

A Brief Review on *Murraya paniculata* (Orange Jasmine): pharmacognosy, phytochemistry and ethnomedicinal uses

Deepa Joshi*, Kashmira J. Gohil

Department of Pharmacology, Anand College of Pharmacy, Agra, Uttar Pradesh, India

Received November 2, 2022

Reviewed January 17, 2023

Accepted February 8, 2023

*Corresponding Author

Deepa Joshi

Department of Pharmacology, Anand
College of Pharmacy, Agra, Uttar Pradesh
282007, India

Tel: +91-783-088-1582

E-mail: deepajoshi390@gmail.com

Objectives: *Murraya paniculata* (family-Rutaceae), popularly known as orange jasmine, is the most important evergreen plant. The Rutaceae family is economically significant due to its diverse edible fruits and essential oils.

Methods: *Murraya paniculata* extracts (MPE) of leaf have been shown to include phenolic compounds, highly oxygenated flavonoids, flavanones, sesquiterpenoids, polymethoxy glycosides, and coumarins. Cyclocitral, methyl salicylate, trans-nerolidol, cubenol, isoger-macrene, -cadinol, and cubeb-11-ene are all abundant in MPE. The usages of various parts of this plant, such as bark, leaves and flower, as a remedy for a variety of ailments as widely recorded in the traditional literature. The plant has anti-diabetic, anti-obesity, antibacterial, anti-implantation, anti-oxidative, cytotoxic, anti-diarrheal, antidepressant and anti-anxiety properties and many others.

Results: The goal of the review is to reignite interest in this potential plant, encouraging researchers to continue their research in order to uncover novel therapeutic compounds for the treatment and management of a range of infections. The current review provided a comprehensive overview of this traditional unique plant.

Conclusion: The review paves a way for exploring its active chemical elements with substantial pharmacological values further for potential benefits of mankind.

Keywords: herbal plants, murraya paniculata, therapeutic uses

INTRODUCTION

Herbal products are gaining popularity in both progressive and developing nations [1]. For thousands of years, medicinal plants have been used based on traditional and folk treatments, and their importance in treating moderate and chronic ailments is growing [2]. *Murraya paniculata* (MP), often called honey bush, orange jasmine, and kamini, is a traditional medicinal plant from the Rutaceae family. It is primarily found in India, Sri Lanka, southern China, Thailand, and eastward over the Malesian region to northeastern Australia and Caledonia [3]. MP is a genus of 12 species and evergreen shrubs [4]. It has the potential to limit *Diaphorina citri* migration into commercial citrus orchards, which is critical for better huanglongbing management [5]. Headache, bruises, gastralgia, stomachaches,

rheumatism, skin irritation, and swelling are all treated with this herb. It is also utilized as a menstrual flow booster and a snake bite treatment [4, 6]. The plant is also utilized for treating toothaches [7]. The reliable databases used for the review were Science Direct, Elsevier, and Research Gate, PubMed, Google Scholar, among others.

This review explicitly elaborated on MP, emphasizing on its recently studied phytoconstituents and pharmacological activities.

DESCRIPTION

The plant can grow to be 8-12 feet tall. MP has a taproot (Fig. 1), with fragrant white flowers and oval-shaped crimson fruits. The fruit length ranges from 5 to 1 inch [8].



Figure 1. *Murraya paniculata* plant.

MP leaves are egg-shaped, and are 2-11 cm long and 1-1.5 cm wide. The leaves have a fragrant aroma, a bitter, spicy flavor, and a brownish green color. They are smooth and shiny on the surface. Leaves can be detected in the mesophyll, or the epidermis without stomata, and the epidermis. Anticlinal cell walls, a lower epidermis covered in hair and rosette-shaped stomata are all characteristic of MP leaf powders [9].

MP is popularly cultivated in tropical countries, and under glass in temperate ones. The ideal altitude is about 200 meters as MP is grown on basalt or calcareous soils. Between March and June, the flowers are harvested (Table 1) [10]. MP's chloroplast genome is 160,280 bp long, with large and small single-copy regions measuring 87,605 and 18,609 bp, respectively, which are separated by two IR regions of 27,033 bp. The sample has a GC percentage of 38.61%. De novo assembly and annotation indicated the presence of unique genes and revealed the presence of 85, 29, and 8 protein-coding, tRNA, and rRNA genes, respectively [11].

