

Inhibition of *Fusarium oxysporum* f. sp. *nicotianae* Growth by Phenylpropanoid Pathway Intermediates

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Fusarium wilt in tobacco caused by the fungus *Fusarium oxysporum* f. sp. *nicotianae* is a disease-management challenge worldwide, as there are few effective and environmentally benign chemical agents for its control. This challenge results in substantial losses in both the quality and yield of tobacco products. Based on an *in vitro* analysis of the effects of different phenylpropanoid intermediates, we found that the early intermediates *trans*-cinnamic acid and *para*-coumaric acid effectively inhibit the mycelial growth of *F. oxysporum* f. sp. *nicotianae* strain FW316F, whereas the downstream intermediates quercetin and caffeic acid exhibit no fungicidal properties. Therefore, our *in vitro* screen suggests that *trans*-cinnamic acid and *para*-coumaric acid are promising chemical agents and natural lead compounds for the suppression of *F. oxysporum* f. sp. *nicotianae* growth.

Keywords: fungicidal, *Fusarium oxysporum*, *para*-coumaric acid, tobacco, *trans*-cinnamic acid

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Nonpathogenic and pathogenic strains of the soil-borne fungus *Fusarium oxysporum* are ubiquitous in agricultural soils worldwide, and pathogenic sub-specific strains (i.e., formae speciales; f. sp.) impact a number of economically important crops (Gordon, 2017). *F. oxysporum* f. sp. *nicotianae* (FON) is the causal agent of Fusarium wilt disease (FW) in tobacco, which causes chlorosis, wilting and necrosis of tobacco leaves that leads to stunted growth and death. FON affects the global tobacco crop due to high seed transmissibility and its ability to survive in soil without a host plant for years (LaMondia, 2015).

To date, the use of FW-resistant cultivars is the most effective FW management strategy. However, breeding programs are time-consuming and costly, and resistant cultivars are not entirely immune to FW infection under high disease pressure (LaMondia, 1995). Therefore, FW chemical management, through pre-planting soil fumigation with methyl bromide or chloropicrin, is commonplace in many plant production systems (Duniway, 2002). However, methyl bromide is now recognized as a Class I ozone-depleting substance and its use has been phased out (UNEP Ozone Secretariat, 2020) and chloropicrin is a strong irritant that decreases tobacco quality and often does not appreciably decrease disease incidence (Fisher et al., 2018; LaMondia, 2015; Schneider et al., 2003; Xie et al., 2015). Other classes of fungicides such as succinate dehydrogenase inhibitors and demethylation inhibitors have shown promise for the control of soil-borne fungal pathogens (Masiello et al., 2019; Miller et al., 2020) but these compounds have not been systematically tested on *Fusarium* species. Moreover, the wide-spread use of fungicides with single-site mechanisms of action necessitates

a diverse set of chemical management tools to slow down the evolution of fungicide-resistant pathogens (Avenot and Michailides, 2010; Lucas et al., 2015). Therefore, there is a continuing demand for cost-effective and environmentally benign chemicals for FW disease management in tobacco and other cropping systems.

The demand for novel fungicides and ecologically non-disruptive FW treatments led to the expansion of research focused on the identification of natural products which can control FW in various plant systems (Bubici et al., 2019; Hazir et al., 2017; Steinkellner and Mammerler, 2007; Wu et al., 2008). This research showed that plant secondary metabolites synthesized by the phenylpropanoid pathway (PPP) can inhibit the growth of some *F. oxysporum* strains (Steinkellner and Mammerler, 2007). The PPP begins with the conversion of phenylalanine to *trans*-cinnamic acid (*t*-CA) by phenylalanine ammonia-lyase (Vogt, 2010). *t*-CA is processed by cinnamic acid 4-hydrolase into *para*-coumaric acid (*p*-CA). *p*-CA is converted into *para*-coumaroyl-CoA, a major branch-point metabolite of the PPP, which is a precursor of various compounds such as caffeic acid and quercetin (Fig. 1) (Vogt, 2010).

