



# Modulation of Reactive Oxygen Species to Overcome 5-Fluorouracil Resistance

Kyung-Soo Chun<sup>1</sup> and Sang Hoon Joo<sup>2,\*</sup>

<sup>1</sup>College of Pharmacy, Keimyung University, Daegu 42601,

<sup>2</sup>Department of Pharmacy, Daegu Catholic University, Gyeongsan 38430, Republic of Korea

## Abstract

5-Fluorouracil (5-FU) remains to be an important chemotherapeutic drug for treating several cancers when targeted therapy is unavailable. Chemoresistance limits the clinical utility of 5-FU, and new strategies are required to overcome the resistance. Reactive oxygen species (ROS) and antioxidants are balanced differently in both normal and cancer cells. Modulating ROS can be one method of overcoming 5-FU resistance. This review summarizes selected compounds and endogenous cellular targets modulating ROS generation to overcome 5-FU resistance.

**Key Words:** Reactive oxygen species, Cancer, Resistance, 5-Fluorouracil

## INTRODUCTION

Despite the introduction of targeted anticancer therapy, 5-fluorouracil (5-FU) remains an important chemotherapeutic drug for treating several cancers, including colorectal, breast, and gastric cancer. 5-FU's cytotoxic mechanism involves the inhibition of thymidylate biosynthesis or the misincorporation of fluorinated nucleotides into newly synthesized DNA or RNA (Longley *et al.*, 2003). It can be effective in the treatment of cancer when targeted therapy is unavailable. As described in previous studies, the development of prodrugs such as capecitabine has improved the limitation of 5-FU due to poor oral absorption (Pazdur *et al.*, 1998). Furthermore, combination chemotherapy improved 5-FU's anticancer effect, as demonstrated by FOLFOX (folinic acid, 5-FU, and oxaliplatin) and FOLFIRI (folinic acid, 5-FU, and irinotecan) (Souglakos *et al.*, 2006). Combining chemotherapeutics with different mechanisms could overcome the heterogeneity of tumor cells and decrease the development of resistance (Frei *et al.*, 1998). Nevertheless, the overall response rate remains less than 50% (Mehrzhad *et al.*, 2016) due to the cells being resistant to chemotherapy.

Studies have been conducted to elucidate the 5-FU resistance mechanism described elsewhere (Blondy *et al.*, 2020). The generation of reactive oxygen species (ROS) frequently correlates with the induction of apoptosis in many cancer cells;

modulation of ROS may be one mechanism by which cancer cells avoid the cytotoxicity induced by 5-FU (Mates and Sanchez-Jimenez, 2000). For example, in human lung carcinoma cells NCI-H1299, the expression of reactive oxygen modulator 1 (Romo1) is elevated, and the cellular level of ROS is high. At the same time, tumor cells maintain high levels of antioxidant enzymes and antiapoptotic Bcl-2 family proteins, most likely to reduce oxidative stress (Hwang *et al.*, 2007). This implies that cancer cells prefer a high level of ROS while keeping the protective mechanisms running to minimize the unwanted toxicity of ROS.

ROS have a versatile role in cancer cell biology (Liou and Storz, 2010). When elevated, ROS are thought to act as mitogens, inducing cancer cell proliferation (Torres and Forman, 2003). DNA damage from oxidative stress may lead to mutations that can either activate oncogenes or inactivate tumor suppressor genes (Wei, 1992). ROS production is minimal in normal cells, and antioxidant functions effectively remove ROS (Fig. 1A). Increased ROS production is frequently observed in cancer cells with a poor prognosis (Kumar *et al.*, 2008). Cancer cells maintain a relatively high level of ROS, likely due to the tumor-promoting effects of ROS such as angiogenesis (Ushio-Fukai and Nakamura, 2008), metastasis (Nishikawa, 2008), and proliferation (Juhász *et al.*, 2017). As shown in Fig. 1B, cancer cells increase the level of antioxidant systems in response to elevated levels of ROS to protect

**Open Access** <https://doi.org/10.4062/biomolther.2022.017>

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Received Jan 28, 2022 Revised Mar 29, 2022 Accepted Mar 30, 2022

Published Online Apr 20, 2022

**\*Corresponding Author**

E-mail: sjoo@cu.ac.kr

Tel: +82-53-850-3614, Fax: +82-53-359-6729

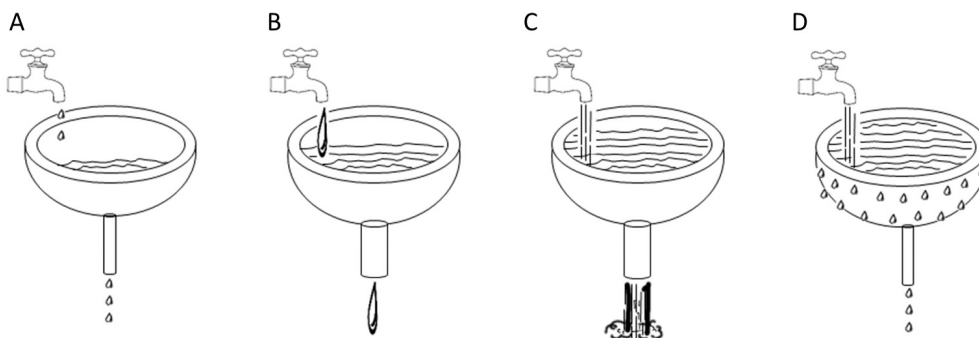
themselves from oxidative stress (Gorrini *et al.*, 2013). Many anticancer drugs, including 5-FU, induce high levels of ROS to exert cytotoxic effects. Cancer cells adapt to the escalated ROS level by expressing even more antioxidant systems (Fig. 1C) (Liu *et al.*, 2016b). When there is insufficient protection from high levels of ROS, cancer cells may not survive (Fig. 1D).

In this study, we summarized our understanding of natural and synthetic compounds (Table 1) and identified possible cellular targets involved with the modulation of cellular ROS levels to overcome 5-FU resistance.

## NATURAL/SYNTHETIC COMPOUNDS THAT MODULATE ROS TO OVERCOME 5-FU RESISTANCE

### Metal chelators

Tetrathiomolybdate, a copper-chelating drug, was initially developed as an anticopper and antiangiogenic agent to treat Wilson's disease (Brewer *et al.*, 1991). Interestingly, it enhances the activity of the anticancer drug doxorubicin, a DNA intercalator in ovarian cancer cells (Kim *et al.*, 2011). Tetrathiomolybdate increased the cytotoxicity of doxorubicin at subcytotoxic levels, likely by targeting antioxidant enzymes such as



**Fig. 1.** Balance between ROS production and antioxidant function. (A) ROS (water in the figure) are produced by various mechanisms (drawn as a water tap), and antioxidant function (drawn as a drain) effectively removes them, allowing physiological ROS levels to remain low. (B) Increased ROS generation is frequently observed in cancer cells, and cancer cells increase the level of antioxidant functions accordingly. The cellular level of ROS increases but not to toxic levels. (C) Even higher antioxidant function accompanies ROS overproduction when cancer cells adapt to chemotherapy. (D) Decrease of antioxidant function may result in cellular toxicity.

**Table 1.** Selected compounds increasing ROS generation to overcome 5-FU resistance

Compound	Cell/tissue type	Effects/Mechanisms
tetrathiomolybdate	Ovarian cancer cells	Stress-mediated apoptosis↑, activation of JNK and p38 MAPK↑
TPEN	Colon cancer HCT116 cells	Mitochondrial membrane potential (MMP)↓
apigenin	Hepatocellular carcinoma cells	Mitochondrial apoptosis↑
Polyphenolics from quince	Colon cancer cells LS174	NF-κB activation↓, cell cycle progression↓, angiogenesis↓
kaempferol	Colon cancer cells LS174	Activation of STAT3↓, angiogenesis↓
shikonin	Gastric cancer SGC-7901	Translocation of AIF and Endo G into nucleus
proanthocyanidin	Breast cancer MDA-MB-231 cells	G2/M cell cycle arrest↑, MMP↓
B63 (curcumin analog)	Gastric cancer cells SGC-7901 etc.	Expression of Thioredoxin reductase 1↓
dimethoxycurcumin	Colon cancer cells SW480, SW620	Expression of Bax and cyt C↑, expression of Bcl-2↓
Sanguisorba officinalis L. radix	Colorectal cancer cells RKO, HCT116	Bax/Bcl-2 disruption↑, autophagy↑
manuka honey	Colon cancer cells HCT116	Expression of EGFR, HER2, Akt and mTOR↓
emodin	Breast cancer MCF7 cells	Expression of E2F1 and NRPARP↓
gypenoside	Colorectal cancer cells SW-480, SW-620 and Caco2	DNA damage induction↑, expression of p53↑
tubeimoside-I	Colorectal cancer cells SW480, SW620, HCT116, and RKO	Activation of AMPK↑
oridonin	Colorectal cancer cells HCT115	Activation of JNK/c-Jun pathway↓
Coptis herb extracts	Lung cancer A549 cells	ROS↑
manahine	Colorectal cancer cells HCT116, SW480	Expression of PTEN and p53 in nucleus↑
caffeine	Liver cancer cells HepG2, HLF, Huh7, etc.	Cleavage of PARP↑, expression of Bcl-2 and Bcl-xL↓
selenocysteine	Skin cancer cells A375	Activation of ERK/Akt signaling↓
allicin	Liver cancer cells SK-Hep-1, BEL-7402	ROS↑, MMP↓
3-bromopyruvate	Liver cancer cells SNU449, Hep3B	ROS↑, MMP↓

copper/zinc-superoxide dismutase (SOD). Furthermore, by generating ROS, tetrathiomolybdate increased the cytotoxicity of several anticancer drugs, including 5-FU and mitomycin C (Kim *et al.*, 2012). The production of ROS induced by tetrathiomolybdate resulted in the activation of stress-mediated apoptosis, JNK, and p38 mitogen-activated protein kinase (MAPK), which increased cytotoxicity.