PHYTOCONSTITUENTS OF DIFFERENT PARTS OF MP

Various phytoconstituents have been discovered, including flavonoids, alkaloids, carbohydrates, phenolic compounds, amino acids and proteins [3, 12-14]. Various chemical compounds have been isolated from several parts of the plant.

1. Leaves, shoots and twigs

Extracts from MP leaves contain coumarins compounds such as murrayanone and murraculatin [15]. Eight highly oxygenat-

Table 1. Botanical classification of *Murraya paniculata* [10]

Kingdom	Plantae
Phylum	Charophyta
Class	Equisetopsida
Subclass	Magnoliidae
Superorder	Rosanae
Order	Sapindales
Family	Rutaceae
Genus	Murraya
Species	Paniculata

ed flavones, namely, 5-hydroxy-6,7,3',4',5'-pentamethoxyflavone (umhengerin), 5-hydroxy-6,7,8,3',4',5'-hexamethoxy flavone (gardenin A), 6,7,8,4'-tetra-methoxy-5,3',5'-trihydroxyflavone (gardenin E), 5,3'-dihydroxy-6,7,8,4',5'-pentamethoxyflavone (gardenin C), 6,7,8,3',4',5'-hexamethoxyflavone, 5-hydroxy-6,7,8,3',4'-pentamethoxyflavone (5-O-desmeth-yl-nobiletin), 5,3'-dihydroxy-6,7,4',5'-tetramethoxyflavone, and 5,3',5'-trihydroxy-6,7,4'-trimethoxyflavone, were found in CHCl₃ extracts of MP leaves [16]. Two flavonoids, 5,6,7,8,3',4',5'-heptamethoxyflavone and 3,5,7,8,3',4',5'-heptamethoxyflavone, were isolated from MP leaf extracts and showed inhibitory activity against human carbonic anhydrase isozyme II (hCAII) at doses of 10.8 and 21.5 M, respectively [17]. Moreover, leaf and shoot extracts contain two glycosides of flavone methyl ethers (5,8,3'-trihydroxy-6,7,4'-trimethoxy flavone 8-O-beta glucopyranoside and 5,8-dihydroxy-6,7,3',4'-tetramethoxy flavone 8-O—beta-glucopyranoside [18]. Polymethoxylated flavonoids were withdrawn from the leaf extracts, and interpreted by the HPLC-DAD-ESI-MS/MS analytical method [19]. A total of 14 secondary metabolites showed abundance in the leaf and twig extracts [20]. Alanditrypinnone, alantryphenone, alantrypinene-B, and alantrylwnone are spiroquinazoline alkaloids isolated from *Eupenicillium* spp. leaves [21]. 2-O-ethylmurrangatin, a secondary leaf metabolite, inhibited lipoxygenase and had moderate respiratory burst inhibitory activity [22].

2. Root

MP root extracts contain indole alkaloid derivatives, such as paniculidines (D-F) with six known analogs, and the HRESIMS, UV, IR, and NMR spectroscopic techniques were used to elucidate the compounds [23]. Coumarins derivatives named panitins A-G with 34 known analogs were also identified [24].

Acetone extraction of root bark extracts have shown the levels of coumarins like minumicrolin isovalerate, murralonginol isovalerate, murrangatin isovalerate, chloculol, and an indole alkaloid called paniculol. The structures of coumarins were identified on the basis of ^1H NMR (270 MHz) spectroscopic method [25]. Yuehchukene, a new dimeric indole alkaloid produced from root extracts, has also been identified and displays anti-implantation activity [26].

3. Flowers

Coumarins like (-)-murracarpin, omphalocarpin, murrayacarpin-A and -B along with known coumarins like scopoletin, scopolin, murracarpin, 5,7-dimethoxy-S-(3'-methyl-2'-oxobutyl)coumarin, mupanidin, 3,5,7,3',4',5'-hexamethoxytlavone (a flavonoid), and murrayaculatine (an indole alkaloid) were extracted from flower extracts [27, 28].