Phenylpropanoids (PPs) have many beneficial properties which have attracted intense research interest (Neelan et al., 2019). Analyses of the fungicidal properties of PPs emphasized the research centered on *t*-CA (Guzman, 2014). Currently, there is no consensus on *t*-CA fungicidal activity even in a single species. For example, studies conducted on *F. oxysporum* indicate that *t*-CA can both promote and inhibit mycelial growth (Wu et al., 2008; Ye et al., 2004; Zhao et al., 2018), and the current hypothesis is that the growth inhibitory activity of *t*-CA on FW-causing pathogens varies due to the high physiological variability of *F. oxysporum* subspecies (Bock et al., 2014; Hao et al., 2010; Hazir et al., 2017; Morales et al., 2017; Nirmaladevi et al., 2016; Wu et al., 2008). The impact of other PPs on FW-causing pathogens has also been analyzed, although not in great detail. For example, *p*-CA and caffeic acid

have both been shown to decrease the sporulation and germination of *Fusarium oxysporum* f. sp. *niveum* (Hao et al., 2010).

Because the wide-spread interest in the identification of environmentally benign FON control is in stark contrast with the lack of information on the anti-FON activity of natural products, here we lay the groundwork for research into the efficacy of PPP intermediates in controlling FON growth, and demonstrate that *t*-CA and its direct downstream derivative, *p*-CA, effectively mitigate the growth of the FON strain FW316F.

FON strain FW316F was single-spore purified from the stem of a TN 97 tobacco plant growing at the University of Kentucky-Spindletop Farm in Lexington, and identified as FON based on conidia morphology (Leslie et al., 2006). For the preparation of the FON inoculum used in all experiments described in this study, FON was grown for two weeks in the dark at 25°C on potato dextrose agar (PDA; Difco, Detroit, MI, USA), scrapped into 15 ml of distilled water, vortexed and briefly centrifuged to pellet mycelial debris. To make the final inoculum, the supernatant was diluted until the solution absorbance at 600 nm was 0.1 ± 0.02 . An aliquot of each inoculum was transferred onto a hemocytometer (C-chip DHC-N01, SKC Inc., Cheonan, Korea), and micrographs were captured using an Olympus SZX12 microscope. The microconidia/ml was determined to be $257,857 \pm 65,183$ (mean \pm standard deviation, $n = 6$) using the open-source program ImageJ (<http://www.imagej.net>).

Considering that previous studies demonstrated the antifungal activity of *t*-CA (Guzman, 2014), we initiated our analyses with this compound. To that end, we prepared a control PDA plate (no additives), test plates (with 0.1 mM and 1 mM *t*-CA; *t*-CA obtained from Sigma-Aldrich, St. Louis, MO, USA), a solvent control plate (with 0.55% dimethyl sulfoxide [DMSO]) and a positive control plate (with 0.2% of the commercial fungicide Cleary 3336F). FON inoculum (100 μ l) was spread onto the prepared plates, which were then incubated in the dark for four days

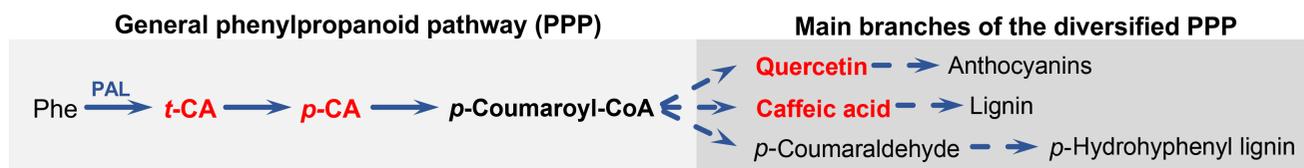


Fig. 1. Simplified scheme of the phenylpropanoid biosynthetic pathway. The phenylpropanoid biosynthetic pathway (PPP) channels carbon from L-phenylalanine (Phe) to a large number of polyphenols. The general PPP begins with Phe which is converted to *trans*-cinnamic acid (*t*-CA) by the enzyme phenylalanine ammonia lyase. *t*-CA is then reduced to *para*-coumaric acid (*p*-CA) which is activated to yield *p*-coumaroyl-CoA. *p*(*ara*)-Coumaroyl-CoA is the branch-point metabolite of all flavonoid and non-flavonoid polyphenols. The compounds tested in this study are highlighted in red.