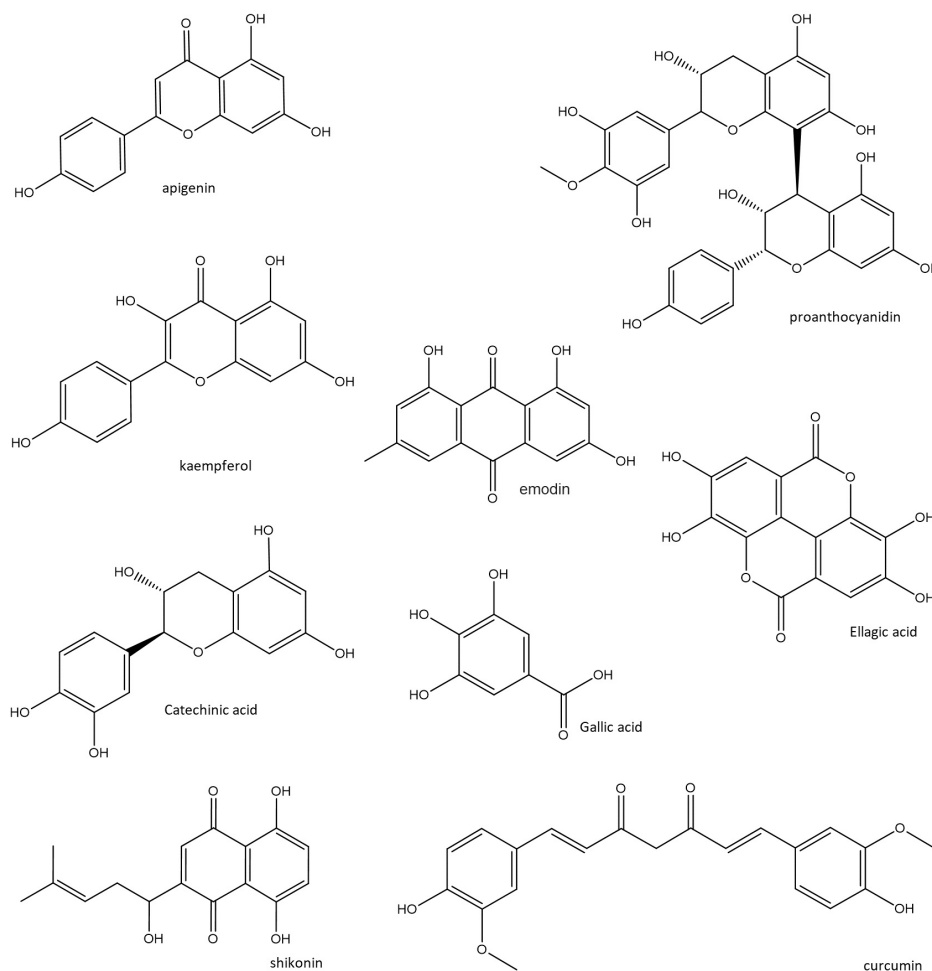
*N,N,N',N'*-tetrakis-[2-pyridylmethyl]-ethylenediamine (TPEN) was reported to have a cancer-specific copper chelation mediated cytotoxicity (Fatfat *et al.*, 2014). Additionally, TPEN treatment resulted in the excessive generation of ROS via the formation of the TPEN-copper complex, leading to cytotoxicity in human colon cancer HCT116 cells. Evidently, elevated copper levels may be important in maintaining the proper level of ROS generation in cancer cells, whereas intracellular copper levels are crucial to maintaining the proper level of ROS generation in cancer cells (Gupte and Mumper, 2009). Furthermore, although cellular copper levels may be a target for cancer treatment, it remains to be seen whether a copper-chelating drug can help overcome 5-FU resistance.

### Phenolic compounds

Interestingly, several antioxidant compounds promote the

production of ROS in cancer cells. Although more research is needed to elucidate the precise mechanisms, these antioxidant compounds are thought to modulate ROS generation and increase the cytotoxicity of 5-FU. Phenolic compounds refer to diverse natural products such as flavanols, flavonols, chalcones, tannins, curcuminoids, etc. Their antioxidant function is usually attributed to the phenolic ring structure (Cai *et al.*, 2006). The following sections list several phenolic compounds (Fig. 2) that have been reported to have synergistic cytotoxicity when combined with 5-FU or to be cytotoxic to 5-FU resistant cancer cells.

Apigenin is a flavonoid compound found in common fruits and vegetables that exhibits anti-inflammatory, antioxidant, and anticancer activity (Shukla and Gupta, 2010). Research revealed that apigenin cotreatment with 5-FU at a subtoxic level demonstrated synergistic cytotoxicity in treating hepatocellular carcinoma (HCC) cells *in vitro* and *in vivo* (Hu *et al.*, 2015). Moreover, the ROS level was increased, and the mitochondrial apoptotic pathway was activated, indicating that apigenin has a pro-oxidant function. Although it remains to be seen whether apigenin is cytotoxic to 5-FU resistant cancer cells, apigenin, which is well-known for its antioxidant activity, appears to also demonstrate some pro-oxidant activity.



**Fig. 2.** Phenolic compounds.

The polyphenolic extract from quince (*Cydonia oblonga* Miller) has shown antiproliferative effects in kidney and colon cancer cells (Carvalho *et al.*, 2010). A Tunisian research group reported that quince peel polyphenolic extract induced ROS production, and the cytotoxic effect of 5-FU was increased in human colon adenocarcinoma LS174 cell (Riahi-Chebba *et al.*, 2015). Although the potential expansion of the cellular work to a preclinical level requires further study, it is worth noting that ROS generation may be linked to the cytotoxicity of 5-FU. Riahi-Chebba *et al.* (2019), conversely, reported that kaempferol, another phenolic compound derived from quince, inhibited the production of ROS while exhibiting the same cytotoxicity as other phenolic compounds and was effective even in 5-FU resistant colon cancer cells. This intriguing result cautions us not to assume that a decrease in ROS levels is cytoprotective, as other mechanisms may simultaneously be responsible for cytotoxicity.

Shikonin, a naphthoquinone derivative found in the shikonin plant (*Lithospermum erythrorhizon*), is known for its cytotoxicity and anti-inflammatory activity (Chen *et al.*, 2002). Similarly, Liang *et al.* (2016) studied the antitumor activity of shikonin on gastric cancer. They observed that shikonin induced ROS generation and enhanced the 5-FU sensitivity *in vitro* and *in vivo*. In addition to the mitochondria-mediated apoptotic pathway, they detected the caspase-independent nuclear translocation of the apoptosis-inducing factor and endonuclease G from mitochondria.

Proanthocyanidin compounds from white fig *Ficus virens* (Chen *et al.*, 2017b) and *Uncaria rhynchophylla* (Chen *et al.*, 2017c) have been shown to have cytotoxic activity on human breast cancer MDA-MB-231 cells. Proanthocyanidins increased cellular ROS and the mitochondrial apoptotic pathway, and synergistic cytotoxicity was observed when proanthocyanidins were combined with 5-FU. Surprisingly, the cytotoxic effect appeared to be cancer cell-specific, and proanthocyanidins alleviated intestinal mucositis in 5-FU-treated rats (Chen *et al.*, 2017b).

Curcumin, a polyphenolic compound frequently found in curry powders, has long been considered an antioxidant (Ak and Gulcin, 2008). Several studies, however, have reported the generation of ROS by curcumin analogs. Researchers created B63, a curcumin analog, as an anticancer agent and discovered that B63 induced ROS-mediated paraptosis in gastric cancer cells (Chen *et al.*, 2019). They showed the inhibition of thioredoxin reductase 1 (TrxR1) by B63 *in vitro*, and the overexpression of TrxR1 negated the proapoptotic activity of B63. Their findings indicate that TrxR1 is a target of B63 and that B63 effectively suppressed the growth of 5-FU-resistant gastric cancer cells. Similarly, dimethoxycurcumin increases ROS production in colon cancer cells, allowing it to exert cytotoxic activity against colon cancer SW480 and SW620 cells when combined with 5-FU (Zhao *et al.*, 2017).

A Chinese research group studied the water extract of *Sanguisorba officinalis* L. radix, for its anticancer activity on human colorectal cancer HCT116 and RKO cells (Liu *et al.*, 2016a). They demonstrated that treating cells with the extract and 5-FU significantly increased ROS generation and that cotreatment increased 5-FU cytotoxicity. Moreover, they reported an increase in autophagy-related markers, light chain LC3, and p62, besides ROS generation, implying that the generation of ROS is not the only explanation for the synergism between *Sanguisorba officinalis* L. radix and 5-FU. The study

demonstrated that gallic acid, catechinic acid, and ellagic acid, three main constituents of *Sanguisorba officinalis* L. radix, are responsible for the synergistic activity.

