RESEARCH ARTICLE

Report on the Unrecorded *Helvella* Genus Found in Taebaek City, South Korea

Sangyoung Park¹, Sohee Kim¹, Eunjin Kim², Ju-Kyung Eo^{3,*}, and Hwayong Lee^{1,*}

¹Department of Forest Science, Chungbuk National University, Cheongju 28644, Koera

²Department of Forest Therapy, Chungbuk National University, Cheongju 28644, Koera

³Ecological Technology Research Team, Division of Ecological Applications Research, National Institute of Ecology, Seocheon 33657, Koera

*Corresponding authors: abiesendo@gmail.com, leehy@chungbuk.ac.kr

ABSTRACT

In July 2023, previously unrecorded fungi belonging to the genus *Helvella* were collected from the city of Taebaek, South Korea. These fungi are morphologically similar to *Helvella crispa*, but their differences include a wide, saddle-shaped apothecium and white hair on the receptacle surface. By analyzing DNA sequences combining the internal transcribed spacer (ITS) and nuclear ribosomal large subunit (nrLSU) regions and comparing them with various related species of *Helvella*, the collected fungi were identified as *H. orienticrispa*.

Keywords: *Helvella,* ITS (internal transcribed spacer), nrLSU (nuclear ribosomal large subunit), unreported *helvella*





pISSN: 0253-651X **eISSN**: 2383-5249

Kor. J. Mycol. 2023 December, 51(4): 411-417 https://doi.org/10.4489/kjm.20230042

Received: October 16, 2023 Revised: December 18, 2023 Accepted: December 20, 2023

© 2023 THE KOREAN SOCIETY OF MYCOLOGY.



This is an Open Access article distributed

under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

The genus *Helvella* (Helvellacae, Peziziomycetes) is characterized by saddle or cup-shaped fruiting bodies with lobes or wrinkles, and is commonly found in valleys, mossy areas, or grassy fields. Some *Helvella* species play an important ecological role in forest ecosystems by growing mycorrhizae [1], while *H. jocatoi* and *H. bachu* are used as food ingredients in some countries [2,3], Therefore, *Helvella* species are valuable forest resources.

Research on the *Helvella* genus has previously been conducted in Europe and the USA, with a primary focus on species diversity and phylogenetic relationships [4-13]. In Europe, using morphological characteristics and molecular biology data, the boundaries of some challenging-to-distinguish *Helvella* species have been clearly delineated, revealing that many *Helvella* species exhibit regional endemism [8]. This highlights the need for future research on Asian *Helvella* species. Since then, active research on Asian *Helvella* species has been conducted in China, revealing the presence of numerous cryptic species within *Helvella* and highlighting the differences in their distribution between Europe and Asia [9,11,13-15]. New and unrecorded *Helvella* species continue to be reported worldwide. In Korea, there had been no reports of *H. costifera* prior to its discovery in 2010 [16].

In this study, *Helvella* spp. discovered in Taebaek City were analyzed using morphological and molecular biological approaches. In the present study, we confirmed that this species has not been recorded in Korea; therefore, it was listed as an unrecorded species.

MATERIALS AND METHODS

Morphological observation

A fungal diversity survey was conducted between July and August, 2023 in Taebaek City, Korea. Before collection, sample photographs were taken using a camera (Olympus OM-D E-M1 Mark III, Tokyo, Japan), and GPS coordinates and habitat information were recorded. The collected samples were air-dried at 40°C for 72 h for the purpose of DNA extraction. Microscopic observations were performed using an optical microscope (BX53, Olympus, Tokyo, Japan). Microstructural measurements were performed using a camera (DP22, Olympus, Tokyo, Japan) integrated with a microscope and its software (DP2-SAL, Olympus, Tokyo, Japan). Twenty ascospores and ten microstructures, excluding the ascospores, were observed.