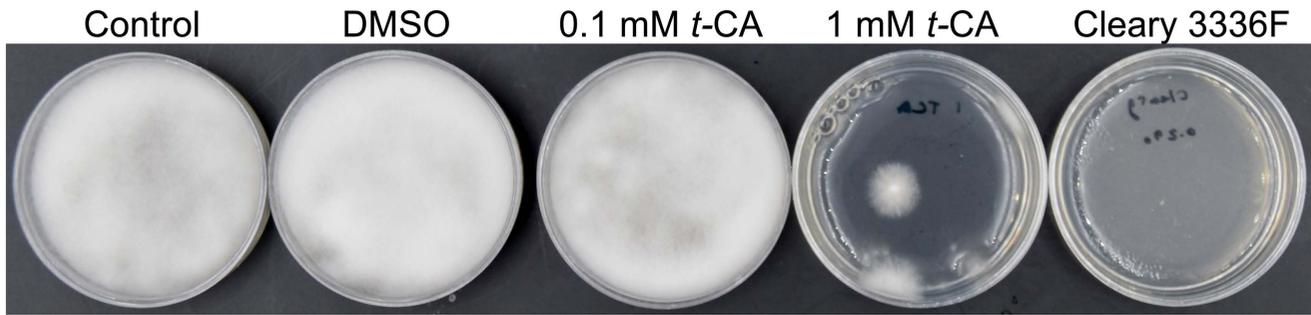


Fig. 2. Trans-cinnamic acid (*t*-CA) inhibits the growth of *Fusarium oxysporum* f. sp. *nicotianae*. *Fusarium oxysporum* f. sp. *nicotianae* inoculum was spread on potato dextrose agar plates without additives (control), with 0.5% dimethyl sulfoxide (DMSO; as a solvent control for the 1 mM *t*-CA treatment), with 0.1 mM or 1 mM *t*-CA, and with 0.2% of the commercial fungicide Cleary 3336F. Plates were incubated in the dark at room temperature for 4 days and photographed.

at room temperature and evaluated (Fig. 2). As expected, the 0.2% Cleary 3336F treatment completely inhibited FON growth. Mycelia grew equally on plates without additives and plates with the solvent control, suggesting that the *t*-CA solvent has no effect on fungal growth at the tested doses. The *t*-CA dose-response plates revealed that whereas 0.1 mM *t*-CA had no antifungal properties, 1 mM *t*-CA led to nearly complete inhibition of mycelial growth, proving to be almost as effective as the 0.2% Cleary 3336F treatment (Fig. 2). Therefore, we concluded that *t*-CA has

dose-dependent antifungal activity against the FON strain FW316F.

Because *t*-CA is the first compound synthesized in the PPP, we next asked if *t*-CA itself or if a downstream metabolite is responsible for the observed fungicidal activity. To that end, we tested three compounds: (1) *p*-CA, a direct downstream derivative of *t*-CA, (2) the late downstream intermediate caffeic acid, which is a lignin precursor, and (3) quercetin, a late intermediate that belongs to the flavanol class of the PP compounds named flavonoids. To compare