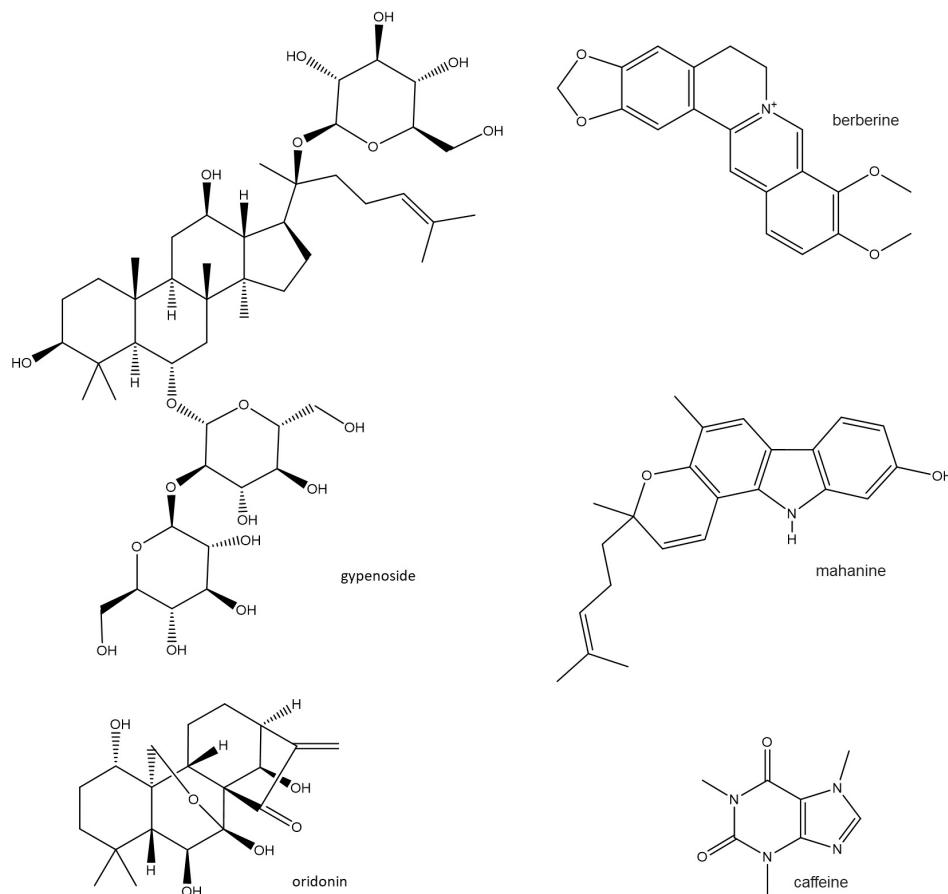
Manuka honey, a type of honey collected from the manuka tree *Leptospermum scoparium*, has antioxidant, anti-inflammatory, and anticancer properties (Afrin *et al.*, 2018b). Reports describe the synergistic cytotoxicity of manuka honey on human colon cancer HCT116 and LoVo cells when combined with 5-FU (Afrin *et al.*, 2018a). Manuka honey, a polyphenol-rich natural product, suppressed cell survival signals in HCT116 and LoVo cells while inducing pro-apoptotic signals and ROS production. Furthermore, the combined treatment reduced the activity of antioxidant enzymes such as SOD, catalase, glutathione peroxidase, glutathione reductase, and the expression of Nrf2, SOD, catalase, and HO-1, resulting in increased cell death due to oxidative stress.

Emodin, a natural anthraquinone compound, has antiproliferative activity in human breast cancer MCF7 cells (Huang *et al.*, 2007). In a later study, tests were conducted to determine whether low-dose emodin could potentiate the activity of 5-FU in MCF7 cells (Zu *et al.*, 2018). Findings revealed that emodin increased 5-FU-induced apoptosis in breast cancer cells by generating ROS. Surprisingly, researchers observed cellular senescence after 5-FU treatment with emodin, which they believe was caused by the upregulation of cyclin-dependent kinase inhibitors and the downregulation of E2F1 and the notch-regulated ankyrin repeat protein (NRARP) protein. Their findings suggested that NRARP is a critical target for inducing cellular senescence.

#### Polycyclic compounds and alkaloids

Several polycyclic compounds and alkaloids (Fig. 3) have been investigated for their role in producing ROS in cancer cells. For instance, gypenosides are triterpenoid saponin compounds whose potential use in cancer treatment has been documented (Ahmad *et al.*, 2019), and they are thought to have potentiated 5-FU's cytotoxicity (Kong *et al.*, 2015). Results showed that p53 and ROS generation mediates the synergism between gypenosides and 5-FU to exert anticancer activity. Additionally, the triterpenoid saponin compound, tubeimoside-I, isolated from *Rhizoma Bolbostemmatidis*, has exhibited antitumor activity in various types of tumors (Yu *et al.*, 1994). Yan *et al.* (2019) discovered that combining 5-FU and tubeimoside-I suppressed the growth of colorectal cancer SW480, SW620, HCT116, and RKO cells in a synergistic manner, whereas tubeimoside-I induced cellular ROS and the activation of AMPK, resulting in cytotoxic autophagy.

Oridonin, a diterpenoid from the medicinal herb *Rabdosia rubescens*, exhibits antitumor activity (Li *et al.*, 2011). Studies assessed oridonin's anticancer effect in colorectal cancer HCT15 cells and compared the 5-FU resistant HCT15 cells and sensitive cells (Zhang *et al.*, 2019). To exert its cytotoxicity, oridonin induced the generation of ROS in both cells and the activation of the JNK/c-Jun pathway. Notably, cotreatment with N-acetylcysteine reversed JNK/c-Jun pathway activation, indicating that ROS generation mediates JNK/c-Jun pathway activation. Although oridonin activated apoptosis in colorectal cancer cells, it appears to activate necroptosis in renal carcinoma 786-O cells (Zheng *et al.*, 2018). Cotreatment of oridonin and 5-FU showed synergistic cytotoxicity, probably through separate mechanisms, and notably, the same compound showed a different mechanism of action.



**Fig. 3.** Polycyclic compounds and alkaloids.

The anticancer effects of the *Coptis* herb extracts and the major alkaloid component berberine have been well-reported, and their cytotoxic effects have been detected in various cancer cell lines (Tang *et al.*, 2009). Furthermore, *Coptis* extract showed cytotoxicity when combined with 5-FU in human lung cancer A549 cells (He *et al.*, 2012). The cytotoxicity of either *Coptis* extract or berberine was associated with an increase in ROS generation in a dose-dependent manner, and when combined with 5-FU, the anticancer effect was enhanced.

Mahanine, an alkaloid from the curry leaf plant (*Murraya koenigii*), has exhibited various biological activities (Ramsewak *et al.*, 1999). Das *et al.* (2014) showed the synergistic enhancement of cytotoxicity of 5-FU when mahanine was used together in human colorectal cancer HCT116 and SW480 cells. Interestingly, the synergistic effect was observed irrespective of p53 status, i.e., both p53<sup>wt</sup> and p53<sup>null</sup> cells were sensitive to mahanine in combination with 5-FU. Although the precise mechanism is unknown, mahanine induced ROS production and led to the accumulation of PTEN and p53 in the nucleus. The increased production of ROS appears to be linked to the activation of tumor suppressor proteins PTEN and p53, resulting in increased cytotoxicity of 5-FU.

Caffeine, a food ingredient found in coffee and tea, slows the growth of liver cancer cells, including HepG2, HLF, Huh7, and PLC/PRF/5 (Okano *et al.*, 2008). Many studies report a synergistic effect of caffeine and cisplatin in various cancers,

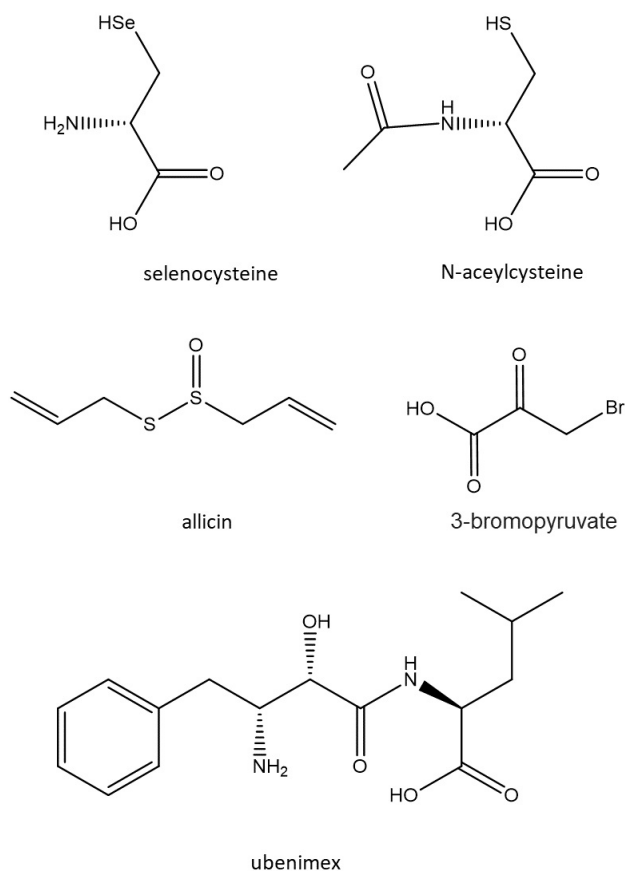
such as the human endometrial cancer cell line RL95-2 (Lin *et al.*, 2021). Recently, Wang *et al.* (2019) reported that the antitumor activity of 5-FU was enhanced by cotreatment of caffeine in HCC HepG3 and SMMMC cells. They discovered that combining 5-FU and caffeine inhibited HCC cell growth and induced apoptosis by increasing ROS production.