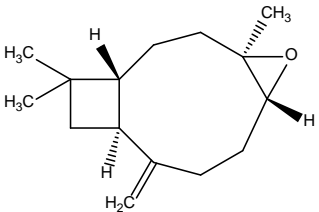
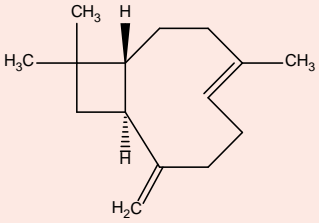
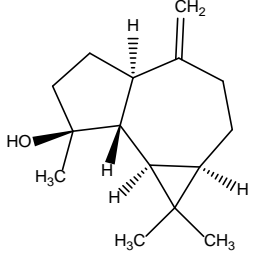
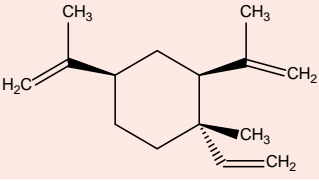
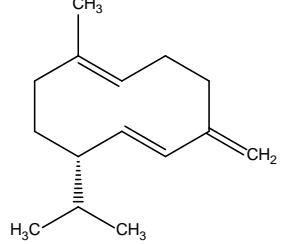
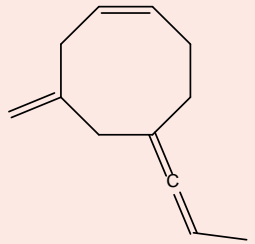
4. Aerial parts

The aerial parts of MP contain coumarins like murrmeranzin, murralonginal, minumicrolin, murrangatin, meranzin hydrate, and hainanmurpanin. Minumicrolin compounds were discovered to have anti-cholinesterase actions [29]. GS & GS-MS were used to identify the 48 categories of volatile compounds found in ethanolic extracts [30].

5. Fruits

MP fruits were used to isolate a water-soluble gum polysaccharide. The polysaccharide was highly branched using hydrolytic assays, methylation analyses, periodate oxidation research and NMR data [31]. Essential oils from the fruit contain α -copaene, -zingiberene, β -caryophyllene, germacrene D, and α -humulene [32]. A chemical component called paniculacin has been identified from an ethanolic MP extract in the form of a colorless oil [4]. The 58 identified components were validated through phytochemical investigation of the essential oil. Caryophyllene oxide, -elemene, -caryophyllene, spathulenol, germacrene D, cyclo-octene, and 4-methylene-6-(1-propenylidene) were the main compounds (Table 2) [33].

Table 2. Major phytoconstituents of MP leave oil [33]

S.No.	Chemical Structure with IUPAC name	Chemical constituent
1.		Caryophyllene oxide
2.		Beta-caryophyllene
3.		Spathulenol
4.		Beta-elemene
5.		Germacrene-D
6.		Cyclotene, 4-methylene-6-(1-propenylidene)

PHARMACOLOGICAL ACTIVITIES

1. Analgesic activity

The analgesic activity of MP bark extracts was evaluated on

Swiss albino rats, which were elicited from doses of 200 and 400 mg/kg body weight [34]. Other studies found that leaf extracts exhibited antinociceptive activities in rats and mice [35, 36].

2. Anti-diarrheal, bronchodilator, and vasodilator activity

In rabbit tissue preparations, the aqueous ethanolic extracts of MP leaves displayed calcium channel blocking actions, which is effective for treating diarrhea. The spasmolytic action of the extract was discovered at a dose of 0.01-0.3 mg/mL, with an EC₅₀ of 0.03610 mg/mL for anti-diarrheal activity. The leaf extract also exhibited bronchodilator and vasodilator activity in rabbits from utilizing isolated tissue preparations [37]. In another study, when the ethanolic MP extract was compared to castor oil, it showed potential anti-diarrheal activity, with a significant reduction in the incidence and severity of diarrhea in an experimental mice model [38].

3. Anti-inflammatory activity

The three types of flavonoids extracted from MP were 5,7,3',4',5'-pentamethoxyflavone (P1), 5,7,3',4'-tetramethoxyflavone (P3), and 5-hydroxy-6,7,8,3',4'-pentamethoxyflavone (P8), which possesses anti-inflammatory action upon nitric oxide inhibition at the dose of 3 μM. Ethanolic leaf extracts exhibited anti-inflammatory activity, which was evaluated by using different anti-inflammatory screening models [35, 38, 39]. In one study, the anti-inflammatory activity of total flavonoids of MP (TFMP) was investigated on high glucose-induced H9c2 cells. TFMP exhibited various inhibitory activities on oxidative stress, inflammation, and apoptosis [40].