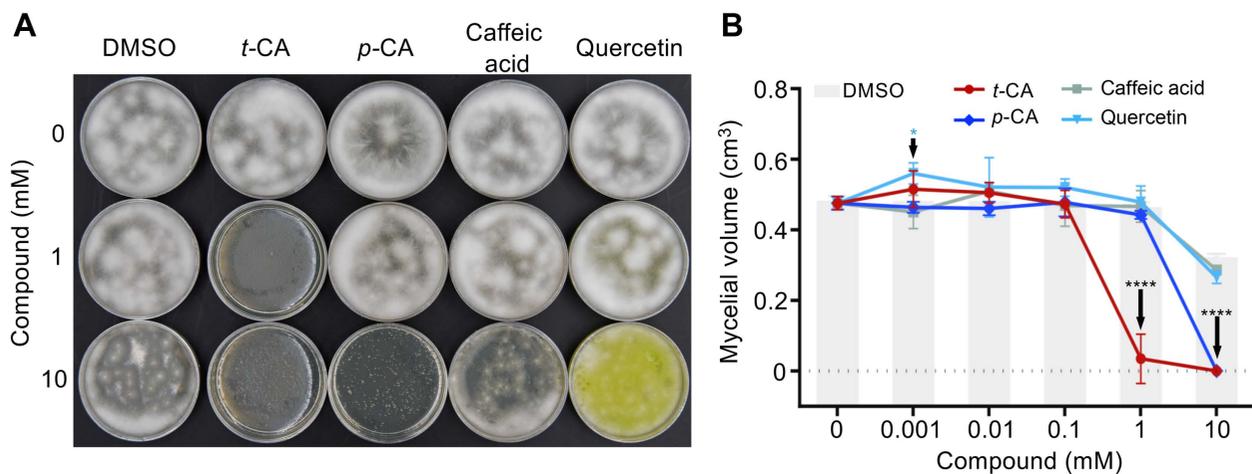


Fig. 3. Effects of phenylpropanoids on *Fusarium oxysporum* f. sp. *nicotianae* (FON) growth. (A) Trans-cinnamic acid (*t*-CA) and *para*-coumaric acid (*p*-CA) inhibit the growth of FON. One hundred microliters of FON inoculum was spread on potato dextrose agar (PDA) media containing the denoted concentrations of compounds and incubated in the dark for 4 days. Quercetin is a yellow-colored plant pigment and at higher concentrations, it stains the media. (B) Quantification of the growth-inhibitory effects of phenylpropanoids. Solvent controls (dimethyl sulfoxide [DMSO]; gray bars) contained the concentration of DMSO used to prepare the compound solution at the labeled tick-mark (e.g., for 0.001 mM, 0.0005% DMSO was used as a control; for 0.01 mM, 0.005% DMSO). The mycelium from each plate was scrapped into tubes containing 1 ml of water after four days of growth on the PDA media. The mycelial suspension was centrifuged at $21,330 \times g$ for 5 min.. The mycelial pellet was then leveled, photographed and the mycelial volume was measured. Data are presented as mean \pm SD ($n = 3$). * $P < 0.05$, **** $P < 0.0001$. *p*-values on the graph are *post-hoc* comparisons of the DMSO solvent controls to the point(s) indicated by the black arrows. The light blue asterisk indicates the comparison between the 0.001 mM quercetin treatment and the DMSO control.

the FON-inhibitory capacity of these three downstream compounds to *t*-CA, we prepared PDA media supplemented with *t*-CA, *p*-CA (Sigma-Aldrich), caffeic acid (3,4-dihydroxy-cinnamic acid, Sigma-Aldrich), and quercetin (5,7,3',4'-flavon-3-ol, Sigma-Aldrich) at concentrations of 0.001 mM, 0.01 mM, 0.1 mM, 1 mM and 10 mM. The FON inoculum (100 μ l) was spread on the plates, and plates were incubated in the darkness at room temperature. After four days of incubation, mycelial growth was inhibited on media containing 1 mM and 10 mM *t*-CA as well as on plates containing 10 mM *p*-CA (Fig. 3A).

To quantify the inhibitory effect of the tested PPs, mycelia were scrapped into 1 ml of water, pelleted by centrifugation, leveled with a spatula, photographed and the mycelial volume was measured using ImageJ software. After three independent replicates, the collected data ($n = 3$) were analyzed using GraphPad Prism 6.0 (two-way ANOVA followed by Tukey's multiple comparisons *post-hoc* test, GraphPad Software Inc., San Diego, CA, USA). $P \leq 0.05$ from the *post-hoc* tests were considered significant and

are marked on the graphs. As previously observed (Fig. 2), there was no significant difference in growth between the control plates and most of the solvent (DMSO) control plates (Fig. 3B). The exception was the DMSO control for the 10 mM treatments, which led to an $\sim 34\%$ reduction in mycelial growth. However, 10 mM *t*-CA and 10 mM *p*-CA completely inhibited mycelial growth, indicating that at this dose, the effect of the test compound was stronger than the effect of the solvent. Quantitative analyses showed that the inhibition of mycelial growth was statistically significant when compared with the solvent control in the 1 mM *t*-CA, 10 mM *t*-CA, and 10 mM *p*-CA treatments (Fig. 3B). Conversely, caffeic acid treatments were not significantly different from the control at any tested doses. Quercetin also did not show any anti-FON activity. In fact, the lowest tested dose of quercetin (0.001 mM) led to a slight increase in mycelial volume (Fig. 3B). Although the exact mechanism of this growth promotion was not further analyzed, quercetin is known to be a strong antioxidant (Xu et al., 2019), and as such, it may have improved mycelial growth by

