared to 10 mM MgSO₄ solution (control) in Chinese cabbage ('Ryeokkwang' and 'Buram-3-ho') under non-stress condition; (B) Leaf wilting score (0-5) under heat and drought conditions; (C) fresh weight after re-watering of Chinese cabbages ('Ryeokkwang' and 'Buram-3-ho'). Plants were treated with bacterial strains H26-2 and H30-3, or 10 mM MgSO₄ solution (untreated control), one week later, heat and drought stresses were treated; Twenty-four h after end of stress treatment, fresh weight were assessed. Error bars indicate the standard errors of 20 replications from two experiments. An asterisk on bars with standard errors indicate significant ($P < 0.05$) differences between treatments according to the least significant difference test.

plant immune enhancer) was not appropriate as a positive control for alleviation of heat and drought stresses in Chinese cabbage (Supplementary Fig. 1), therefore we tested two strains for our further studies. Strains H26-2 and H30-3 had *in vitro* plant growth-promoting (PGP) traits including ACC deaminase activity, phosphate solubilization, production of indoleacetic acid, and EPS (data not shown). Moreover, strains H26-2 and H30-3 produced siderophore on CAS agar plates; but these strains did not produce HCN on TSA.

When strains H26-2 and H30-3 were applied to two cultivars of Chinese cabbage, these strains significantly ($P <$

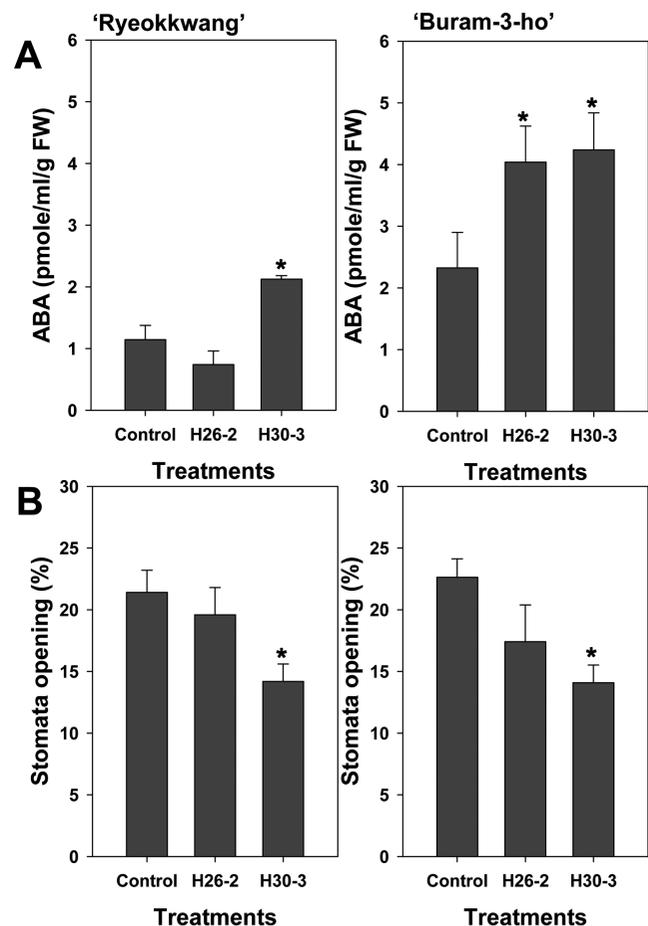


Fig. 2. (A) Abscisic acid (ABA) contents and (B) stomatal opening (%) in leaves of Chinese cabbages ('Ryeokkwang' and 'Buram-3-ho') at 24 h after heat and drought stresses treatment. Plants were drenched with strain H26-2 and H30-3, and 10mM MgSO₄ solution as untreated control, one week later, heat and drought stresses were performed. Error bars indicate the standard errors of eight (for ABA contents) or 16 (for stomatal opening) replications from two experiments. An asterisk on bars with error bars indicate significant ($P < 0.05$) differences between treatments according to the least significant difference test.

