

# Phylogenetic Characteristics of *Fasciola hepatica* Isolated from a Korean Patient

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**Abstract:** Fascioliasis is a parasitic infection caused by liver flukes. Although several cases have been reported in Korea, phylogenetic analysis of isolates is lacking. In this study, a 66-year-old woman with right upper quadrant (RUQ) abdominal pain was diagnosed as fascioliasis involving abdominal muscle by imaging study. She received praziquantel treatment, but symptoms were not improved. Lateral movement of the abscess lesion was followed. Trematode parasite was surgically removed from the patient's rectus abdominis muscle. The fluke was identified as *Fasciola hepatica* based on sequence analysis of 18S rDNA. To determine the phylogenetic position of this *Fasciola* strain (named Korean *Fasciola* 1; KF1), the *cox1* gene (273 bp) was analyzed and compared with the genes of 17 *F. hepatica* strains isolated from cows, sheep, goats, and humans from various countries. Phylogenetic analysis showed that KF1 was closely related with the isolates from China goat.

**Key words:** *Fasciola hepatica*, genetic analysis, phylogenetic

Fascioliasis in livestock and humans are caused by the genus *Fasciola* [1]. *Fasciola* spp. are prevalent in temperate climates. Fascioliasis is one of the neglected zoonotic diseases declared by the World Health Organization (WHO) [2]. Incidence of fascioliasis has been significantly increased since 1980, affecting approximately 2.4 million people globally each year. In humans, infection occurs through ingestion of contaminated water containing *Fasciola* metacercaria(e). The larvae pass through the stomach, break through the duodenal barrier, penetrate the abdominal cavity, and enter the hepatobiliary system. When the ingested fluke moves to the bile duct through the abdominal cavity and liver parenchyma, extensive bleeding and inflammation occur, causing thickening and expansion of the bile duct and gall bladder [3,4]. The symptoms and signs of human fascioliasis occur in 2 stages. The hepatic stage of the disease occurs when juvenile *Fasciola* enters the liver and begins to migrate to the biliary root canal; this is referred to as the acute or invasive stage. This stage occurs 1-3 months after the ingestion of metacercariae, in which typical

symptoms and signs include fever, hives, hepatomegaly, and pain in the right hypochondrium. When the worm infects biliary tract, cholangitis, cholestasis, right upper quadrant (RUQ) pain, and eosinophilia are usually developed [5-7].

Genetic diversity of *Fasciola* populations has been demonstrated via molecular analysis of their nuclear and mitochondrial DNAs (mtDNAs). The polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) analysis of mtDNA revealed 52 unique sequences in 221 flukes [8]. A study on *Fasciola* isolates from livers of sheep and cattle suggested grouping of these isolates into 3 main lineages (*F. hepatica*, *F. gigantica*, and *F. indica*), using the *cox1* (encoding cytochrome c oxidase subunit) gene. Ai et al. [1] also inferred phylogeny using the *cox1*, which is a useful marker for studying the genetic differentiation and phylogenetic relationship of *Fasciola*.

Infection with *Fasciola* spp. has been reported in East Asia, including Korea (Table 1) and Japan, while few molecular taxonomic studies have been conducted on Korean species [10]. In this study, the genetic characteristics of *Fasciola* isolated from a Korean patient were compared with those of several previously reported isolates using the *cox1* gene sequencing and multiple alignment methods.

A 66-year-old Korean woman living in Pusan visited the Pusan National University Hospital on July 11, 2021, with an ab-

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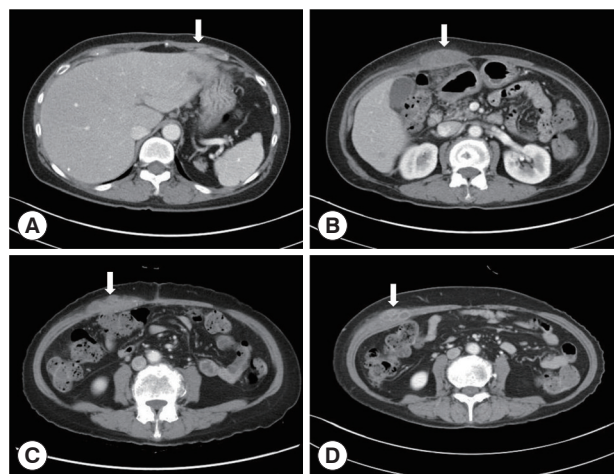
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**Table 1.** Information of human fascioliasis patients in Korea