DNA extraction and phylogenetic analysis

For the purpose of phylogenetic analysis, genomic DNA was extracted from the specimens using Exgene Plant SV Mini (Geneall, Seoul, Korea). Polymerase chain reaction (PCR) was performed to amplify partial sequences from two partial genes (internal transcribed spacer [ITS] and nuclear ribosomal large subunit [nrLSU]) using previously published primer sets. The primer sets used in this study were: ITS1 and ITS4 for ITS [17], and hLSUf and hLSUr for nrLSU [18] (Table 1). Amplification reactions were performed in 20 μL reaction volumes containing 1 μL DNA template, 1 μL per primer (10 μM), 10 μL GainBlueTM Hot Start Master Mix, 2X (GainBio, Daejeon, Korea), and 7 µL ddH₂O. The PCR conditions involved an initial pre-denaturation step at 95°C for 4 minutes, followed by 40 cycles of denaturation at 95°C for 30 seconds, annealing at 64°C for 30 seconds, extension at 72°C for 60 seconds, followed by a final extension step at 72°C for 10 minutes. PCR products were visualized on a 1% agarose gel and purified using an Expin PCR SV kit (GeneAll, Seoul, Korea). Sanger sequencing was performed by Cosmogenentech (Daejeon, Korea). Sequences were aligned and edited in MUSCLE 5.1 [19] using default parameters. Ambiguously aligned regions and gaps in the alignment were excluded before analysis. Alignments were constructed separately for each gene fragment using MUSCLE 5.1 and then concatenated using Geneious Prime 2023.2 (https:// www.geneious.com). Poorly aligned sites were identified using Gblocks v. 0.91b [20] with the default parameters. All the identified ambiguous sites were excluded from the analysis. All phylogenetic analyses were performed based on maximum likelihood (ML) analysis in IQtree2.2.2.7 [21] available on the CIPRES Web portal [22] using a TIM+I+G model with 1,000 bootstrap replicates. The optimal substitution model was selected using jModelTest 2.1.6 [23].

Table 1. Primer information.

Primer name	Sequences $(5' \rightarrow 3')$	References
ITS1	TCC GTA GGT GAA CCT GCG C	White et al. [17]
ITS4	TCC TCC GCT TAT TGA TAT GC	
H_LSUf1	AGC GGA GGA AAG AAA CCA ACA	Landeros et al. [18]
H_LSUr2	TCC CAA CAG CTA TGC TCC TAC	

ITS: internal transcribed spacer; nrLSU: nuclear ribosomal large subunit.

RESULTS

Molecular phylogenetics

The nucleotide sequences of the 31 samples (a combination of from the ITS and nrLSU genes) were used to construct a phylogenetic tree. Reference sequences for this analysis were obtained from Zhao's study [14]. The taxa included in the phylogenetic tree comprised five species belonging to the Crispa clade (*H. crispa*, *H. orienticrispa*, *H. involuta*, *H. pseudoreflexa*, *H. zhongtiaoensis*) and one species belonging to the Lacunosa clade (*H. rugosa*). Except for *H. crispa*, all samples were collected from China [14]. The nucleotide sequence of the sample (KH058) used in this study, which was collected from the southwestern region of China, exhibited high bootstrap support (96%) and formed a monophyletic clade. In particular, it demonstrated a monophyletic relationship with the reference specimen *H. orienticrispa* (HKAS74319) with 99% bootstrap support (Fig. 1).

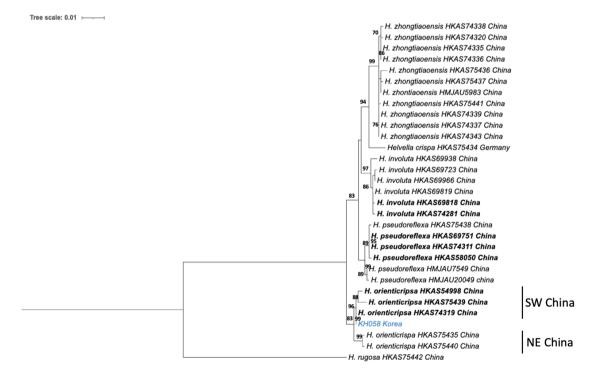


Fig. 1. Phylogenetic tree inferred from maximum likelihood (ML) analysis using ITS-nrLSU sequence. Sequence generated for this study are highlighted in blue. Type species are indicated in bold. ITS, internal transcribed spacer; nrLSU, nuclear ribosomal large subunit; SW, Southwestern China; NE, Northeastern China.

Taxonomy

동양주름안장버섯(신칭) *Helvella orienticrispa* Q. Zhao, Zhu L. Yang & K.D. Hyde, in Zhao, Tolgor, Zhao, Yang & Hyde, *Phytotaxa* 239(2): 136 (2015)