0.05) increased fresh weight of Chinese cabbage in both cultivars ('Ryeokkwang' and 'Boram-3-ho') compared to the control under non-stressed condition (Fig. 1A). Under heat and drought conditions, cultivar ('Ryeokkwang' and 'Boram-3-ho') and treatment (Control, H26-2 and H30-3) showed significant difference at $P = 0.0195$ and $P < 0.0001$, respectively. In 'Ryeokkwang', strains H26-2 and H30-3 significantly ($P < 0.05$) reduced leaf wilting compared to the control (Fig. 2B), while there was no difference in fresh weight 24 h after re-watering (Fig. 2C). In 'Boram-3-ho', tested two strains significantly ($P < 0.05$) reduced leaf wilting (Fig. 2B) and promote recovery compared to a control (Fig. 2C). These results implied two strains could be involved in both PGP and amelioration of abiotic stresses including heat and drought in Chinese cabbages. Some plant-associated bacteria are known as improving crop growth and development under unfavorable single stress including drought or saline soils (Kang et al., 2014; Singh and Jha, 2016). Bresson et al. (2013) and Ghosh et al. (2017) found that PGPR strain *Phyllobacterium brassicacearum* STM196 and *Pseudomonas putida* alleviates drought stress, respectively.

To understand the effects of the two strains on the regulation of water content in plants, we measured ABA content and stomatal opening in plant leaves under heat and drought stresses. Abscisic acid in leaves of Chinese cabbage treated with strain H30-3 was significantly ($P < 0.05$) higher than in the control 24 h after the beginning of stresses in both cultivars (Fig. 2A). Strain H26-2 also showed significantly ($P < 0.05$) higher ABA content in leaves of 'Boram-3-ho'. Based on stomatal observations, strain H30-3 reduced stomatal opening compared to the

control 24 h after stresses in both cultivars (Fig. 2B). On the other hand, stomatal opening (%) of H26-2 in leaves showed a decreased tendency; however, there was no statistically significant difference compared to the control (Fig. 2B). ABA is widely known as a stress hormone because of its rapid accumulation in plants and its role as a mediator under stress conditions. Under salinity and drought stresses, ABA helps to maintain the water status of plants by regulating guard cells and leaf growth and inducing expression of genes encoding enzymes and proteins related to dehydration tolerance (Zhang et al., 2006; Zhang et al., 2010; Zhu, 2002). Considering our present results regarding ABA contents and stomatal observation in plants, strains H26-2 and H30-3 induce the accumulation of ABA and it leads to water control in tested plants by regulation of stomatal opening under abiotic stresses.

Bacterial colonization and population maintainability in rhizosphere soils under unfavorable conditions can be important factors involved in plant-microbe-environmental interactions (Sandhya et al., 2009; Sherameti et al., 2008). When bacterial colonization in rhizosphere of Chinese cabbages was monitored during heat and drought stresses, strains H26-2 Rif and H30-3 Rif colonized the rhizosphere of both cultivars in Chinese cabbages, the bacterial colonization did not affect by days after stresses during stresses in both cultivar's rhizosphere (Table 1). Similarly, under non-stress conditions, colonization of H26-2 Rif and H30-3 Rif in rhizosphere soils did not change over the entire tested period, except for H26-2 Rif in 'Boram-3-ho' at 5 d (Table 1). According to comparison between non-stress and stresses, in 'Ryeokkwang', strain H26-2 Rif showed a higher colonization at 1 DAS under stress conditions

Table 1. Bacterial colonization in rhizosphere soils of Chinese cabbages treated with spontaneous rifampicin-resistant mutants, H26-2 Rif and H30-3 Rif under normal and stressed conditions

Cultivars	DAS ^a	Bacterial colonization [Log(cfu/g of soils) ^b			
		H26-2 Rif		H30-3 Rif	
		Non-stress	Stresses	Non-stress	Stresses
Ryeokkwang	1	4.83 ± 0.19 a ^c	5.33 ± 0.11 A*	4.26 ± 0.22 a	4.00 ± 0.01 A
	3	4.85 ± 0.20 a	4.96 ± 0.13 A	3.66 ± 0.08 a	4.01 ± 0.08 A*
	5	4.74 ± 0.19 a	4.98 ± 0.25 A	4.21 ± 0.21 a	3.83 ± 0.21 A
Boram-3-ho	1	5.17 ± 0.13 a*	4.62 ± 0.20 A	3.88 ± 0.18 a	4.04 ± 0.28 A
	3	5.35 ± 0.09 a	5.06 ± 0.28 A	3.83 ± 0.03 a	3.83 ± 0.10 A
	5	4.72 ± 0.08 b	4.56 ± 0.20 A	4.05 ± 0.13 a	4.04 ± 0.10 A