4. Antioxidant activity

Utilizing DPPH scavenging and other techniques, the antioxidant activity of a methanolic extract of leaves was ascertained. The antioxidant capacity of MP methanol extract was shown to be greater than that of the standard antioxidant (trolox) [41]. Another study looked at the antioxidant activity of MP extracts in water, ethanol, and hexane. According to the data, ethanol extracts at 500 g/mL had a 67.77% antioxidant activity when compared to the standard reference (500 g/mL of alpha tocopherol), which had a 72.24% antioxidant activity [42].

5. Antibacterial activity

The MP leaf extracts exhibited an antibacterial activity on Gram-positive and Gram-negative through the disc diffusion and micro-dilution methods. The extracts contained phenols and flavonoids in high amounts, which contribute to the antibacterial activity [43]. There is notable inhibition of growth of all bacterial strains [44]. Another study found that the flower extract showed a zone of inhibition of bacteria which was measured by high-media scales [45]. An essential oil of MP also possessed antibacterial properties [46]. One study found that ethanolic leaf extract exhibited antibacterial activity against extended-spectrum β-lactamase *Klebsiella pneumoniae*, which causes nosocomial infections and is resistant to beta-lactam antibiotics [47].

6. Anticancer activity

The isolated sterol from MP leaves exhibited antitumor activity against cancer cell lines. The cytotoxic actions of sterols were determined by an MTT assay [48]. In one investigation, the cytotoxic effect of ethyl acetate extracts of MP leaves was investigated on human gingival fibroblasts and monocytes [49]. Different bark extracts according to solvents were tested using a brine shrimp lethality bioassay [50]. In a different study, a flavonoid glycoside isolated from MP twigs was able to inhibit adherence, movement, and invasion of lung adenocarcinoma A549 cells [51].

7. Antifungal activity

Ethanolic and aqueous MP extracts possess antifungal activity against the *Trichophyton rubrum* [45]. One recent study suggested that MP leaf extracts can potentially inhibit fungal growth [52].

8. Anthelmintic activity

In vivo anthelmintic activity was dramatically increased after MP leaves were fed in one study, indicating that gastrointestinal nematodes, growth rates, and hematological abnormalities in goats were reduced [53]. In another study, MP leaf extracts showed anthelmintic activity against *Tricho strongylus* sp., *Haemonchus* sp., and *Cooperia* sp., and the infusion of 7% leaf extracts reduced larval development, infective larvae and adult

trichostronglidae the most effectively [54].

9. Antianxiety and antidepressant activity

The anti-anxiety and anti-depressant properties of several solvent extracts of MP leaf were also recorded. The extract boosted the number of animals entering the anti-anxiety model and decreased immobility in mice in the anti-depression model, according to previous findings [55].

10. Gastroprotective and renoprotective quality

The ethanol extract of MP significantly inhibited ethanol HCl induced gastric lesions and decrease the levels of hormones and cytokines, such as TNF- α , IL-6, IL-1 β , MTL and GAS at the high dose. The result suggested that MP protected the gastric mucosa by the expansion of inflammation, and preventing the ethanol-HCl-induced necrosis and apoptosis [56]. Another study found that the extracted total flavonoids from the ethanol extract of the leaves had a protective effect on the kidneys in rats that were hyperlipidemic and had diabetes that had been induced by streptozotocin [57].

11. Anti-obesity activity

Pancreatic lipase activity was inhibited by the ethanolic and aqueous extract of MP leaves, indicating anti-obesity activity [58].

12. Anti-hyperglycemic activity

The anti-diabetic action of hydro alcoholic extract of MP leaves was studied in streptozotocin induced diabetic rats. Reduced blood glucose levels in diabetic rats, which varied depending on the dose, were used to corroborate this effect. The findings revealed that 400 mg/kg of extract was equal to the reference dose of glibenclamide [59]. Another study used an alloxan-induced diabetes model to scrutinize the effects of MPE on blood glucose levels in diabetic and non-diabetic rats. After 14 days of treatment, glucose levels in diabetic rats had decreased. After 21 days of therapy, non-diabetic rats displayed a reduction in blood glucose levels [60]. A hydro-alcoholic extract was given to alloxan-induced diabetic rats at doses of 100, 200, and 400 mg/kg for 60 days. Glycemic, cholesterol, and triglyceride levels were all reduced by the extract. Diabetes-induced mor-

phological changes in the liver, pancreas, and kidney were also decreased. Fructosamine and glycated hemoglobin levels were reduced as well [61].