#### Role of other small molecules in ROS production

As described below, reports suggest that other small molecules (Fig. 4) may modulate ROS generation in cancer cells. First, selenocystine is the oxidation product of selenocysteine, which has a diselenide bond connecting two amino acids. It induces apoptosis in human cancer cells such as A375, HepG2, and MCF7 by increasing ROS production (Chen and Wong, 2009). Fan *et al.* (2013) investigated whether selenocystine cotreatment could increase the cytotoxicity of 5-FU in human melanoma A375 cells. They observed significant selenocystine-induced DNA damage mediated by ROS production and the inactivation of the extracellular-signal-regulated kinase (ERK) and Akt signaling pathways, resulting in anticancer synergism. Furthermore, the induction of ROS-mediated apoptosis in melanoma cells by 3,3'-diselenodipropionic acid, a selenocysteine derivative, is another example of potentially overcoming anticancer drug resistance (Cao *et al.*, 2014).

Alliin, a compound in garlic, has drawn considerable attention as an antimicrobial antioxidant (Chan *et al.*, 2013). Zou





**Fig. 4.** Other small molecules.

*et al.* (2016) tested whether the anticancer activity of 5-FU in human HCC SK-Hep-1 and BEL-7402 cells and in nude mice increased with allicin and 5-FU cotreatment. They discovered that cotreatment with allicin increased ROS production and sensitization of HCC cells to 5-FU. The synergistic effect was reversed by N-acetylcysteine treatment, indicating that the anticancer activity is mediated by ROS generation. Their study also demonstrated that cotreatment with allicin and 5-FU significantly inhibited the growth of HCC xenograft tumors in nude mice; although commonly thought to be an antioxidant, allicin increased ROS generation when combined with combined 5-FU.

3-Bromopyruvate is an inhibitor of hexokinase (Ko *et al.*, 2001), the key enzyme of glycolysis. The researchers reported that 3-bromopyruvate induced the ROS-mediated cell death of hepatoma SNU449 and Hep3B cells (Kim *et al.*, 2008). Upon treatment with 3-bromopyruvate, both cell lines underwent necrosis and apoptosis in an ATP depletion-dependent manner due to increased intracellular ROS and the disruption of mitochondrial function. Furthermore, the combination of 3-bromopyruvate and 5-FU inhibited tumor growth *in vivo* and *in vitro* (Chong *et al.*, 2017).

## ENDOGENOUS CELLULAR TARGETS TO OVERCOME 5-FU RESISTANCE

### Nuclear factor erythroid 2-related factor 2 (Nrf2)

The transcription factor Nrf2 mediates antioxidant response (Moi *et al.*, 1994). Nrf2 exists in the cytoplasm as the Nrf2-Keap1 complex in the absence of oxidative stress. The cellular level of Nrf2 is kept low by continuous degradation via the ubiquitin-proteasome system, which is mediated by Keap1, the Nrf2 key repressor (Zhang, 2006). Several cysteine residues of Keap1 are modified when exposed to oxidative stress, resulting in the dissociation of the Nrf2-Keap1 complex. Nrf2, which is released by Keap1, enters the nucleus and binds to the DNA in the antioxidant response element (ARE) region to regulate the expression of several genes involved in antioxidant function, such as glutamate-cysteine ligase catalytic subunit (Solis *et al.*, 2002), thioredoxin reductase (Soriano *et al.*, 2009), and heme oxygenase-1 (HO-1) (Jarmi and Agarwal, 2009). When expressed, these antioxidants may impart some degree of protection to cells under oxidative stress. Overexpression of Nrf2 in gastric cancer serves as a prognostic marker for 5-FU resistance, lending credence to Nrf2's prosurvival role (Hu *et al.*, 2013). Similarly, Nrf2 has a role in developing 5-FU resistance in colon cancer HT-29 cells (Akhdar *et al.*, 2009). Kang *et al.* (2014) discovered hypomethylation of Nrf2 promoter CpG islands in 5-FU resistance colorectal cancer SNU5/5-FUR cells compared with nonresistant cancer cells, indicating that Nrf2 upregulation led to 5-FU resistance.

Besides its antioxidant function, Nrf2 regulates the expression of drug-metabolizing enzymes and drug transporters, resulting in a decrease in 5-FU efficacy (Bai *et al.*, 2016). A team of researchers reported that 2',4'-dihydroxy-6'methoxy-3',5'-dimethylchalcone, an inhibitor of Nrf2/ARE pathway, could reverse 5-FU resistance in HCC BEL-7402 cells by inhibiting the 5-FU efflux (Wei *et al.*, 2018).

### ROS/mitogen-activated protein kinases pathway

JNK, c-Jun N-terminal kinase, belongs to MAPKs. The function of JNK is related to both cell survival (Wu *et al.*, 2019) and death (Dhanasekaran and Reddy, 2008). Based on the stimuli, JNK signaling can be either prosurvival or pro-apoptotic, and the signaling pathway is not directly linked to the cytotoxic effect of 5-FU. It appears that either activation or inactivation of the proper signaling pathway could place an additional burden on cells treated with 5-FU, potentially increasing 5-FU cytotoxicity.

Compared with differentiated and chemosensitive pancreatic cancer stem cells, the JNK signaling pathway is activated in pancreatic cancer stem cells (Okada *et al.*, 2014; Suzuki *et al.*, 2015). Researchers established that the JNK signaling pathway is activated in the pancreatic cancer stem cells (Suzuki *et al.*, 2015). Pretreatment of cells with SP600125, a JNK inhibitor, resulted in the sensitization of the cells to 5-FU and gemcitabine. The cytotoxic effects of these chemotherapeutics were accompanied by an increase in ROS production. Furthermore, the use of N-acetylcysteine, a free radical scavenger, reduced the intracellular level of ROS and allowed the cells to remain resistant to 5-FU; this is an example of the detrimental use of an antioxidant in chemotherapy. The synergistic cytotoxicity of 5-FU and the compounds mentioned above, tetrathiomolybdate (Kim *et al.*, 2012) and oridonin (Zhang *et al.*, 2019), is associated with the generation of ROS and the

activation of JNK. 5-FU cytotoxicity appears to be enhanced by oxidative stress and JNK activation, potentially overcoming 5-FU resistance.

Similarly, coronarin D, a diterpene compound derived from grapes, has anticancer activity. *Zingiberaceae* (Bailey, 2020) is involved with the activation of JNK signaling and ROS generation, as shown in human nasopharyngeal cancer cells (Chen *et al.*, 2017a). It was recently reported that coronarin D induces the apoptosis of 5-FU resistant human oral cancer cells. The cytotoxicity is related to the JNK signaling pathway (Hsieh *et al.*, 2020).

Besides JNK signaling, activation of p38 MAP kinase is linked to 5-FU cytotoxicity. According to Xie *et al.* (2016), the overexpression of nicotinamide N-methyltransferase causes a decrease in ROS levels, and the inactivation of p38 signaling is involved in 5-FU resistance in colorectal cancer SW480 cells. Moon *et al.* (2020) reported that the activation of p38 by yeast extract resulted in the antitumor effect on 5-FU resistant colorectal cancer SNU-C5 cells.

Nevertheless, the role of the ERK pathway in 5-FU sensitivity appears to be pro-survival. Kim *et al.* (2016) discovered ERK overexpression in SUNC5/FUR cells, resistant to 5-FU. Furthermore, sensitization was achieved by transfecting 5-FU resistant cells with siRNA against ERK. The role of aluminum chloride in inducing 5-FU resistance was further investigated, revealing that ERK activation facilitated the survival of HCC HepG2 cells during 5-FU treatment (Li *et al.*, 2019a). By contrast, U0126, an ERK inhibitor, reversed aluminum chloride-induced 5-FU resistance; it also remains unclear whether ERK activation is required for cells to remain resistant to 5-FU. A Japanese group, for example, reported a 5-FU resistant human squamous carcinoma UM-SCC-23 cell line that activated both ERK and Akt signals (You *et al.*, 2009). Nevertheless, U0126 could not reverse the resistance, whereas Akt inhibition was. Furthermore, Wang *et al.* (2017b) reported that 5-FU resistance caused by ADAM12 overexpression increased phosphorylated Akt but not phosphorylated ERK in breast cancer SKBR3 MDA-MB-231 cells.

#### PI3K/Akt pathway

Phosphatidylinositol 3-kinase (PI3K)/Akt pathway is considered one of the key signaling pathways that confers cancer cells' resistance to chemotherapy (Liu *et al.*, 2020). Researchers reported the constitutive activation of Akt signaling in 5-FU resistant squamous carcinoma UM-SCC-23 cells (You *et al.*, 2009). By contrast, the inhibition of Akt signaling was noticed when the synergistic cytotoxicity of 5-FU was observed by the cotreatment of several compounds, including violacein (Kodach *et al.*, 2006), curcumin (Zhang *et al.*, 2017), and kaempferol (Li *et al.*, 2019b). Recent reports state that celecoxib, a COX-2 inhibitor, induced apoptosis by inhibiting Akt in 5-FU resistant gastric carcinoma AGS cells (Choi *et al.*, 2021). The inhibition of COX-2 is thought to have resulted in the down-regulation of Akt and the induction of apoptosis. MK-2206's direct inhibition of Akt also increases the cytotoxicity of 5-FU in gastric cancer SGC-7901 and MKN45 cells (Jin *et al.*, 2016). Hence, it remains to be confirmed whether MK-2206 could be used to treat 5-FU resistant cells.