^a DAS; days after stress treatment

^b Bacterial colonization was determined by counting colony-forming unit (cfu) in pot soils at 1, 3, 5 d after stress treatment on Chinese cabbage plants.

^c Mean ± standard errors with same letters are not significantly different between DAS at each non-stress or stresses in cultivars according to the least significantly difference test at $P < 0.05$. An asterisk indicates significant difference between non-stress vs. stresses at each DAS according to the t-test. Experiments were conducted twice with four replicates each.

compared to the non-stress conditions; H30-3 Rif did at 3 DAS. In 'Buram-3-ho', H26-2 Rif reduced colonization at 1 DAS under the stress conditions compared to non-stress condition (Table 1). Strain H26-2 and H30-3 did not drastically affect rhizosphere colonization during stresses and they maintained their populations. Similarly, Sherameti et al. (2008) found that root colonization by microorganisms can play a pivotal role for increasing plant growth and productivity under non-optimal environmental conditions. Moreover, like our strains, EPS-producing bacteria can effectively colonize roots and increase the stability of root-adhering soils. Santaella et al. (2008) revealed that the EPS of *Rhizobium* sp. YAS34 are targeted to specific parts of roots and they are critical to root colonization.

To determine the effect of bacterial EPS on plant tolerance to abiotic stresses, EPS of H26-2 and H30-3 were tested in plate assay and *in planta*. In the plate assay in MS media amended with mannitol, cultivars did not affect

cotyledon greening ($P = 0.2485$) and number of lateral root ($P = 0.1191$) (Table 2). In 'Ryeokkwang', treatment did not show significant difference in cotyledon greening and number of lateral root, however, in 'Buram-3-ho', treatment show statistical significance in cotyledon greening ($P = 0.0231$) and number of lateral root ($P = 0.0494$) (Table 2). In 'Buram-3-ho' EPS of H26-2 increased cotyledon greening and EPS of H30-3 promoted number of lateral root compared to a control (Table 2). *In planta*, cultivars did not affect fresh weight ($P = 0.2937$) under non-stress condition, however, showed significant difference on leaf wilting score ($P < 0.0001$) and fresh weight ($P = 0.0006$) after re-watering under stress conditions (Table 2). In 'Ryeokkwang', treatments including control, EPS of H26-2, and EPS of H30-3 had influenced on only fresh weight ($P = 0.0491$) after re-watering under stress conditions (Table 2). EPS of H30-3 statistically recovered fresh weight compared to a control but leaf wilting scores were not different

Table 2. Effect of exopolysaccharides of H26-2 and H30-3 on cotyledon greening (%) and number of lateral roots after germination in plate assay, and fresh weight and leaf wilting score in Chinese cabbage under non-stressed or heat and drought stressed conditions