13. Toxicity studies

The plant was traditionally utilized as a folk medicine. There were no deaths or CNS or ANS toxicity after an acute oral administration of the extract (2,000 mg/kg and 5,000 mg/kg single dose) [62, 63]. In rats, subacute oral administration (100, 200, or 400 mg/kg for 28 days) revealed no effects on body weight, food intake, or water intake [63].

CONCLUSION

MP holds considerable therapeutic potential for conventional and pharmacological management of diseases. Significant research has not yet been conducted on phytoconstituents from MP extracts. Identified phytochemicals should also be subjected to pharmacological investigation to shed light on the molecular processes of these unreported secondary metabolites for the protection human and animal populations from diseases and other health issues. Literature showed that methanol and hydroalcoholic extracts of MP had shown high pharmacological activity. Moreover, a large body of pharmacological research has demonstrated that MP is useful against cancer, diabetes, hyperlipidemia, infections, free radicals, and other diseases. Additionally, no harmful effects have been established by pre-clinical data. More investigations into the pharmacological and toxicological characteristics of the phytochemicals in MP are required to completely comprehend them. This review suggests further pharmacological investigation into MP to ascertain its therapeutic potential against particular deadly illnesses. Future studies are also required to assess its pharmacological properties, including its toxicity to plants, environmental effects, and other potential uses. Additionally, we anticipate that this review will benefit researchers for their further studies.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest, financial or otherwise.