#### Autophagy pathway

Autophagy, a self-eating process involving the autophagosome (Glick *et al.*, 2010), has two states, either pro-survival or

pro-death, depending on the mode of activation and which cells are affected. Initially, the pro-survival role of autophagy was reported because inhibition of autophagy was associated with increased cytotoxicity of 5-FU in human colon cancer colon26 and HT29 cells *in vitro* and *in vivo* (Li *et al.*, 2010). The pro-survival role of autophagy was again shown in HCT116 p53<sup>-/-</sup> cells (Sui *et al.*, 2014) and human HCC Bel-7402 cells (Wang *et al.*, 2017a). The involvement of pro-survival autophagy in 5-FU resistance has been demonstrated for several cellular components such as TSPAN9 (Qi *et al.*, 2020) and claudin-1 (Tong *et al.*, 2019). Zhang *et al.* (2017) reported synergistic cytotoxicity from the combination of curcumin and 5-FU, which included a reduction in pro-survival autophagy mediated by AMPK and Unc-51 Like Autophagy Activating Kinase 1 (ULK1). By contrast, several compounds have been shown to increase the cytotoxicity of 5-FU by inducing pro-death autophagy.  $\beta$ -Elemene, a sesquiterpene compound found in various plants, induces pro-death autophagy in 5-FU resistant colorectal cancer HCT116 p53<sup>-/-</sup> cells (Zhang *et al.*, 2020). Similarly, when combined with 5-FU, withaferin-A, a natural product with a steroidal lactone structure, induces endoplasmic reticulum stress-mediated autophagy (Alnuqaydan *et al.*, 2020) in CRC cells (SW480, HT29, HCT116).

#### Aminopeptidase N (CD13)

Aminopeptidase N, also known as CD13, is a cell-surface-anchored zinc peptidase with various functions, including peptide cleavage, endocytosis, and signaling (Mina-Osorio, 2008). It was initially identified as a cell surface marker CD13 for myeloid leukemia cells (Sakai *et al.*, 1987), and the signaling function of aminopeptidase N appears to be independent of enzyme activity. Aminopeptidase expression has been linked to a poor prognosis and angiogenesis in cancer cells such as nonsmall cell lung cancer (Tokuhara *et al.*, 2006), pancreatic carcinoma (Ikeda *et al.*, 2003), and colon cancer (Hashida *et al.*, 2002). When substrates or inhibitors bind to CD13, the conformation of the dimeric CD13 structure changes, resulting in signal transduction (Xu *et al.*, 1997), and CD13 protects cells from apoptosis by reducing ROS-induced DNA damage.

Ubenimex, also known as bestatin, is a dipeptide compound produced by actinomycetes (Umezawa *et al.*, 1976). It specifically blocks and antagonizes CD13 (Look *et al.*, 1989). Haraguchi *et al.* (2010) reported that ubenimex or a CD13-neutralizing antibody inhibited CD13 in hepatocellular cancer HuH7 cells. They revealed that combining 5-FU and ubenimex increased ROS production and improved liver cancer therapy. Additionally, Dou *et al.* (2017) investigated the therapeutic potential of BC-02, a conjugation compound of ubenimex and 5-FU, in the successful inhibition of the growth and self-renewal of liver cancer stem cells. Similarly, Sun *et al.* (2015) reported that 4cc, a synthetic inhibitor of aminopeptidase N, could increase 5-FU cytotoxicity by generating ROS in human liver cancer HCC cells. It remains to be seen whether CD13 inhibitors can be used to treat cancer cells other than HCC.

## CONCLUSION

In this study, we have compiled a list of compounds that can be used alone or combined with 5-FU to modulate ROS generation. Although the precise mechanisms underlying their cytotoxicity remain unknown, several endogenous cellular tar-

gets have been identified, as described above. These compounds and cellular targets could help develop new strategies for combating 5-FU resistance.

## CONFLICT OF INTEREST

The authors claim no conflicts of interest.

## ACKNOWLEDGMENTS

This work was supported by research grants from Daegu Catholic University in 2020.

## REFERENCES

Afrin, S., Giampieri, F., Forbes-Hernandez, T. Y., Gasparri, M., Amici, A., Cianciosi, D., Quiles, J. L. and Battino, M. (2018a) Manuka honey synergistically enhances the chemopreventive effect of 5-fluorouracil on human colon cancer cells by inducing oxidative stress and apoptosis, altering metabolic phenotypes and suppressing metastasis ability. *Free Radic. Biol. Med.* **126**, 41-54.

Afrin, S., Giampieri, F., Gasparri, M., Forbes-Hernandez, T. Y., Cianciosi, D., Reboredo-Rodriguez, P., Amici, A., Quiles, J. L. and Battino, M. (2018b) The inhibitory effect of Manuka honey on human colon cancer HCT-116 and LoVo cell growth. Part 1: the suppression of cell proliferation, promotion of apoptosis and arrest of the cell cycle. *Food Funct.* **9**, 2145-2157.

Ahmad, B., Khan, S., Nabi, G., Gamallat, Y., Su, P., Jamal, Y., Duan, P. and Yao, L. (2019) Natural gypenosides: targeting cancer through different molecular pathways. *Cancer Manag. Res.* **11**, 2287-2297.

Ak, T. and Gulcin, I. (2008) Antioxidant and radical scavenging properties of curcumin. *Chem. Biol. Interact.* **174**, 27-37.

Akhdar, H., Loyer, P., Rauch, C., Corlu, A., Guillouzo, A. and Morel, F. (2009) Involvement of Nrf2 activation in resistance to 5-fluorouracil in human colon cancer HT-29 cells. *Eur. J. Cancer* **45**, 2219-2227.

Alnuqaydan, A. M., Rah, B., Almutary, A. G. and Chauhan, S. S. (2020) Synergistic antitumor effect of 5-fluorouracil and withaferin-A induces endoplasmic reticulum stress-mediated autophagy and apoptosis in colorectal cancer cells. *Am. J. Cancer Res.* **10**, 799-815.

Bai, X., Chen, Y., Hou, X., Huang, M. and Jin, J. (2016) Emerging role of NRF2 in chemoresistance by regulating drug-metabolizing enzymes and efflux transporters. *Drug Metab. Rev.* **48**, 541-567.

Bailly, C. (2020) Anticancer activities and mechanism of action of the labdane diterpene coronarin D. *Pathol. Res. Pract.* **216**, 152946.

Blondy, S., David, V., Verdier, M., Mathonnet, M., Perraud, A. and Christou, N. (2020) 5-Fluorouracil resistance mechanisms in colorectal cancer: from classical pathways to promising processes. *Cancer Sci.* **111**, 3142-3154.

Brewer, G. J., Dick, R. D., Yuzbasiyan-Gurkin, V., Tankanow, R., Young, A. B. and Kluin, K. J. (1991) Initial therapy of patients with Wilson's disease with tetrathiomolybdate. *Arch. Neurol.* **48**, 42-47.

Cai, Y. Z., Mei, S., Jie, X., Luo, Q. and Corke, H. (2006) Structure-radical scavenging activity relationships of phenolic compounds from traditional Chinese medicinal plants. *Life Sci.* **78**, 2872-2888.

Cao, W., Li, X., Zheng, S., Zheng, W., Wong, Y. S. and Chen, T. (2014) Selenocysteine derivative overcomes TRAIL resistance in melanoma cells: evidence for ROS-dependent synergism and signaling crosstalk. *Oncotarget* **5**, 7431-7445.

Carvalho, M., Silva, B. M., Silva, R., Valentao, P., Andrade, P. B. and Bastos, M. L. (2010) First report on *Cydonia oblonga* Miller anticancer potential: differential antiproliferative effect against human kidney and colon cancer cells. *J. Agric. Food Chem.* **58**, 3366-3370.

Chan, J. Y., Yuen, A. C., Chan, R. Y. and Chan, S. W. (2013) A review of the cardiovascular benefits and antioxidant properties of allicin. *Phytother. Res.* **27**, 637-646.

Chen, J. C., Hsieh, M. C., Lin, S. H., Lin, C. C., Hsi, Y. T., Lo, Y. S., Chuang, Y. C., Hsieh, M. J. and Chen, M. K. (2017a) Coronarin D induces reactive oxygen species-mediated cell death in human nasopharyngeal cancer cells through inhibition of p38 MAPK and activation of JNK. *Oncotarget* **8**, 108006-108019.

Chen, T. and Wong, Y. S. (2009) Selenocysteine induces reactive oxygen species-mediated apoptosis in human cancer cells. *Biomed. Pharmacother.* **63**, 105-113.

Chen, X., Chen, X., Zhang, X., Wang, L., Cao, P., Rajamanickam, V., Wu, C., Zhou, H., Cai, Y., Liang, G. and Wang, Y. (2019) Curcuminoid B63 induces ROS-mediated paraptosis-like cell death by targeting TrxR1 in gastric cells. *Redox. Biol.* **21**, 101061.

