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Total polyphenol and ferulic acid analysis of a new variety of corn, Bandiburichodang, according to steaming time and roasting temperature

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Abstract Bandiburichodang (BDC) is a new variety of Zea mays L. Total polyphenol content (TPC) assay and quantitative analysis of ferulic acid (FA) were performed to determine the steaming, roasting conditions of BDC kernels that lead to the highest content. TPC levels increased after roasting under all conditions. TPC levels in samples steamed at 115 °C for 25 min were 3.157 mg/g before roasted, and increased to 3.825 and 4.739 mg/g after roasting at 160 and 200 °C, respectively. Whether BDC kernels were roasted was relevant with TPC content. BDC kernels were extracted to perform quantitative analysis of FA. Roasting temperature affected FA content: the higher the temperature, the lower the content. BDC kernels that were steamed at 115 °C for 25 min had 0.178 mg/g of FA content before roasting, and levels decreased to 0.132 and 0.115 mg/g after roasting. Under different roasting conditions, FA content decreased 15 to 50%. We hypothesize that this phenomenon is due to a breakdown of phenolic compounds or cell wall disruption.

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Keywords Bandiburichodang · Ferulic acid · High-performance liquid chromatography/Photodiode array detector · Roasting · Total polyphenol content · *Zea mays*

Introduction

Zea mays L., otherwise known as maize, is a crop that is cultivated worldwide. Z. mays is a staple food in many continents including Africa, Asia, and Latin America [1]. Approximately 60% of the global yield of Z. mays is produced by USA and China [2]. Z. mays is beloved all over the world for its rich carbohydrate content and its numerous uses in animal feeds [3]. It is an excellent source of bioactive compounds, such as polyphenols, phenolic acids, flavonoids, anthocyanins, and carotenoids [4].

Ferulic acid (FA) is a phenolic compound in *Z. mays* kernels [5]. It is found in the seeds and leaves of numerous plants [6]. FA was isolated from *Ferula foetida*, where its name originated. FA is known for its various therapeutic activities: antioxidative effects against diabetes, cancer, and other diseases and for its superior cytoprotective ability [7,8]. In fact, FA is frequently a component of Chinese phytomedicine, and awareness of FA is high in China because of this connection [9]. In addition to anticancer, antiproliferative, and antibacterial effects, FA has also been observed to promote wound healing in studies with diabetic rats. Wounds treated with FA re-epithelialized in a shorter time, compared with untreated wounds [10]. Studies have shown that *Z. mays* has a high FA content. A study by Adom and Liu (2002), showed that corn had higher free FA content than other grains such as rice, oats, and wheat [11].

Bandiburichodang (BDC), a new variety of *Z. mays*, was first developed by Mr. Gunhwa Park, Agricultural Corporation Company Maru, Pyeongtaek, Korea. BDC is a yellow chodang, or super sweet corn, species of *Z. mays* [12]. Super sweet corn is often harvested as green corn and used as a snack food [13].

detected well in the new variety. The existence of a correlation between how BDC kernels are pretreated (steaming time, steaming temperature, and roasting temperature) and FA contents of the kernels has been suggested.

This study hypothesized that in addition to steaming, the roasting of BDC kernels might influence the phenolic content. Hence, a quantitative analysis of FA by high-performance liquid chromatography (HPLC) and total polyphenol content (TPC) assays with BDC kernel samples was performed.

Materials and Methods

Plant materials

BDC (National Seed Resource Variety Protection No. 5008) was cultivated by Mr. Gunhwa Park, Agricultural Corporation Company Maru, Pyeongtaek, Korea (Fig. 1). Kernel samples were made using different steaming and roasting conditions. BDC plants were grown according to the instructions of the Rural Development Administration, Korea. Mr. Gunhwa Park modified the spacing of individual plants, such as 15 plants per square foot with distances of 30 cm between stems and 70 cm between rows.

Instruments and reagents

Quantitative analysis was performed using the Waters Alliance 2695 Separations Module (USA), and Water 996 Photodiode Array detector (USA). INNO C_{18} columns (4.6×250 mm, 5 µm) for high-performance liquid chromatography (HPLC) were purchased from Youngjin Biochrom Co., Korea, and acetic acid (glacial, 100%) from Sigma-Aldrich, Germany. HPLC-grade water, methanol (MeOH), and acetonitrile (ACN) were obtained from J. T. Baker (Phillipsburg, PA, USA). Acetic acid (AA), tannic acid (TA) and FA (Fig. 2) were provided by Natural Product Institute of Science and Technology (www.nist.re.kr), Anseong, Korea.

Extraction of BDC kernels

BDC kernels were extracted with ethanol (EtOH) using a Soxhlet



Fig. 1 Image of BDC kernels

reflux evaporator. Ten grams of dried BDC kernel samples were measured and then ground into fine powder before extraction with 300 mL for 3 h. Each experiment was performed in triplicate. Dehydrated extracts were collected after evaporation with a rotary vacuum evaporator.