Cultivars, treatments	In plate assay ^a		<i>In planta</i> ^b		
	(MS media amended with 400 mM mannitol)		Non-stress	Stresses	
	Cotyledon greening (%)	Number of lateral root	Fresh weight (g)	Leaf wilting score (0-5)	Fresh weight (g)
Cultivars	F value = 1.37, P value = 0.2485	F value = 2.50, P value = 0.1191	F value = 1.13 P value = 0.2937	F value = 25.30 P value < 0.0001	F value = 13.53 P value = 0.0006
Ryeokkwang	67.5 ± 4.8 a	7.5 ± 0.4 a	8.8 ± 0.8 a	4.2 ± 0.1 a	1.0 ± 0.1 b
Buram-3-ho	75.4 ± 4.7 a	6.8 ± 0.3 a	7.8 ± 0.6 a	2.9 ± 0.2 b	3.3 ± 0.6 a
Ryeokkwang					
Treatments	F value = 0.58, P value = 0.5732	F value = 0.79, P value = 0.4644	F value = 0.32 P value = 0.7326	F value = 0.18 P value = 0.8370	F value = 3.49 P value = 0.0491
Control	67.5 ± 5.3 a	6.5 ± 0.3 a	9.6 ± 1.5 a	4.1 ± 0.1 a	0.8 ± 0.1 b
EPS of H26-2	75.0 ± 11.2 a	6.6 ± 0.5 a	8.9 ± 1.3 a	4.1 ± 0.1 a	1.0 ± 0.1 ab
EPS of H30-3	60.0 ± 12.7 a	7.3 ± 0.6 a	8.1 ± 1.3 a	4.2 ± 0.1 a	1.3 ± 0.2 a
Buram-3-ho					
Treatments	F value = 4.53, P value = 0.0231	F value = 2.95, P value = 0.0494	F value = 0.01 P value = 0.9860	F value = 4.00 P value = 0.0337	F value = 5.03 P value = 0.0164
Control	65.0 ± 5.6 b	6.5 ± 0.4 a	8.0 ± 1.0 a	3.8 ± 0.2 a	1.1 ± 0.4 b
EPS of H26-2	90.0 ± 10.0 a	7.6 ± 0.5 ab	7.8 ± 0.9 a	2.8 ± 0.4 ab	4.0 ± 1.0 a
EPS of H30-3	90.0 ± 5.6 a	8.4 ± 0.7 b	7.7 ± 1.2 a	2.3 ± 0.5 b	4.8 ± 1.1 a

^aThe numbers of seedlings with open and green leaves after seed germination were recorded at 3 and 4 d after seeding in repeated experiments; number of lateral root was evaluated at 17 d after seeding. Values ± standard errors of each cultivar and treatment in a column followed by same letters are not significantly different according to the least significant difference test at $P < 0.05$. Experiment were conducted twice with ten replicates each.

^bPlants were treated with EPS (10 µg/ml) of strains H26-2 and H30-3, and 10 mM MgSO₄ solution (control), one week later, heat and drought stress were treated; 24 h after end of stress treatment, leaf wilting score was assessed, and fresh weight after re-watering was assessed. Values ± standard errors of each treatment in a column followed by same letters are not significantly different according to the least significant difference test at $P < 0.05$. Experiments were conducted twice with eight replicates each.

(Table 2). In ‘Buram-3-ho’, treatments showed significance at leaf wilting score ($P = 0.0337$) and fresh weight ($P = 0.0164$) after re-watering under stress conditions. Treatment of EPS of H30-3 reduced leaf wilting and after re-wa-

tering, EPS of two tested strains promote recovery of fresh weight in ‘Buram-3-ho’ (Table 2). However, EPS of H26-2 and H30-3 did not affect plant growth in non-stress conditions in both cultivars (Table 2). Bacterial EPS can protect