Chen, X., Yang, L., Oppenheim, J. J. and Howard, M. Z. (2002) Cellular pharmacology studies of shikonin derivatives. *Phytother. Res.* **16**, 199-209.

Chen, X. X., Lam, K. H., Chen, Q. X., Leung, G. P., Tang, S. C. W., Sze, S. C., Xiao, J. B., Feng, F., Wang, Y., Zhang, K. Y. and Zhang, Z. J. (2017b) Ficus virens proanthocyanidins induced apoptosis in breast cancer cells concomitantly ameliorated 5-fluorouracil induced intestinal mucositis in rats. *Food Chem. Toxicol.* **110**, 49-61.

Chen, X. X., Leung, G. P., Zhang, Z. J., Xiao, J. B., Lao, L. X., Feng, F., Mak, J. C., Wang, Y., Sze, S. C. and Zhang, K. Y. (2017c) Proanthocyanidins from *Uncaria rhynchophylla* induced apoptosis in MDA-MB-231 breast cancer cells while enhancing cytotoxic effects of 5-fluorouracil. *Food Chem. Toxicol.* **107**, 248-260.

Choi, S. M., Cho, Y. S., Park, G., Lee, S. K. and Chun, K. S. (2021) Celecoxib induces apoptosis through Akt inhibition in 5-fluorouracil-resistant gastric cancer cells. *Toxicol. Res.* **37**, 25-33.

Chong, D., Ma, L., Liu, F., Zhang, Z., Zhao, S., Huo, Q., Zhang, P., Zheng, H. and Liu, H. (2017) Synergistic antitumor effect of 3-bromopyruvate and 5-fluorouracil against human colorectal cancer through cell cycle arrest and induction of apoptosis. *Anticancer Drugs* **28**, 831-840.

Das, R., Bhattacharya, K., Sarkar, S., Samanta, S. K., Pal, B. C. and Mandal, C. (2014) Mahanine synergistically enhances cytotoxicity of 5-fluorouracil through ROS-mediated activation of PTEN and p53/p73 in colon carcinoma. *Apoptosis* **19**, 149-164.

Dhanasekaran, D. N. and Reddy, E. P. (2008) JNK signaling in apoptosis. *Oncogene* **27**, 6245-6251.

Dou, C., Fang, C., Zhao, Y., Fu, X., Zhang, Y., Zhu, D., Wu, H., Liu, H., Zhang, J., Xu, W., Liu, Z., Wang, H., Li, D. and Wang, X. (2017) BC-02 eradicates liver cancer stem cells by upregulating the ROS-dependent DNA damage. *Int. J. Oncol.* **51**, 1775-1784.

Fan, C., Chen, J., Wang, Y., Wong, Y. S., Zhang, Y., Zheng, W., Cao, W. and Chen, T. (2013) Selenocysteine potentiates cancer cell apoptosis induced by 5-fluorouracil by triggering reactive oxygen species-mediated DNA damage and inactivation of the ERK pathway. *Free Radic. Biol. Med.* **65**, 305-316.

Fatfat, M., Merhi, R. A., Rahal, O., Stoyanovsky, D. A., Zaki, A., Haidar, H., Kagan, V. E., Gali-Muhtasib, H. and Machaca, K. (2014) Copper chelation selectively kills colon cancer cells through redox cycling and generation of reactive oxygen species. *BMC Cancer* **14**, 527.

Frei, E., 3rd, Elias, A., Wheeler, C., Richardson, P. and Hryniuk, W. (1998) The relationship between high-dose treatment and combination chemotherapy: the concept of summation dose intensity. *Clin. Cancer Res.* **4**, 2027-2037.

Glick, D., Barth, S. and Macleod, K. F. (2010) Autophagy: cellular and molecular mechanisms. *J. Pathol.* **221**, 3-12.

Gorrini, C., Harris, I. S. and Mak, T. W. (2013) Modulation of oxidative stress as an anticancer strategy. *Nat. Rev. Drug Discov.* **12**, 931-947.

Gupte, A. and Mumper, R. J. (2009) Elevated copper and oxidative stress in cancer cells as a target for cancer treatment. *Cancer Treat. Rev.* **35**, 32-46.

Haraguchi, N., Ishii, H., Mimori, K., Tanaka, F., Ohkuma, M., Kim, H. M., Akita, H., Takiuchi, D., Hatano, H., Nagano, H., Barnard, G. F., Doki, Y. and Mori, M. (2010) CD13 is a therapeutic target in human liver cancer stem cells. *J. Clin. Invest.* **120**, 3326-3339.

Hashida, H., Takabayashi, A., Kanai, M., Adachi, M., Kondo, K., Kohno, N., Yamaoka, Y. and Miyake, M. (2002) Aminopeptidase N is involved in cell motility and angiogenesis: its clinical significance in