FUNDING

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

ORCID

Deepa Joshi, <https://orcid.org/0000-0003-2990-1040>

Kashmira J. Gohil, <https://orcid.org/0000-0003-4142-3379>

REFERENCES

- Gohil KJ, Patel JA. A review on *Bacopa monniera*: current research and future prospects. *Int J Green Pharm.* 2010;4(1):1-9.
- Gohil KJ, Patel JA, Gajjar AK. Pharmacological review on *Centella asiatica*: a potential herbal cure-all. *Indian J Pharm Sci.* 2010;72(5):546-56.
- Gautam MK, Goel RK. Exploration of preliminary phytochemical studies of leaves of *Murraya paniculata* (L.). *Int J Pharm Life Sci.* 2012;3(8):1871-4.
- Saeed S, Shah S, Mehmood R, Malik A. Paniculacin, a new coumarin derivative from *Murraya paniculata*. *J Asian Nat Prod Res.* 2011;13(8):724-7.
- Tomaseto AF, Marques RN, Fereres A, Zanardi OZ, Volpe HXL, Alquézar B, et al. Orange jasmine as a trap crop to control *Diphorina citri*. *Sci Rep.* 2019;9(1):2070.
- Kong YC, Ng GKH, Wat CKH, But PPH. Pharmacognostic differentiation between *Murraya paniculata* (L.) Jack and *Murraya koenigii* (L.) Spreng. *Int J Crude Drug Res.* 1986;24(3):167-70.
- Faisal M, Sarker MMH, Rahman A, Hossain AI, Rahman S, Bashar ABMA, et al. *Murraya paniculata* (L.) Jack: a potential plant for treatment of toothache. *J Dent Oral Disord Ther.* 2014; 2(3):1-3.
- Gilman EF. Fact Sheet FPS-416: *Murraya paniculata*. Gainesville: University of Florida; 1999.
- Wardani E, Harahap Y, Mun'im A, Bahtiar A. Influence of extraction on the yield, phytochemical and LCMS profile from standardized kemuning leaf (*Murraya paniculata* (L.) Jack). *Pharmacogn J.* 2019;11(6 Suppl):1455-62.
- Wilson A, Kuchlmayr B, Moon C, McCusker A, Zhang X. *Flora of Australia* volume 26: Meliaceae, Rutaceae, Zygophyllaceae. Melbourne: ABRIS, Canberra/CSIRO Publishing; 2013. pp. 502-3.
- Liu H, Zhao Y, Zhou J, Ma Q, Wang X, Hua Z. Complete chloroplast genome sequence of *Murraya paniculata* (Rutaceae): a widely used folk medicinal herb. *Mitochondrial DNA B Resour.* 2020;5(3):3696-7.
- Ng MK, Abdulhadi-Noaman Y, Cheah YK, Yeap SK, Alitheen NB. Bioactivity studies and chemical constituents of *Murraya paniculata* (Linn) Jack. *Int Food Res J.* 2012;19(4):1307-12.
- Sayar K, Paydar M, Pinguan-Murphy B. Pharmacological properties and chemical constituents of *Murraya paniculata* (L.) Jack. *Med Aromat Plants.* 2014;3(4):1000173.
- Monir TSB, Afroz S, Jahan I, Hossain T. Phytochemical study and antioxidant properties of aqueous extracts of *Murraya paniculata* Leaf. *J Appl Life Sci Int.* 2020;23(4):1-8.
- Wu TS. Coumarins from the leaves of *Murraya paniculata*. *Phytochemistry.* 1988;27(7):2357-8.
- Kinoshita T, Firman K. Highly oxygenated flavonoids from *Murraya paniculata*. *Phytochemistry.* 1996;42(4):1207-10.
- Sangkaew A, Samritsakulchai N, Sanachai K, Rungrotmongkol T, Chavasiri W, Yompakdee C. Two flavonoid-based compounds from *Murraya paniculata* as novel human carbonic anhydrase isozyme II inhibitors detected by a resazurin yeast-based assay. *J Microbiol Biotechnol.* 2020;30(4):552-60.
- Zhang Y, Li J, Shi S, Zan K, Tu P. Glycosides of flavone methyl ethers from *Murraya paniculata*. *Biochem Syst Ecol.* 2012;43:10-3.
- Zhang JY, Li N, Che YY, Zhang Y, Liang SX, Zhao MB, et al. Characterization of seventy polymethoxylated flavonoids (PMFs) in the leaves of *Murraya paniculata* by on-line high-performance liquid chromatography coupled to photodiode array detection and electrospray tandem mass spectrometry. *J Pharm Biomed Anal.* 2011;56(5):950-61.
- Liang H, Zhao M, Tu P, Jiang Y. Polymethoxylated flavonoids from *Murraya paniculata* (L.) Jack. *Biochem Syst Ecol.* 2020;93:104162.
- Barros FAP, Rodrigues-Filho E. Four spiroquinazoline alkaloids from *Eupenicillium* sp. isolated as an endophytic fungus from leaves of *Murraya paniculata* (Rutaceae). *Biochem Syst Ecol.* 2005;33(3):257-68.
- Shaikh A, Choudhary MI. Bioassay studies of 2'-O-ethylmurrangatin isolated from a medicinal plant, *Murraya paniculata*. *Turk J Biol.* 2011;35(6):751-5.
- Wang XT, Zeng KW, Zhao MB, Tu PF, Li J, Jiang Y. Three new indole alkaloid derivatives from the roots of *Murraya paniculata*. *J Asian Nat Prod Res.* 2018;20(3):201-8.
- Wang X, Liang H, Zeng K, Zhao M, Tu P, Li J, et al. Panitins A-G: coumarin derivatives from *Murraya paniculata* from Guangxi Province, China show variable NO inhibitory activity. *Phytochemistry.* 2019;162:224-31.
- Ito C, Furukawa H, Ishii H, Ishikawa T, Haginiwa J. The chemical composition of *Murraya paniculata*. The structure of five new coumarins and one new alkaloid and the stereochemistry of murrangatin and related coumarins. *J Chem Soc Perkin Trans 1.*