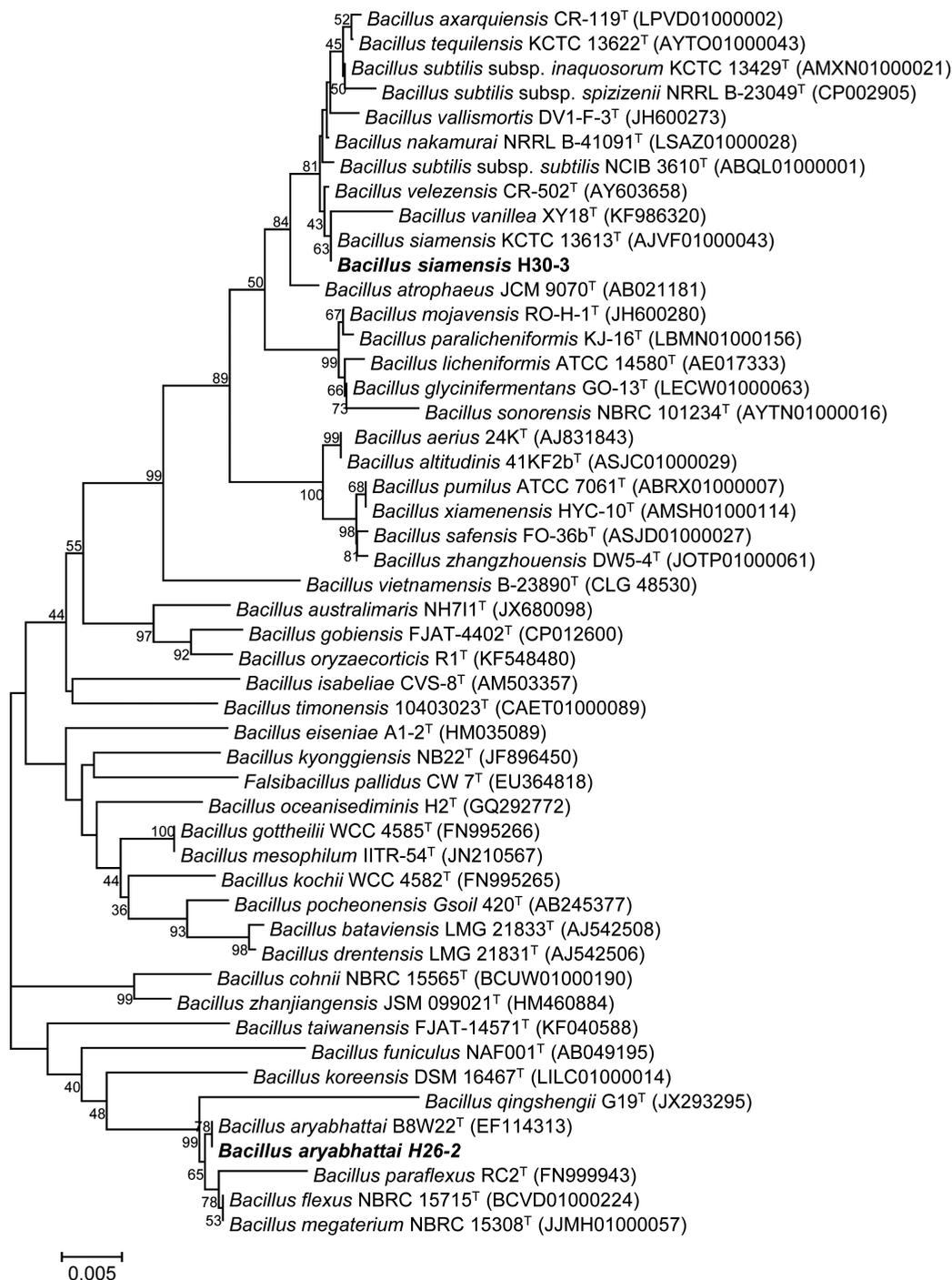


Fig. 3. A phylogenetic tree performed by the neighbor-joining method showing the relationship of *Bacillus aryabhatai* H26-2 and *Bacillus siamensis* H30-3 with other species of the genus *Bacillus* based on 16S rRNA sequences analysis. Numbers at the branching points are the bootstrap values (> 50%) for 1,000 replicates. A scale bar indicates 5 nucleotide substitutions per 1000 nucleotides of the sequence. Strains H26-2 and H30-3 are shown in bold type. T, type strain.

microorganisms from water stress by enhancement of water retention and regulation of diffusion (Chenu and Roberson, 1996); they also help to attach and colonize the roots (Bashan and Hoguein, 1997). Exopolysaccharides released into the soil by EPS-producing bacteria can form a protective capsule around soil aggregates and plants can increase water deficit tolerance under drought conditions (Sandhya et al., 2009). In sand amended with purified EPS, water content was considerably higher than that in untreated controls at a low water potential and bacterial EPS dissolved nutrients in the bacterial microenvironment during drought stress. Therefore, soil treated with EPS dries more slowly compared with controls under drought stress (Alami et al., 2000). These results suggest that EPS can hold water in the surrounding microenvironment during drought stress, buffering against changes during abiotic stresses. Our results showed that EPS of strain H30-3 effectively reduced leaf wilting under heat and drought stresses and helped plants to recover after re-watering in ‘Boram-3-ho’ Chinese cabbage. In ‘Ryeokkwang’, extracted EPS of the two strains did not mitigate leaf wilt. Our results suggest that EPS of strain H30-3 can cause cultivar-dependent responses to heat and drought stresses in Chinese cabbage, indicating that EPS of strains H26-2 or H30-3 can interact with plants specifically. On the other hand, tested bacterial EPS did not affect plant growth in non-stress conditions. Therefore, EPS of H30-3 can be one of bacterial determinants for alleviating heat and drought stresses.