Apothecium broadly saddle-shaped to irregularly lobed, 3-4 cm high, 2-4 cm broad; margin rolled to hymenium when young, but extended and split at maturity, never fused with stipe; hymenium glabrous, pale cream to white, becoming yellowish when dried; receptacle surface finely pubescent, blunt ribs covering almost 1/3 of apothecium, pale cream to white, and yellowish when dried. Stipe 3-7 cm long, 1-1.5 cm thick, with deep lengthwise grooves and cross-veins; ribs blunt, mostly double-edged, finely pubescent, slightly brittle, pale cream to white, cream when dried, basal mycelium white. Medullary excipulum 130-260 μ m broad, textura intricata, hyaline, composed of thick-walled hyphae 2.5-6 μ m broad. Ectal excipulum 70-200 μ m, outmost cells catenuliform in long fascicled tufts, hyaline, evenly indigo blue in cotton blue with cylindrical to subclavate, slightly thick-walled end cells 24-36×10-14 μ m. Asci 264-312×10-18 μ m, 8-spored, subcylindrical to clavate. Ascospores (16.2-) 18.3-18.5 (-19.8)×(10.6-) 12.1-12.3 (-13.5) μ m, [Q=(1.38) 1.51-1.53 (1.73) (Q_{av}=1.52±0.09)], broadly ellipsoid to ellipsoid, walls smooth, slightly thickened. Paraphyses filiform, 3-4.2 μ m broad, slightly exceeding the asci, with a yellow refractive content in Cotton blue; apex slightly enlarged 6-8.3 μ m broad (Fig. 2).

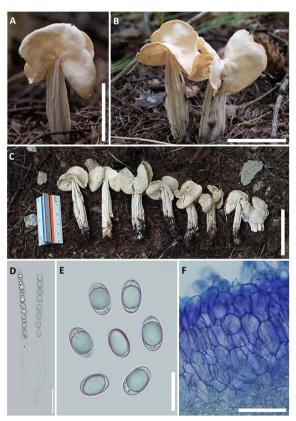


Fig. 2. Morphological characteristics of *Helvella orienticrispa*. A-C: Ascomata of KH058. D: Asci of KH058 in water. E: Ascospores of KH058 in water. F: Ectal excipulum of KH058 in cotton blue. (Scale bar: A-C=5 cm; D, F=50 μm; E=20 μm).

Habitat.

Solitary or scattered on the ground, in *Larix kaempferi* (Lamb.) Carrière and *Quercus mongolica* Fisch. ex Ledeb forest

Specimen examined. Republic of Korea, Taebaek City, Geomryongso, Gangwon Province, 22 Jul. 2023 (KH058); N 37.23058. E 128.03514. GenBank no. OR647322 (ITS), and OR647324 (nrLSU).

Remark.

H. orienticrispa is very similar to the species recorded in Korea namely, *H. crispa*, to the extent that it is difficult to distinguish between them with the naked eye. However, *H. crispa* typically has irregularly shaped apothecium, with fine brown hairs on the receptacle surface [24], while *H. orienticrispa* is characterized by typically having broad saddle-shaped ascocarps and fine white tufts on the receptacle surface, allowing for differentiation.

DISCUSSION

H. orienticrispa, a previously unrecorded species in Korea, was discovered in the present study, and grows mainly in coniferous forests [14]. This species is distinguished from other *Helvella* species by its broadly spread, saddle-shaped apothecium and its upper and outer surfaces of the ascocarp cup, which are both cream colored and have fine white hairs on the outer surface. On the other hand, *H. crispa*, was first reported in Europe in 1823; as phylogenetic classification studies of *Helvella* species have progressed worldwide, it has been found to possess regional endemism, and several cryptic species with very similar morphologies have also been identified [8,11,14]. Recent phylogenetic classification studies of *Helvella* species in China suggest that *H. crispa* is not present in China [11], instead, several new species similar to *H. Crispa*, such as *H. involuta*, *H. orienticrispa*, *H. pseudoreflexa*, and *H. zhongtiaoensis*, have been recently reported [14]. Therefore, it is necessary to evaluate the presence or absence of *H. crispa* in South Korea.

CONFLICT OF INTERESTS

No conflict of interest was reported by the authors.

ACKNOWLEDGEMENTS

This work was supported by a grant from the National Institute of Ecology (NIE), funded by the Ministry of Environment (MOE) of the Republic of Korea (NIE-B-2023-18).

REFERENCES

 Hwang J, Zhao Q, Yang ZL, Wnag Z, Townsend JP. Solving the ecological puzzle of mycorrhizal associations using data from annotated collections and environmental samples-an example of saddle fungi. Environ Microbiol Rep 2015;7:658-67.