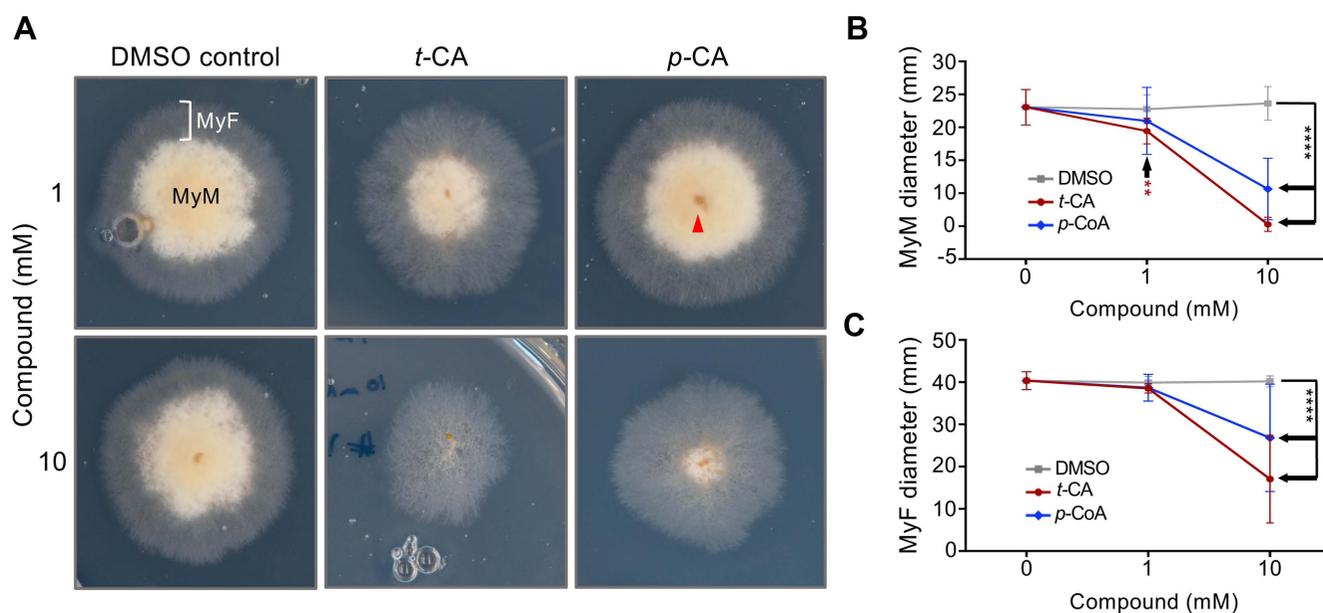


Fig. 4. Trans-cinnamic acid (*t*-CA) and para-coumaric acid (*p*-CA) inhibit *Fusarium oxysporum* f. sp. *nicotianae* (FON) mycelial growth originating from tobacco seed surface transmission. (A) *t*-CA and *p*-CA decrease mycelial front diameter originating from FON-contaminated KT97 seeds. Tobacco seeds of the FON-susceptible variety KT97 were sterilized and suspended in 1 ml of sterile distilled water and 100 μ l of FON inoculum was added to the solution. *t*-CA, *p*-CA or an equivalent amount of dimethyl sulfoxide (DMSO) were then added to the seed/FON mix at the denoted concentrations. After briefly vortexing the mix, the seeds were sown on tobacco growth media. Plates were then incubated for 4 days in a growth chamber under continuous light after which they were photographed. Photographs are 40 \times 40 mm. Red arrowhead points to a tobacco seed. MyM, mycelial mat; MyF, mycelial front. (B, C) *t*-CA and *p*-CA decrease mycelial mat diameter originating from FON contaminated KT97 seeds. Experiment was performed as in A and data were gathered from photographs using ImageJ. Data presented are mean \pm SD ($n = 15$). **** $P < 0.0001$. P -values on the graph are the result of *post-hoc* comparisons of the DMSO solvent controls to the point(s) indicated by the black arrows. The red asterisks in B indicate the comparison between the 1 mM *t*-CA treatment and the DMSO control.

reducing oxidative stress. Taken together, our data suggest that the simple, early PPP intermediates *t*-CA and *p*-CA effectively inhibit the growth of FON strain FW316F, and of the tested intermediates, *t*-CA has the highest potency.