Year	Sex	Age	Infection site	First symptom	Second symptom	First/final diagnosis	Gene study	Reference
1976	F	42	Bile duct	Urticaria, dyspepsia, epigastric discomfort	Epigastric colicky pain attacks	-/fascioliasis	-	[13]
1984	F	27	Cecum	Nausea, vomiting, epigastric tenderness.	Palpable mass	Bowel obstruction/fascioliasis	-	[14]
1991	F	32	Left chest wall	Palpable mass	-	-/fascioliasis	-	[19]
1994	M	28	Intraocular	Headache, motor weakness	Corneal edema with hyphema	Corneal edema/intraocular fascioliasis	-	[15]
2006	F	46	Pancreas	Left upper quadrant pain	Peripheral eosinophilia, hyperamylasemia, hyperlipasemia	Acute pancreatitis/pancreatic fascioliasis	-	[16]
2014	M	87	Bile duct	Abdominal pain and discomfort	Peripheral eosinophilia	Gallbladder stone and intraductal cholangiocarcinoma/fascioliasis	ITS-1	[20]
2015	F	56	Colon	Discomfort and pain in the abdomen	Tenderness in the abdomen, eosinophilia	Cysticercosis/fascioliasis	-	[12]
2021	F	66	Abdomen	Right upper quadrant (RUQ) pain	Pain in the subcostal region, eosinophilia	Parametric infection/fascioliasis	Cox-1	This study

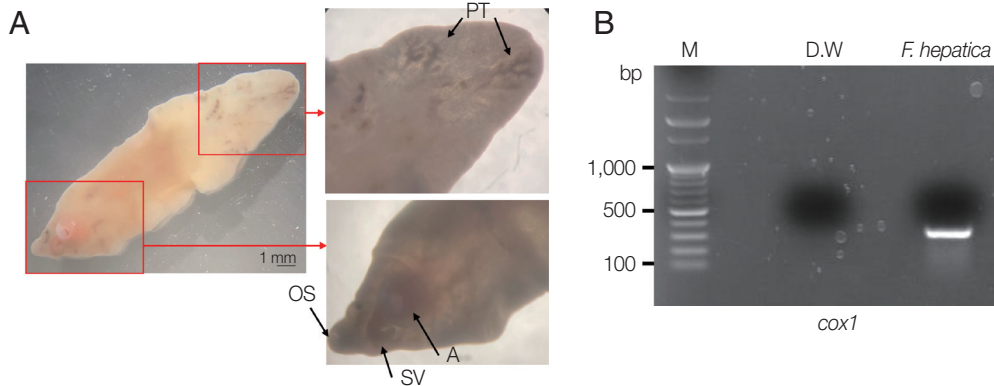
M, male; F, female.

dominal mass. She had been suffered from hypothyroidism. In June 2021, she was diagnosed with a hepatic abscess by computed tomography (CT) performed during an evaluation of RUQ pain at another hospital. The symptoms improved by symptomatic treatment and she was discharged after administration of antibiotics (ceftriaxone and metronidazole) (Fig. 1A). She visited again our hospital due to recurrence of the RUQ pain. The mass was palpable in the abdominal cavity, we checked the mass with CT. Some swelling and cell infiltration was confirmed, and parasitic infection was suspected (Fig. 1B). She received praziquantel (25 mg/kg 3 days). Most fascioliasis patients showed eosinophilia and abdominal distension [11]. Abdominal distension along with eosinophilia was also confirmed in this patient. In August 2021, liver abscess was percutaneously drained, after which she was discharged. However, she complained of new pain in the subcostal region (Fig. 1C). Praziquantel treatment were maintained, but her symptoms did not improve. New multiple abscesses were found below the previous lesion on abdominal CT. Compared to the previous CT, a lateral movement of the abscess was observed with maximum size of 3.4 cm (Fig. 1D). The CT showed abdominal distension. The laboratory test showed eosinophilia (7.3%, normal range 0-6.9%) with low number of segmented neutrophil (38.8%, normal range 40.0-70.4%). The mean corpuscular hemoglobin concentration, eosinophil count, and erythrocyte sedimentation rate increased, and the levels of segmented neutrophils decreased. A biopsy revealed a leaf-like parasite. The