- human colon cancer. *Gastroenterology* **122**, 376-386.
- He, C., Rong, R., Liu, J., Wan, J., Zhou, K. and Kang, J. X. (2012) Effects of Coptis extract combined with chemotherapeutic agents on ROS production, multidrug resistance, and cell growth in A549 human lung cancer cells. *Chin. Med.* **7**, 11.
- Hsieh, M. Y., Hsieh, M. J., Lo, Y. S., Lin, C. C., Chuang, Y. C., Chen, M. K. and Chou, M. C. (2020) Modulating effect of Coronarin D in 5-fluorouracil resistance human oral cancer cell lines induced apoptosis and cell cycle arrest through JNK1/2 signaling pathway. *Biomed. Pharmacother.* **128**, 110318.
- Hu, X. F., Yao, J., Gao, S. G., Wang, X. S., Peng, X. Q., Yang, Y. T. and Feng, X. S. (2013) Nrf2 overexpression predicts prognosis and 5-FU resistance in gastric cancer. *Asian Pac. J. Cancer Prev.* **14**, 5231-5235.
- Hu, X. Y., Liang, J. Y., Guo, X. J., Liu, L. and Guo, Y. B. (2015) 5-Fluorouracil combined with apigenin enhances anticancer activity through mitochondrial membrane potential (DeltaPsi<sub>m</sub>)-mediated apoptosis in hepatocellular carcinoma. *Clin. Exp. Pharmacol. Physiol.* **42**, 146-153.
- Huang, Q., Lu, G., Shen, H. M., Chung, M. C. and Ong, C. N. (2007) Anti-cancer properties of anthraquinones from rhubarb. *Med. Res. Rev.* **27**, 609-630.
- Hwang, I. T., Chung, Y. M., Kim, J. J., Chung, J. S., Kim, B. S., Kim, H. J., Kim, J. S. and Yoo, Y. D. (2007) Drug resistance to 5-FU linked to reactive oxygen species modulator 1. *Biochem. Biophys. Res. Commun.* **359**, 304-310.
- Ikedo, N., Nakajima, Y., Tokuhara, T., Hattori, N., Sho, M., Kanehiro, H. and Miyake, M. (2003) Clinical significance of aminopeptidase N/CD13 expression in human pancreatic carcinoma. *Clin. Cancer Res.* **9**, 1503-1508.
- Jarmi, T. and Agarwal, A. (2009) Heme oxygenase and renal disease. *Curr. Hypertens. Rep.* **11**, 56-62.
- Jin, P., Wong, C. C., Mei, S., He, X., Qian, Y. and Sun, L. (2016) MK-2206 co-treatment with 5-fluorouracil or doxorubicin enhances chemosensitivity and apoptosis in gastric cancer by attenuation of Akt phosphorylation. *Oncotargets Ther.* **9**, 4387-4396.
- Juhász, A., Markel, S., Gaur, S., Liu, H., Lu, J., Jiang, G., Wu, X., Antony, S., Wu, Y., Melillo, G., Meitzler, J. L., Haines, D. C., Butcher, D., Roy, K. and Doroshov, J. H. (2017) NADPH oxidase 1 supports proliferation of colon cancer cells by modulating reactive oxygen species-dependent signal transduction. *J. Biol. Chem.* **292**, 7866-7887.
- Kang, K. A., Piao, M. J., Kim, K. C., Kang, H. K., Chang, W. Y., Park, I. C., Keum, Y. S., Surh, Y. J. and Hyun, J. W. (2014) Epigenetic modification of Nrf2 in 5-fluorouracil-resistant colon cancer cells: involvement of TET-dependent DNA demethylation. *Cell Death Dis.* **5**, e1183.
- Kim, J. K., Kang, K. A., Piao, M. J., Ryu, Y. S., Han, X., Fernando, P. M., Oh, M. C., Park, J. E., Shilnikova, K., Boo, S. J., Na, S. Y., Jeong, Y. J., Jeong, S. U. and Hyun, J. W. (2016) Endoplasmic reticulum stress induces 5-fluorouracil resistance in human colon cancer cells. *Environ. Toxicol. Pharmacol.* **44**, 128-133.
- Kim, J. S., Ahn, K. J., Kim, J. A., Kim, H. M., Lee, J. D., Lee, J. M., Kim, S. J. and Park, J. H. (2008) Role of reactive oxygen species-mediated mitochondrial dysregulation in 3-bromopyruvate induced cell death in hepatoma cells: ROS-mediated cell death by 3-BrPA. *J. Bioenerg. Biomembr.* **40**, 607-618.
- Kim, K. K., Kwar, N. M., Singh, R. K., Lange, T. S., Brard, L. and Moore, R. G. (2011) Tetrathiomolybdate induces doxorubicin sensitivity in resistant tumor cell lines. *Gynecol. Oncol.* **122**, 183-189.
- Kim, K. K., Lange, T. S., Singh, R. K., Brard, L. and Moore, R. G. (2012) Tetrathiomolybdate sensitizes ovarian cancer cells to anticancer drugs doxorubicin, fenretinide, 5-fluorouracil and mitomycin C. *BMC Cancer* **12**, 147.
- Ko, Y. H., Pedersen, P. L. and Geschwind, J. F. (2001) Glucose catabolism in the rabbit VX2 tumor model for liver cancer: characterization and targeting hexokinase. *Cancer Lett.* **173**, 83-91.
- Kodach, L. L., Bos, C. L., Duran, N., Peppelenbosch, M. P., Ferreira, C. V. and Hardwick, J. C. (2006) Violacein synergistically increases 5-fluorouracil cytotoxicity, induces apoptosis and inhibits Akt-mediated signal transduction in human colorectal cancer cells. *Carcinogenesis* **27**, 508-516.
- Kong, L., Wang, X., Zhang, K., Yuan, W., Yang, Q., Fan, J., Wang, P. and Liu, Q. (2015) Gypenosides synergistically enhances the anti-tumor effect of 5-fluorouracil on colorectal cancer *in vitro* and *in vivo*: a role for oxidative stress-mediated DNA damage and p53 activation. *PLoS ONE* **10**, e0137888.
- Kumar, B., Koul, S., Khandrika, L., Meacham, R. B. and Koul, H. K. (2008) Oxidative stress is inherent in prostate cancer cells and is required for aggressive phenotype. *Cancer Res.* **68**, 1777-1785.
- Li, C. Y., Wang, E. Q., Cheng, Y. and Bao, J. K. (2011) Oridonin: an active diterpenoid targeting cell cycle arrest, apoptotic and autophagic pathways for cancer therapeutics. *Int. J. Biochem. Cell Biol.* **43**, 701-704.
- Li, J., Hou, N., Faried, A., Tsutsumi, S. and Kuwano, H. (2010) Inhibition of autophagy augments 5-fluorouracil chemotherapy in human colon cancer *in vitro* and *in vivo* model. *Eur. J. Cancer* **46**, 1900-1909.
- Li, M., Cui, Z. G., Zakki, S. A., Feng, Q., Sun, L., Feril, L. B., Jr. and Inadera, H. (2019a) Aluminum chloride causes 5-fluorouracil resistance in hepatocellular carcinoma HepG2 cells. *J. Cell. Physiol.* **234**, 20249-20265.
- Li, Q., Wei, L., Lin, S., Chen, Y., Lin, J. and Peng, J. (2019b) Synergistic effect of kaempferol and 5-fluorouracil on the growth of colorectal cancer cells by regulating the PI3K/Akt signaling pathway. *Mol. Med. Rep.* **20**, 728-734.
- Liang, W., Cai, A., Chen, G., Xi, H., Wu, X., Cui, J., Zhang, K., Zhao, X., Yu, J., Wei, B. and Chen, L. (2016) Shikonin induces mitochondria-mediated apoptosis and enhances chemotherapeutic sensitivity of gastric cancer through reactive oxygen species. *Sci. Rep.* **6**, 38267.
- Lin, C. K., Liu, S. T., Wu, Z. S., Wang, Y. C. and Huang, S. M. (2021) Mechanisms of cisplatin in combination with repurposed drugs against human endometrial carcinoma cells. *Life (Basel)* **11**, 160.
- Liou, G. Y. and Storz, P. (2010) Reactive oxygen species in cancer. *Free Radic. Res.* **44**, 479-496.
- Liu, M. P., Liao, M., Dai, C., Chen, J. F., Yang, C. J., Liu, M., Chen, Z. G. and Yao, M. C. (2016a) *Sanguisorba officinalis* L synergistically enhanced 5-fluorouracil cytotoxicity in colorectal cancer cells by promoting a reactive oxygen species-mediated, mitochondria-caspase-dependent apoptotic pathway. *Sci. Rep.* **6**, 34245.
- Liu, R., Chen, Y., Liu, G., Li, C., Song, Y., Cao, Z., Li, W., Hu, J., Lu, C. and Liu, Y. (2020) PI3K/AKT pathway as a key link modulates the multidrug resistance of cancers. *Cell Death Dis.* **11**, 797.
- Liu, Y., Li, Q., Zhou, L., Xie, N., Nice, E. C., Zhang, H., Huang, C. and Lei, Y. (2016b) Cancer drug resistance: redox resetting renders a way. *Oncotarget* **7**, 42740-42761.
- Longley, D. B., Harkin, D. P. and Johnston, P. G. (2003) 5-Fluorouracil: mechanisms of action and clinical strategies. *Nat. Rev. Cancer* **3**, 330-338.
- Look, A. T., Ashmun, R. A., Shapiro, L. H. and Peiper, S. C. (1989) Human myeloid plasma membrane glycoprotein CD13 (gp150) is identical to aminopeptidase N. *J. Clin. Invest.* **83**, 1299-1307.
- Mates, J. M. and Sanchez-Jimenez, F. M. (2000) Role of reactive oxygen species in apoptosis: implications for cancer therapy. *Int. J. Biochem. Cell Biol.* **32**, 157-170.
- Mehrzad, V., Roayaei, M., Peikar, M. S., Nouranian, E., Mokarian, F., Khani, M. and Farzannia, S. (2016) Bevacizumab plus FOLFOX or FOLFIRI regimens on patients with unresectable liver-only metastases of metastatic colorectal cancer. *Adv. Biomed. Res.* **5**, 10.
- Mina-Osorio, P. (2008) The moonlighting enzyme CD13: old and new functions to target. *Trends Mol. Med.* **14**, 361-371.
- Moi, P., Chan, K., Asunis, I., Cao, A. and Kan, Y. W. (1994) Isolation of NF-E2-related factor 2 (Nrf2), a NF-E2-like basic leucine zipper transcriptional activator that binds to the tandem NF-E2/AP1 repeat of the beta-globin locus control region. *Proc. Natl. Acad. Sci. U.S.A.* **91**, 9926-9930.
- Moon, D., Kang, H. K., Kim, J. and Yoon, S. P. (2020) Yeast extract induces apoptosis and cell cycle arrest via activating p38 signal pathway in colorectal cancer cells. *Ann. Clin. Lab. Sci.* **50**, 31-44.
- Nishikawa, M. (2008) Reactive oxygen species in tumor metastasis. *Cancer Lett.* **266**, 53-59.
- Okada, M., Shibuya, K., Sato, A., Seino, S., Suzuki, S., Seino, M. and Kitanaka, C. (2014) Targeting the K-Ras--JNK axis eliminates can-