Due to the high transmissibility of FON in seeds (La-Mondia, 2015), we opted to test if *t*-CA and *p*-CA can be used to inhibit FON spread stemming from surface-contaminated tobacco seeds. To that end, we aliquoted 10 ± 2 mg of surface-sterilized seeds of the FON-susceptible tobacco variety TN 97 (FW. Rickard Seeds, Inc., Winchester, KY, USA) into tubes, added 100 μ l of FON inoculum to each tube and then briefly vortexed the mix. A volume of *t*-CA or *p*-CA DMSO stock solution was added to the tubes to attain concentrations of 1 mM or 10 mM. For solvent controls, an equivalent volume of DMSO was added to the seed/FON inoculum mix. The treated FON-contaminated seeds were sown on full-strength Murashige and Skoog media containing 3% sucrose and 0.7% Phytoagar (Phyto-Tech, Shawnee Mission, KS, USA) and incubated for 4 days in a growth chamber under continuous light (17-Watt fluorescent lights, color temperature 4100) at a constant temperature (28°C). The control seeds developed thick, opaque mycelial mats (MyM) and thinner, translucent mycelial fronts (MyF) around the periphery of the MyM, whereas the MyM around the seeds treated with 1 mM and 10 mM *t*-CA and 10 mM *p*-CA was reduced in size (Fig. 4A).

To quantify the impact of the *t*-CA and *p*-CA seed treatments on FON seed-surface transmission, we measured the MyF and MyM diameters using ImageJ software (Fig. 4B and C). After three independent replicates, the collected data ($n = 15$) were analyzed using GraphPad Prism 6.0. Two-way ANOVA was used to compare means between groups, and Tukey's multiple comparisons was used as a *post-hoc* test. $P \geq 0.05$ from the *post-hoc* tests were considered significant. Compared to the DMSO treatments, 1 mM and 10 mM *t*-CA led to significant, dose-dependent reductions in MyM and MyF diameters. Similar to the *in vitro* growth experiments (Fig. 3), *p*-CA was less effective than *t*-CA in decreasing the spread of FON from infected seeds, as indicated by the decrease in MyM and MyF diameter only at the highest tested dose (Fig. 4B and C). We observed complete inhibition of mycelial growth around infected seeds in $40\% \pm 2\%$ of cases when treated with 10 mM *t*-CA and in $20\% \pm 2\%$ of 10 mM *p*-CA treatments. These results suggest that compared to *t*-CA, *p*-CA is approximately half as effective at inhibiting FON mycelial growth. Collectively, our results indicate that the PP intermediates *t*-CA and *p*-CA are both promising natural products for the control of FON spread in tobacco produc-

tion systems, and that *t*-CA is the most effective PP tested for the inhibition of the mycelial growth of FON strain FW316F.

Natural products have been a strong lead in the discovery and development of novel fungicides (Daoubi et al., 2005; Li et al., 2018; Musso et al., 2012). In the present study, we demonstrate that both *t*-CA and *p*-CA are potentially useful anti-FON agents and possible lead compounds for the development of new synthetic fungicides. More research is required to determine if the antifungal capacity of *t*-CA and *p*-CA generalizes to all FON strains. Of equal importance is the testing of the FON-inhibitory capacity of PPP intermediates on susceptible tobacco plants exposed to FON. The use of PPP derivatives to mitigate FON infection has potential limitations. For example, *t*-CA and *p*-CA are poorly soluble in aqueous solutions (Mota et al., 2008), which complicates the creation of formulations for field applications. Furthermore, using plant metabolites to control FON may cause off-target effects on plant growth and development. However, the strong fungicidal activity of *t*-CA and *p*-CA, their availability, and low environmental risk give a strong impetus for a detailed understanding of the mechanism of their FON growth-inhibitory activity, which in turn may aid in the development of novel environmentally friendly fungicides.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

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