**Fig. 1.** Abdominal computerized tomography (CT). (A) CT performed at the time of admission for evaluation of pain in the right upper quadrant (RUQ). (B) RUQ pain recurred after treatment of liver abscess. CT was taken to evaluate the abdominal wall mass. (C) Abdominal CT performed with new pain in the subcostal area. (D) There was no improvement after maintenance treatment with praziquantel. CT was taken again at Pusan National University Hospital. The arrows indicated a mass lesion.

length was approximately 1.7-1.8-long, and the width was 0.5-0.8-long. The morphology of the posterior testis of the *F. hepatica* was observed under a microscope. The fluke did not have hooks or spines but were characterized by suckers. The acetabulum was located at the anterior part of the worm (Fig. 2A). The granuloma contained acute and chronic inflammatory reactions accompanied by numerous eosinophilic invasive

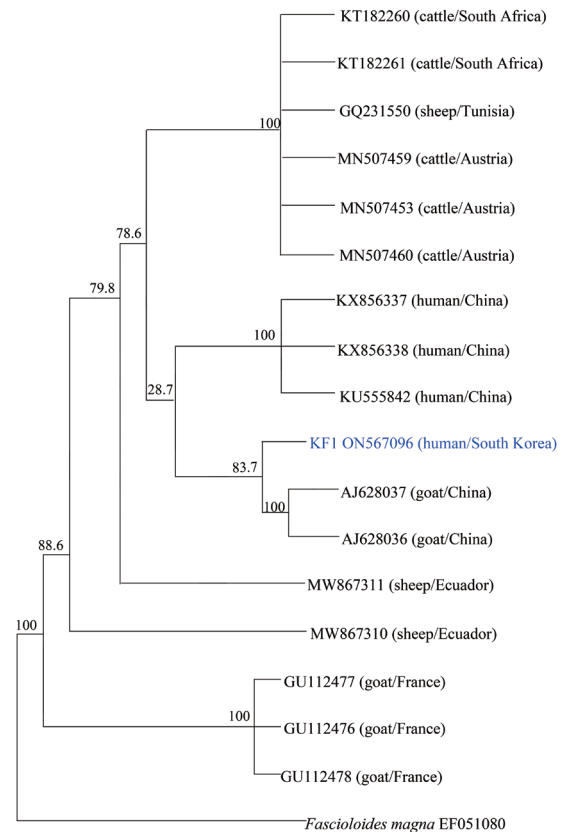


**Fig. 2.** Morphological characteristics of liver fluke and electrophoretic analysis of PCR products. (A) Morphological characteristics of *F. hepatica* isolated from the patient. OS, oral sucker; A, acetabulum (ventral sucker); SV, seminal vesicle; PT, posterior testis. (B) *Cox1* partial PCR product (M, 100-kb ladder).

micronecrosis.

This patient came to our hospital with a liver abscess, but was diagnosed with abdominal muscular fascioliasis invoking RUQ pain. A parasitic infection caused by *Fasciola hepatica* was confirmed by histological observation of biopsied material. *F. hepatica* usually thrives in the hepatobiliary system, it often migrates aberrantly and causes ectopic fascioliasis [12]. Ectopic fascioliasis in the various sites, such as the cecum, ascending colon, brain, eyes, spine, subcutaneous tissue, neck, and inguinal lymph nodes, have been reported in Korea [13-15]. In 2006, a very rare case of pancreatic infection was also reported in Korea [16]. *F. hepatica* infection should be suspected in the differential diagnosis of parasitic infection based on the patient's dietary history, eosinophilia, symptoms, and results of ELISA tests.