- 1990;7:2047-55.
26. Kong YC, Ng KH, Wat KH, Wong A, Saxena IF, Cheng KF, et al. Yuehchukene, a novel anti-implantation indole alkaloid from *Murraya paniculata*. *Planta Med.* 1985;51(4):304-7.
 27. Wu TS, Liou MJ, Kuoh CS. Coumarins of the flowers of *Murraya paniculata*. *Phytochemistry.* 1989;28(1):293-4.
 28. Wu TS, Chan YY, Leu YL, Huang SC. A flavonoid and indole alkaloid from flowers of *Murraya paniculata*. *Phytochemistry.* 1994;37(1):287-8.
 29. Saied S, Nizami SS, Anis I. Two new coumarins from *Murraya paniculata*. *J Asian Nat Prod Res.* 2008;10(5-6):515-9.
 30. Shah S, Saied S, Mahmood A, Malik A. Phytochemical screening of volatile constituents from aerial parts of *Murraya paniculata*. *Pak J Bot.* 2014;46(6):2051-6.
 31. Mondal SK, Ray B, Ghosal PK, Teleman A, Vuorinen T. Structural features of a water soluble gum polysaccharide from *Murraya paniculata* fruits. *Int J Biol Macromol.* 2001;29(3):169-74.
 32. Olawore NO, Ogunwande IA, Ekundayo O, Adeleke KA. Chemical composition of the leaf and fruit essential oils of *Murraya paniculata* (L.) Jack. (Syn. *Murraya exotica* Linn.). *Flavour Fragr J.* 2005;20(1):54-5.
 33. Chowdhury JU, Bhuiyan MNI, Yusuf M. Chemical composition of the leaf essential oils of *Murraya koenigii* (L.) Spreng and *Murraya paniculata* (L.) Jack. *Bangladesh J Pharm.* 2008;3(2):59-63.
 34. Podder MK, Das BN, Saha A, Ahmed M. Analgesic activity of bark of *Murraya paniculata*. *Int J Med Med Sci.* 2011;3(4):105-8.
 35. Narkhede MB, Ajmire PV, Wagh AE. Evaluation of antinociceptive and anti-inflammatory activity of ethanol extract of *Murraya paniculata* leaves in experimental rodents. *Int J Pharm Pharm Sci.* 2012;4(1):247-50.
 36. Sharker SM, Shahid IJ, Hasanuzzaman M. Antinociceptive and bioactivity of leaves of *Murraya paniculata* (L.) Jack, Rutaceae. *Rev Bras Farmacogn.* 2009;19(3):746-8.
 37. Saqib F, Ahmed MG, Janbaz KH, Dewanjee S, Jaafar HZ, Zia-Ul-Haq M. Validation of ethnopharmacological uses of *Murraya paniculata* in disorders of diarrhea, asthma and hypertension. *BMC Complement Altern Med.* 2015;15:319.
 38. Rahman MA, Hasanuzzaman M, Uddin N, Shahid I. Antidiarrhoeal and anti-inflammatory activities of *Murraya paniculata* (L.) Jack. *Pharmacologyonline.* 2010;3:768-76.
 39. Wu J, Liu K, Shi X. The anti-inflammatory activity of several flavonoids isolated from *Murraya paniculata* on murine macrophage cell line and gastric epithelial cell (GES-1). *Pharm Biol.* 2016;54(5):868-81.
 40. Zou J, Sui D, Fu W, Li Y, Yu P, Yu X, et al. Total flavonoids extracted from the leaves of *Murraya paniculata* (L.) Jack alleviate oxidative stress, inflammation and apoptosis in a rat model of diabetic cardiomyopathy. *J Funct Foods.* 2021;76:104319.
 41. Zhu CH, Lei ZL, Luo YP. Studies on antioxidative activities of methanol extract from *Murraya paniculata*. *Food Sci Hum Wellness.* 2015;4(3):108-14.
 42. Sundaram M, Sivakumar, Karthikeyan, Bhuvaneshwari, Aishwarya, Thirumalai, et al. Studies on in vitro antibacterial, antifungal property and antioxidant potency of *Murraya paniculata*. *Pak J Nutr.* 2011;10(10):925-9.
 43. Gautam MK, Gangwar M, Nath G, Rao CV, Goel RK. In-vitro antibacterial activity on human pathogens and total phenolic, flavonoid contents of *Murraya paniculata* Linn. leaves. *Asian Pac J Trop Biomed.* 2012;2(3 Suppl):S1660-3.
 44. Sonter S, Mishra S, Dwivedi MK, Singh PK. Chemical profiling, in vitro antioxidant, membrane stabilizing and antimicrobial properties of wild growing *Murraya paniculata* from Amarkantak (M.P.). *Sci Rep.* 2021;11(1):9691.
 45. Punasiya R, Dindorkar G, Pillai S. Antibacterial and antifungal activity of flower extract of *Murraya paniculata* L. *Asian J Res Pharm Sci.* 2020;10(1):17-20.
 46. Dosoky NS, Satyal P, Gautam TP, Setzer WN. Composition and biological activities of *Murraya paniculata* (L.) Jack essential oil from Nepal. *Medicines (Basel).* 2016;3(1):7.
 47. Wiyogo IO, Endraswari PD, Setiawati Y. Antibacterial activity of ethanol extract of Kemuning (*Murraya Paniculata*) against *Klebsiella pneumoniae* ESBL by in vitro test. *Indones J Trop Infect Dis.* 2021;9(2):101-6.
 48. Baker DHA, Ibrahim EA, Kandeil A, Baz FKE. Sterols bioactivity of *Ruta graveolens* L. and *Murraya paniculata* L. *Int J Pharm Pharm Sci.* 2017;9(2):103-8.
 49. Rodanant P, Khetkam P, Suksamrarn A, Kuvatanasuchati J. Coumarins and flavonoid from *Murraya paniculata* (L.) Jack: antibacterial and anti-inflammation activity. *Pak J Pharm Sci.* 2015;28(6):1947-51.
 50. Rumzhum NN, Rahman MM, Hossain MF. Free radical scavenging activity and cytotoxic potential of crude extractives of *Murraya paniculata* (L.) bark. *Res J Pharmacogn Phytochem.* 2012;4(1):18-22.
 51. Shi Q, Jiang Z, Yang J, Cheng Y, Pang Y, Zheng N, et al. A flavonoid glycoside compound from *Murraya paniculata* (L.) interrupts metastatic characteristics of A549 cells by regulating STAT3/NF- κ B/COX-2 and EGFR signaling pathways. *AAPS J.* 2017;19(6):1779-90.
 52. Nesa M, Hosen ME, Khan MAI, Kabir MH, Zaman R. In-vitro antifungal activity of *Azadirachta indica*, *Ocimum tenuiflorum* & *Murraya paniculata* leaf extract against three phytopathogenic fungi. *Am J Pure Appl Sci.* 2021;3(5):113-8.
 53. Boonkusol D, Detraksa J, Duangsrikaew K, Tongbai W. In vivo efficacy of *Murraya paniculata* leaf in controlling natural hel-