Strain H30-3 significantly reduced incidence of soft rot caused by *P. carotovorum* subsp. *carotovorum* PCC21 only in ‘Boram-3-ho’, however, strain H26-2 did not show biocontrol activity against soft rot in both cultivars of Chinese cabbages. As similar results of effects of EPS on heat and drought stresses, strain H30-3 showed cultivar-different response to soft rot suppression. In our further study, whether EPS or other bacterial determinants of strain H30-3 can cause cultivar-different response to disease suppression; how to specify and recognize cultivar will be studied.

Strains H26-2 and H30-3 were identified as *Bacillus aryabhatai*, and *B. siamensis*, respectively, based on 16S rRNA sequence analysis (data not shown). Strain H26-2 (1,511 bp) showed 100% similarity to *B. aryabhatai* B8W22^T (EF114313); strain H30-3 (1,381 bp) exhibited 99.93% similarity to *B. siamensis* KCTC 13613^T (AJVF01000043). These strains were deposited as *B. aryabhatai* H26-2 (KACC 92165P) and *B. siamensis* H30-3 (KACC 92187P) in the Korean Agricultural Culture Collection (KACC) of the National Institute of Agricultural Biotechnology (Wanju, Korea). Among diverse PGPR, *Bacillus* spp. are widely known as potential candidates for tol-

Table 3. Biocontrol activity of tested bacterial strains against soft rot caused by *Pectobacterium carotovorum* subsp. *carotovorum* PCC21 in two cultivars of Chinese cabbages

Treatment	Disease incidence (%)	
	Ryeokkwang	Boram-3-ho
Control	60.0 ± 4.7 ab	72.0 ± 6.8 a
BTH	46.0 ± 8.5 b	46.0 ± 7.9 b
H26-2	62.0 ± 8.7 ab	60.0 ± 8.4 ab
H30-3	70.0 ± 7.5 a	40.0 ± 7.9 b

^aValues ± standard errors of treatments in a column followed by same letters are not significantly different according to the least significant difference test at $P < 0.05$. Experiment was conducted twice with three replicates with 15 seedling each.

erance against abiotic stresses such as drought and extreme temperature (Tiwari et al., 2017). They have structural and physiological advantages including a multi-layered cell wall and secretion of various antibiotics, peptides, signal molecules, and extracellular enzymes. These traits can help their survival under different environmental stress conditions for a long period (Kumar et al., 2012). Bacterial structural characters, such as the thick cell walls of *Bacillus* species and EPS production, could act to maintain populations under stress conditions.

Taken together, our results suggest that *B. aryabhatai* H26-2 and *B. siamensis* H30-3 have effects on both plant growth promotion and alleviation of heat and drought stresses in Chinese cabbage. To cope with the stresses, strains H26-2 and H30-3 reduced leaf wilting by increasing ABA contents and reducing stomatal opening in Chinese cabbages. Moreover, the two strains were not altered for colonization in the rhizosphere and maintained their population during stresses. According to our results, EPS of strain H30-3 might interact with Chinese cabbages cultivar specifically, and it is pivotal one of bacterial determinants in mitigating heat and drought stresses in ‘Boram-3-ho’ Chinese cabbage. As well as mitigation of abiotic stresses, strain H30-3 could reduce soft rot incidence (%) in ‘Boram-3-ho’ Chinese cabbage. Further studies to elucidate the mechanisms related to physiological and transcriptional changes in plants and cultivar-dependent response by strain H30-3 will be studied to better understand mitigation of abiotic and biotic stresses conferred by plant-bacteria interactions.

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