- cer stem-like cells and prevents pancreatic tumor formation. *Oncotarget* **5**, 5100-5112.
- Okano, J., Nagahara, T., Matsumoto, K. and Murawaki, Y. (2008) Caffeine inhibits the proliferation of liver cancer cells and activates the MEK/ERK/EGFR signalling pathway. *Basic Clin. Pharmacol. Toxicol.* **102**, 543-551.
- Pazdur, R., Hoff, P. M., Medgyesy, D., Royce, M. and Brito, R. (1998) The oral fluorouracil prodrugs. *Oncology (Williston Park)* **12**, 48-51.
- Qi, Y., Qi, W., Liu, S., Sun, L., Ding, A., Yu, G., Li, H., Wang, Y., Qiu, W. and Lv, J. (2020) TSPAN9 suppresses the chemosensitivity of gastric cancer to 5-fluorouracil by promoting autophagy. *Cancer Cell Int.* **20**, 4.
- Ramsewak, R. S., Nair, M. G., Strasburg, G. M., DeWitt, D. L. and Nitiss, J. L. (1999) Biologically active carbazole alkaloids from *Murraya koenigii*. *J. Agric. Food Chem.* **47**, 444-447.
- Riahi-Chebbi, I., Haoues, M., Essafi, M., Zakraoui, O., Fattouch, S., Karoui, H. and Essafi-Benkhadir, K. (2015) Quince peel polyphenolic extract blocks human colon adenocarcinoma LS174 cell growth and potentiates 5-fluorouracil efficacy. *Cancer Cell Int.* **16**, 1.
- Riahi-Chebbi, I., Souid, S., Othman, H., Haoues, M., Karoui, H., Morel, A., Srairi-Abid, N., Essafi, M. and Essafi-Benkhadir, K. (2019) The Phenolic compound Kaempferol overcomes 5-fluorouracil resistance in human resistant LS174 colon cancer cells. *Sci. Rep.* **9**, 195.
- Sakai, K., Hattori, T., Sagawa, K., Yokoyama, M. and Takatsuki, K. (1987) Biochemical and functional characterization of MCS-2 antigen (CD13) on myeloid leukemic cells and polymorphonuclear leukocytes. *Cancer Res.* **47**, 5572-5576.
- Shukla, S. and Gupta, S. (2010) Apigenin: a promising molecule for cancer prevention. *Pharm. Res.* **27**, 962-978.
- Solis, W. A., Dalton, T. P., Dieter, M. Z., Freshwater, S., Harrer, J. M., He, L., Shertzer, H. G. and Nebert, D. W. (2002) Glutamate-cysteine ligase modifier subunit: mouse *Gclm* gene structure and regulation by agents that cause oxidative stress. *Biochem. Pharmacol.* **63**, 1739-1754.
- Soriano, F. X., Baxter, P., Murray, L. M., Sporn, M. B., Gillingwater, T. H. and Hardingham, G. E. (2009) Transcriptional regulation of the AP-1 and Nrf2 target gene sulfiredoxin. *Mol. Cells* **27**, 279-282.
- Souglakos, J., Androulakis, N., Syrigos, K., Polyzos, A., Ziras, N., Athanasiadis, A., Kakolyris, S., Tsousis, S., Kouroussis, C., Vamvakas, L., Kalykaki, A., Samonis, G., Mavroudis, D. and Georgoulas, V. (2006) FOLFOXIRI (folinic acid, 5-fluorouracil, oxaliplatin and irinotecan) vs FOLFIRI (folinic acid, 5-fluorouracil and irinotecan) as first-line treatment in metastatic colorectal cancer (MCC): a multicentre randomised phase III trial from the hellenic oncology research group (HORG). *Br. J. Cancer* **94**, 798-805.
- Sui, X., Kong, N., Wang, X., Fang, Y., Hu, X., Xu, Y., Chen, W., Wang, K., Li, D., Jin, W., Lou, F., Zheng, Y., Hu, H., Gong, L., Zhou, X., Pan, H. and Han, W. (2014) JNK confers 5-fluorouracil resistance in p53-deficient and mutant p53-expressing colon cancer cells by inducing survival autophagy. *Sci. Rep.* **4**, 4694.
- Sun, Z. P., Zhang, J., Shi, L. H., Zhang, X. R., Duan, Y., Xu, W. F., Dai, G. and Wang, X. J. (2015) Aminopeptidase N inhibitor 4cc synergizes antitumor effects of 5-fluorouracil on human liver cancer cells through ROS-dependent CD13 inhibition. *Biomed. Pharmacother.* **76**, 65-72.
- Suzuki, S., Okada, M., Shibuya, K., Seino, M., Sato, A., Takeda, H., Seino, S., Yoshioka, T. and Kitanaka, C. (2015) JNK suppression of chemotherapeutic agents-induced ROS confers chemoresistance on pancreatic cancer stem cells. *Oncotarget* **6**, 458-470.
- Tang, J., Feng, Y., Tsao, S., Wang, N., Curtain, R. and Wang, Y. (2009) Berberine and Coptidis rhizoma as novel antineoplastic agents: a review of traditional use and biomedical investigations. *J. Ethnopharmacol.* **126**, 5-17.
- Tokuhara, T., Hattori, N., Ishida, H., Hirai, T., Higashiyama, M., Kodama, K. and Miyake, M. (2006) Clinical significance of aminopeptidase N in non-small cell lung cancer. *Clin. Cancer Res.* **12**, 3971-3978.
- Tong, H., Li, T., Qiu, W. and Zhu, Z. (2019) Claudin-1 silencing increases sensitivity of liver cancer HepG2 cells to 5-fluorouracil by inhibiting autophagy. *Oncol. Lett.* **18**, 5709-5716.
- Torres, M. and Forman, H. J. (2003) Redox signaling and the MAP kinase pathways. *BioFactors* **17**, 287-296.
- Umezawa, H., Aoyagi, T., Suda, H., Hamada, M. and Takeuchi, T. (1976) Bestatin, an inhibitor of aminopeptidase B, produced by actinomycetes. *J. Antibiot.* **29**, 97-99.
- Ushio-Fukai, M. and Nakamura, Y. (2008) Reactive oxygen species and angiogenesis: NADPH oxidase as target for cancer therapy. *Cancer Lett.* **266**, 37-52.
- Wang, M., Huang, C., Su, Y., Yang, C., Xia, Q. and Xu, D. J. (2017a) Astragaloside II sensitizes human hepatocellular carcinoma cells to 5-fluorouracil via suppression of autophagy. *J. Pharm. Pharmacol.* **69**, 743-752.
- Wang, X., Wang, Y., Gu, J., Zhou, D., He, Z., Wang, X. and Ferrone, S. (2017b) ADAM12-L confers acquired 5-fluorouracil resistance in breast cancer cells. *Sci. Rep.* **7**, 9687.
- Wang, Z., Gu, C., Wang, X., Lang, Y., Wu, Y., Wu, X., Zhu, X., Wang, K. and Yang, H. (2019) Caffeine enhances the anti-tumor effect of 5-fluorouracil via increasing the production of reactive oxygen species in hepatocellular carcinoma. *Med. Oncol.* **36**, 97.
- Wei, H. (1992) Activation of oncogenes and/or inactivation of anti-oncogenes by reactive oxygen species. *Med. Hypotheses* **39**, 267-270.
- Wei, X., Mo, X., An, F., Ji, X. and Lu, Y. (2018) 2',4'-Dihydroxy-6'-methoxy-3',5'-dimethylchalcone, a potent Nrf2/ARE pathway inhibitor, reverses drug resistance by decreasing glutathione synthesis and drug efflux in BEL-7402/5-FU cells. *Food Chem. Toxicol.* **119**, 252-259.
- Wu, Q., Wu, W., Fu, B., Shi, L., Wang, X. and Kuca, K. (2019) JNK signaling in cancer cell survival. *Med. Res. Rev.* **39**, 2082-2104.
- Xie, X., Liu, H., Wang, Y., Zhou, Y., Yu, H., Li, G., Ruan, Z., Li, F., Wang, X. and Zhang, J. (2016) Nicotinamide N-methyltransferase enhances resistance to 5-fluorouracil in colorectal cancer cells through inhibition of the ASK1-p38 MAPK pathway. *Oncotarget* **7**, 45837-45848.
- Xu, Y., Wellner, D. and Scheinberg, D. A. (1997) Cryptic and regulatory epitopes in CD13/aminopeptidase N. *Exp. Hematol.* **25**, 521-529.
- Yan, J., Dou, X., Zhou, J., Xiong, Y., Mo, L., Li, L. and Lei, Y. (2019) Tubeimoside-I sensitizes colorectal cancer cells to chemotherapy by inducing ROS-mediated impaired autophagosomes accumulation. *J. Exp. Clin. Cancer Res.* **38**, 353.
- You, F., Aoki, K., Ito, Y. and Nakashima, S. (2009) AKT plays a pivotal role in the acquisition of resistance to 5-fluorouracil in human squamous carcinoma cells. *Mol. Med. Rep.* **2**, 609-613.
- Yu, L., Ma, R., Wang, Y. and Nishino, H. (1994) Potent anti-tumor activity and low toxicity of tubeimoside 1 isolated from *Bolbostemma paniculatum*. *Planta Med.* **60**, 204-208.
- Zhang, D., Zhou, Q., Huang, D., He, L., Zhang, H., Hu, B., Peng, H. and Ren, D. (2019) ROS/JNK/c-Jun axis is involved in oridonin-induced caspase-dependent apoptosis in human colorectal cancer cells. *Biochem. Biophys. Res. Commun.* **513**, 594-601.
- Zhang, D. D. (2006) Mechanistic studies of the Nrf2-Keap1 signaling pathway. *Drug Metab. Rev.* **38**, 769-789.
- Zhang, P., Lai, Z. L., Chen, H. F., Zhang, M., Wang, A., Jia, T., Sun, W. Q., Zhu, X. M., Chen, X. F., Zhao, Z. and Zhang, J. (2017) Curcumin synergizes with 5-fluorouracil by impairing AMPK/ULK1-dependent autophagy, AKT activity and enhancing apoptosis in colon cancer cells with tumor growth inhibition in xenograft mice. *J. Exp. Clin. Cancer Res.* **36**, 190.
- Zhang, R., Pan, T., Xiang, Y., Zhang, M., Feng, J., Liu, S., Duan, T., Chen, P., Zhai, B., Chen, X., Wang, W., Chen, B., Han, X., Chen, L., Yan, L., Jin, T., Liu, Y., Li, G., Huang, X., Zhang, W., Sun, Y., Li, Q., Zhang, Q., Zhuo, L., Xie, T., Wu, Q. and Sui, X. (2020) beta-Elemene reverses the resistance of p53-deficient colorectal cancer cells to 5-fluorouracil by inducing pro-death autophagy and cyclin D3-dependent cycle arrest. *Front. Bioeng. Biotechnol.* **8**, 378.
- Zhao, H., Liu, Q., Wang, S., Dai, F., Cheng, X., Cheng, X., Chen, W., Zhang, M. and Chen, D. (2017) *In vitro* additive antitumor effects of dimethoxycurcumin and 5-fluorouracil in colon cancer cells. *Cancer Med.* **6**, 1698-1706.
- Zheng, W., Zhou, C. Y., Zhu, X. Q., Wang, X. J., Li, Z. Y., Chen, X. C., Chen, F., Che, X. Y. and Xie, X. (2018) Oridonin enhances the cytotoxicity of 5-FU in renal carcinoma cells by inducing necroptotic death. *Biomed. Pharmacother.* **106**, 175-182.

Zou, X., Liang, J., Sun, J., Hu, X., Lei, L., Wu, D. and Liu, L. (2016) Allicin sensitizes hepatocellular cancer cells to anti-tumor activity of 5-fluorouracil through ROS-mediated mitochondrial pathway. *J. Pharmacol. Sci.* **131**, 233-240.

Zu, C., Qin, G., Yang, C., Liu, N., He, A., Zhang, M. and Zheng, X. (2018) Low dose Emodin induces tumor senescence for boosting breast cancer chemotherapy via silencing NRARP. *Biochem. Biophys. Res. Commun.* **505**, 973-978.