To molecularly identify *F. hepatica*, PCR was performed on worm's DNA sample. We also establish the phylogenetic classification of this specimen using the *F. hepatica* 18S rRNA and *cox1* gene sequences. Their genetic mutations were compared with 17 previously reported strains. Amplification of the *cox1* gene was performed using primers *cox1* F 5'-TTTGCCITGGTTTGGAGTTA-3' and *cox1* R 5'-CCACACAACAGGATCCCA-TA-3'. The PCR conditions were optimized according to previous reports [17]. Through PCR amplification, a target band with a length of approximately 273 bp was obtained for the *cox1* gene (Fig. 2B). DNA sequencing was performed using Cosmo Genetech (Seoul, Korea). Nucleotide sequences were grouped according to host species (cattle, sheep, goat, and *Homo sapiens*) and region (South Africa, Tunisia, Austria, Ecuador, France, and China). Isolates and reference sequences were used for multiple alignments and phylogenetic distances were calculated using MEGA version 6.0. Phylogenetic trees were constructed using the neighbor-joining method in the MEGA version 6.0 using *cox1* partial sequences isolated from a Korean



**Fig. 3.** Phylogenetic tree of *F. hepatica* isolates generated using *cox1* partial gene sequences. All of *F. hepatica* was represented by Genbank accession numbers. The genetic relationship between *F. hepatica* KF1 (ON567096) and 17 corresponding reference sequences was analyzed by the neighbor-joining method using MEGA, version 6.0.

patient (KF1) with 17 previously reported isolates. As shown in Fig. 3, the phylogenetic tree clearly separated these homologs according to the host and region. KF1 was found to be closely related to *F. hepatica* from goats in China, and the genetic characteristics of most of the isolates were geographically related, with no relationship was observed along with host differences.

Fascioliasis is generally diagnosed by detection of the characteristic eggs in feces. However, at low levels of infection, the sensitivity is very low even after repeated stool examinations. Serological tests are appropriate for diagnosing patients at the invasive stage, as they allow for early detection of infection. Anti-*Fasciola* antibodies can be detected 2-4 weeks after infection [6,18].

A limitation related to the interpretation of the phylogenetic results in this study is that only one Korean isolate was used for the analysis. Phylogenetic analysis has revealed that goat cattle, and sheep share *F. hepatica* strain that can transmit to humans. In the context of the prevention of human fascioliasis, the pattern and phylogenetic aspects of parasitic infections appear to be very important. In Korea, there was genetic confirmation of *F. hepatica*, but there were no related data in the phylogenetic analysis. A study on the distribution of *F. hepatica* is needed for future epidemiological investigations of domestic infections.

## CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

## REFERENCES

1. Ai L, Cai YC, Lu Y, Chen JX, Chen SH. Human cases of fascioliasis in Fujian Province, China. *Korean J Parasitol* 2017; 55: 55-60. <https://doi.org/10.3347/kjp.2017.55.1.55>
2. Caravedo MA, Cabada MM. Human fascioliasis: current epidemiological status and strategies for diagnosis, treatment, and control. *Res Rep Trop Med* 2020; 11: 149-158. <https://doi.org/10.2147/RRTM.S237461>
3. Parkinson M, O'Neill SM, Dalton JP. Endemic human fasciolosis in the Bolivian Altiplano. *Epidemiol Infect* 2007; 135: 669-674. <https://doi.org/10.1017/S095026880600728X>
4. Kaya M, Beştaş R, Cetin S. Clinical presentation and management of *Fasciola hepatica* infection: single-center experience. *World J Gastroenterol* 2011; 17: 4899-4904. <https://doi.org/10.3748/wjg.v17.i44.4899>
5. Aksoy DY, Kerimoglu U, Oto A, Erguven S, Arslan S, Unal S, Batman F, Bayraktar Y. Infection with *Fasciola hepatica*. *Clin Microbiol Infect* 2005; 11: 859-861. <https://doi.org/10.1111/j.1469-0691.2005.01254.x>
6. Mas-Coma S, Bargues MD, Valero MA. Diagnosis of human fascioliasis by stool and blood techniques: update for the present global scenario. *Parasitology* 2014; 141: 1918-1946. <https://doi.org/10.1017/S0031182014000869>
7. Schiappacasse RH, Mohammadi D, Christie AJ. Successful treatment of severe infection with *Fasciola hepatica* with praziquantel. *J Infect Dis* 1985; 152: 1339-1340. <https://doi.org/10.1093/infdis/152.6.1339>
8. Elliott T, Muller A, Brockwell Y, Murphy N, Grillo V, Toet HM, Anderson G, Sangster N, Spithill TW. Evidence for high genetic diversity of NAD1 and COX1 mitochondrial haplotypes among triclabendazole resistant and susceptible populations and field isolates of *Fasciola hepatica* (liver fluke) in Australia. *Vet Parasitol* 2014; 200: 90-96. <https://doi.org/10.1016/j.vetpar.2013.11.019>
9. Akhlaghi E, Mohammadi MA, Ziaali N, Baneshi MR, Nasibi S, Kamyabi H, Rostami S, Harandi MF. Morphometric and molecular study of *Fasciola* isolates from ruminants in Iran. *Turkiye Parazitoloj Derg* 2017; 41: 192-197. <https://doi.org/10.5152/tpd.2017.5214>
10. Agatsuma T, Arakawa Y, Iwagami M, Honzako Y, Cahyaningsih U, Kang SY, Hong SJ. Molecular evidence of natural hybridization between *Fasciola hepatica* and *F. gigantica*. *Parasitol Int* 2000; 49: 231-238. [https://doi.org/10.1016/S1383-5769\(00\)00051-9](https://doi.org/10.1016/S1383-5769(00)00051-9)
11. Park HJ, Choi GS, Jung M, Lee SU. *Fasciola hepatica* induced hepatic abscess treated with triclabendazole. *Korean J Gastroenterol* 2021; 77: 39-44. <https://doi.org/10.4166/kjg.2020.152>
12. Kim AJ, Choi CH, Choi SK, Shin YW, Park YK, Kim L, Choi SJ, Han JY, Kim JM, Chu YC, Park IS. Ectopic Human *Fasciola hepatica* infection by an adult worm in the mesocolon. *Korean J Parasitol* 2015; 53: 725-730. <https://doi.org/10.3347/kjp.2015.53.6.725>
13. Lee SH, Cho SY, Seo BS, Choe KJ, Chi JG. A human case of ectopic fascioliasis in Korea. *Korean J Parasitol* 1982; 20: 191-200. <https://doi.org/10.3347/kjp.1982.20.2.191>
14. Park CI, Kim H, Ro JY, Gutierrez Y. Human ectopic fascioliasis in the cecum. *Am J Surg Pathol* 1984; 8: 73-77. <https://doi.org/10.1097/00000478-198401000-00008>
15. Cho SY, Yang HN, Kong Y, Kim JC, Shin KW, Koo BS. Intraocular fascioliasis: a case report. *Am J Trop Med Hyg* 1994; 50: 349-353. <https://doi.org/10.4269/ajtmh.1994.50.349>
16. Lee OJ, Kim TH. Indirect evidence of ectopic pancreatic fascioliasis in a human. *J Gastroenterol Hepatol* 2006; 21: 1631-1633. <https://doi.org/10.1111/j.1440-1746.2006.03185.x>
17. Choi IW, Kim HY, Quan JH, Ryu JG, Sun R, Lee YH. Monitoring of *Fasciola* species contamination in water dropwort by cox1 mitochondrial and ITS-2 rDNA sequencing analysis. *Korean J Parasitol* 2015; 53: 641-645. <https://doi.org/10.3347/kjp.2015.53.5.641>
18. Gonzales Santana B, Dalton JP, Vasquez Camargo F, Parkinson M, Ndao M. The diagnosis of human fascioliasis by enzyme-linked immunosorbent assay (ELISA) using recombinant cathepsin L protease. *PLoS Negl Trop Dis* 2013; 7: e2414. <https://doi.org/10.1371/journal.pntd.0002414>
19. Chang EC, Choi HL, Park YW, Kong Y, Cho SY. Subcutaneous fascioliasis: a case report. *Korean J Parasitol* 1991; 29: 403-405. <https://doi.org/10.3347/kjp.1991.29.4.403>
20. Kang BK, Jung BK, Lee YS, Hwang IK, Lim H, Cho J, Hwang JH, Chai JY. A case of *Fasciola hepatica* infection mimicking cholangiocarcinoma and ITS-1 sequencing of the worm. *Korean J Parasitol* 2014; 52: 193-196. <https://doi.org/10.3347/kjp.2014.52.2.193>