- minthiasis in goat. *Am J Anim Vet Sci*. 2019;14(2):95-100.
54. Tresia GE, Evvyernie D, Tiuria R. Phytochemical screening and in vitro ovicidal, larvacidal, and nematicidal effects of *Murraya paniculata* (L.) Jack extract on gastrointestinal parasites of goats. *Media Peternak*. 2016;39(3):173-9.
55. Sharma P, Batra S, Kumar A, Sharma A. In vivo antianxiety and antidepressant activity of *Murraya paniculata* leaf extracts. *J Integr Med*. 2017;15(4):320-5.
56. Lu M, Du Z, Yuan S, Ma Q, Han Z, Tu P, et al. Comparison of the preventive effects of *Murraya exotica* and *Murraya paniculata* on alcohol-induced gastric lesions by pharmacodynamics and metabolomics. *J Ethnopharmacol*. 2021;281:114567.
57. Zou J, Yu X, Qu S, Li X, Jin Y, Sui D. Protective effect of total flavonoids extracted from the leaves of *Murraya paniculata* (L.) Jack on diabetic nephropathy in rats. *Food Chem Toxicol*. 2014; 64:231-7.
58. Iswantini D, Silitonga RF, Martatilofa E, Darusman LK. Zingiber cassumunar, *Guazuma ulmifolia*, and *Murraya paniculata* extracts as antiobesity: in vitro inhibitory effect on pancreatic lipase activity. *HAYATI J Biosci*. 2011;18(1):6-10.
59. Gautam MK, Gupta A, Vijaykumar M, Rao CV, Goel RK. Studies on the hypoglycemic effects of *Murraya paniculata* Linn. extract on alloxan-induced oxidative stress in diabetic and non-diabetic models. *Asian Pac J Trop Dis*. 2012;2 Suppl 1:S186-91.
60. Gautam M, Gupta A, Rao CV, Goel R. Antihyperglycemic and antioxidant potential of *murraya paniculata* linn. Leaves: a pre-clinical study. *J Pharm Res*. 2012;5(3):1334-7.
61. Menezes CDA, de Oliveira Garcia FA, de Barros Viana GS, Pinheiro PG, Felipe CFB, de Albuquerque TR, et al. *Murraya paniculata* (L.) (Orange Jasmine): potential nutraceuticals with ameliorative effect in alloxan-induced diabetic rats. *Phytother Res*. 2017;31(11):1747-56.
62. Gautam MK, Singh A, Rao CV, Goel RK. Toxicological evaluation of *Murraya paniculata* (L.) leaves extract on rodents. *Am J Pharmacol Toxicol*. 2012;7(2):62-7.
63. Menezes IR, Santana TI, Varela VJ, Saraiva RA, Matias EF, Boli-gon AA, et al. Chemical composition and evaluation of acute toxicological, antimicrobial and modulatory resistance of the extract of *Murraya paniculata*. *Pharm Biol*. 2015;53(2):185-91.