- Landeros F, Ferrusca-Rico FM, Guzmán-Dávalos L, Esquivel-Naranjo EU, Matías-Ferrer N, Burrola-Aguilar C, Viurcos-Martínez GA, Garibay-Orijel R. Helvella jocatoi sp. nov. (Pezizales, Ascomycota), a new species from H. lacunosa complex with cultural importance in central Mexico Abies religiosa forests. Phytotaxa 2021;498:1-11.
- 3. Zhao Q, Sulayman M, Zhu X-T, Zhao Y-C, Yang Z-L, Hude KD. Species clarification of the culinary Bachu mushroom in western China. Mycologia 2016;108:828-36.
- 4. Dissing H, Lange M. Notes on the genus Helvella in North America. Mycologia 1967;59:349-60.
- 5. Abbott SP, Currah R. The genus Helvella in Alberta. Mycotaxon 1988;33:229-50.
- Nguyen NH, Landeros F, Garibay-Orijel R, Hansen K, Vellinga EC. The *Helvella lacunosa* species complex in western North America: cryptic species, misapplied names and parasites. Mycologia 2013;105:1275-86.
- 7. Landeros F, Iturriaga T, Guzmán-Dávalos L. Type studies in *Helvella* (Pezizales) 1. Mycotaxon 2012;119:35-63.
- 8. Skrede I, Carlsen T, Schumacher T. A synopsis of the saddle fungi (Helvella: Ascomycota) in Europe-species delimitation, taxonomy and typification. Persoonia 2017;39:201-53.
- Wang X-C, Liu T-Z, Chen S-L, Li Y, Zhuang W-Y. A four-locus phylogeny of rib-stiped cupulate species of *Helvella* (Helvellaceae, Pezizales) with discovery of three new species. MycoKeys 2019;60:45-67.
- 10. Løken SB, Skrede I, Schumacher T. The *Helvella corium* species aggregate in Nordic countries-phylogeny and species delimitation. Fungal Syst Evol 2020;5:169-86.
- 11. Mao N, Xu Y, Zhang Y, Zhou H, Huang X, Hou C-L, Fan L. Phylogeny and species diversity of the genus *Helvella* with emphasis on eighteen new species from China. Fungal Syst Evol, 2023;12:111-52.
- 12. Skrede I, Løken SB, Mathiesen C, Schumacher T. Additions to the knowledge of the genus *Helvella* in Europe. New records and de novo description of five species from the Nordic region. F Fungal Syst Evol 2023;11:71-84.
- 13. Wang X-C, Zhuang W-Y, Zhao R-L. Species diversity of *Helvella lacunosa* clade (Pezizales, Ascomycota) in China and description of sixteen new species. J Fungi 2023;9:697.
- 14. Zhao Q, Tolgor B, Zhao Y, Yang ZL, Hyde KD. Species diversity within the *Helvella crispa* group (Ascomycota: Helvellaceae) in China. Phytotaxa 2015;239:130-42.
- 15. Xu R-J, Li L, Zhao Q. *Helvella cystidiata sp. nov.* (Helvellaceae, Ascomycota) from Tibetan Plateau, China. Phytotaxa 2022;560:82-92.
- 16. Han S-K, Oh S-H, Kim H-J. Eight unrecorded higher fungi identified at the Korea National Arboretum. Mycobiology 2010;38:81-88.
- 17. White TJ, Bruns T, Lee S, Tayler J. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. PCR protocols: a guide to methods and applications. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ, editors. Cambridge: Academic Press; 1990. p. 315-22.
- 18. Landeros F, Iturriaga T, Rodríguez A, Vargas-Amado G, Guzmán-Dávalos L. Advances in the phylogeny of *Helvella* (Fungi: Ascomycota), inferred from nuclear ribosomal LSU sequences and morphological data. Rev Mex Biodivers 2015;86:856-71.
- Edgar RC. MUSCLE: a multiple sequence alignment method with reduced time and space complexity. BMC Bioinform 2004;5:113.
- 20. Castresana J. Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. Mol Biol Evol 2000;17:540-52.

- 21. Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ. IQ-TREE: a fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. Mol Biol Evol 2015;32:268-74.
- 22. Miller MA, Pfeiffer W, Schwartz T. Creating the CIPRES science gateway for inference of large phylogenetic trees. In 2010 gateway computing environments workshop (GCE); 2010 Nov 14; New Orleans, USA. LA: IEEE Xplore; 2010. p. 1-8.
- 23. Darriba D, Taboada GL, Doallo R, Posada D. jModelTest 2: more models, new heuristics and parallel computing. Nat Methods 2012;9:772.
- 24. Dissing H. The genus *Helvella* in Europe with special emphasis on the species found in Norden. Odense: Denmark by Andelsbogtrykkeriet; 1966.