Total polyphenol content (TPC)

One gram of each sample was dissolved in 10 mL distilled water, and sonicated for 30 min. After centrifugation at 4,000 rpm for 10 min, the supernatant liquid was collected and filtered through 0.45 µm polyvinylidene fluoride (PVDF) membrane for testing. TA was used as a standard of the TPC assay. One milligram of TA was dissolved in 1 mL distilled water and sequentially diluted for testing. TPC was measured by adding 7.5 mL distilled water to each test tube, followed by 1 mL each standard and sample solutions, 0.5 mL of Folin-Denis reagent, and 1 mL 35% sodium carbonate. Samples were incubated in a dark room for 1 h and then the absorbance at 760 nm was measured with a microplate reader.

Preparation of standard and sample solutions for HPLC

BDC extracts (20 mg) were dissolved in 1mL of MeOH (20 mg/ mL), and sonicated. FA (1 mg) were dissolved in 1 mL MeOH, and then sequentially diluted to produce a calibration curve. Both the standard and the sample solutions was filtered through 0.2 μ m PVDF membrane filter.

HPLC conditions

A wavelength of detector was set to 324 nm. The mobile phase consisted of 0.3% AA in water (A) and ACN (B). The gradient conditions were as follows; 90% of A at 0 min, 70% of A at 20 min, 50% of A at 25 min, 0% of A at 30 min, and 90% of A at 40 min. The injection volume was set to 10 μ L, flow rate to 1 mL/ min, and column temperature to 35 °C.

Calibration curve

When forming the calibration curve, the value of the X axis ($\mu g/mL$) signifies the concentration of FA, and the Y axis value (mAU) indicates the area of FA (Fig. 3). To generate a calibration curve, seven FA solutions with different concentrations (100-1.56



Fig. 2 Chemical structure of FA



Fig. 3 HPLC chromatogram of FA

Table 1	Calibration	curve	of FA
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Compound	t _R	Calibration equation ^a	Correlation factor, r^{2b}
FA	18.5	Y = 44170 X - 13156	0.9998

^aY = peak area, X = concentration of standards (μ g/mL)

 ${}^{b}r^{2}$ = correlation coefficient based on three data points in the calibration curves

ppm) and associated peak areas (4420304-75203 mAU). The total FA content (mg/g) was calculated by multiplying C, V, D, P and dividing by W (C: concentration of standard, V: total volume of the test solution, D: dilution factor, P: standard purity, W: sample weight.). The calibration curve of FA showed good linearity, with the correlation factor (r^2) of 0.9998 (Table 1).

Results and Discussion

Z. mays is used in diverse food items, such as chips, tortillas, and bread [15]. Getting a healthy dose of functional food has become a concern for consumers [16]. Our goal was to identify treatment conditions that isolate the highest quantity of functional phenolic compounds from BDC kernels. Therefore, we evaluated different steaming and roasting conditions and measured the yields of TPC. After extraction with EtOH, TPC assays on each sample and quantitative analysis using HPLC to measure content of FA, a marker component of BDC kernels, were performed.

We found that TPC increased after roasting. One proposed reason is that roasting improves extraction efficiency. TPC levels increased after roasting under all conditions. TPC levels in samples steamed at 115°C for 25 min were 3.157 mg/g before roasted and increased to 3.825 and 4.739 mg/g after roasting at 160 and 200 °C, respectively, and TPC levels in samples steamed at a same temperature for 50 min were 2.916 mg/g before roasted

and increased to 6.535 and 4.834 mg/g after roasting at 160 and 200 °C, respectively. When steamed at 121 °C for 25 min, TPC levels were 3.342 mg/g before roasted and elevated to 4.375 and 5.399 mg/g after roasting at 160 and 200 °C, respectively, and for samples steamed at a same temperature for 50 min were 3.918 mg/g before roasted and increased to 8.336 and 7.574 mg/g after roasting at 160 and 200 °C, respectively (Table 2). Boateng et al. (2008) explained that the heating process improves the extractability of these substances by disrupting the cell wall and degrading insoluble phenolic compounds [17]. Thermal processing is studied to weaken the cell wall and cause unwanted color changes, because of the heat [18]. Kim et al. (2014) discovered that carrot cell walls that were treated with heat more than 5 min were broken and irregularly shaped [19]. However, in 50 min of steaming conditions, certain decrease in TPC levels has been observed when roasting temperature increased. We assumed this phenomenon was a result of extreme heating conditions. Yadev et al. (2012) found out that as steaming time increased, tannin content decreased [20].

Because there are many phenolic compounds, there are diverse functionalities. As secondary metabolites, phenolic compounds affect plant color, volume, and resistance against infectious agents [21]. In humans, they are associated with pharmaceutical functions, such as cancer suppression [22]. Phenolic compounds are anticipated to have utilizations in various industries, such as food and cosmetics. For example, phenolic compounds have been

Table 2 TPC in distilled water extracts of BDC kernels, according to steaming conditions and roasting temperature

Steaming time (min)	Steaming temperature		Roasting temperature	
		Raw (mg/g extract)	160 °C (mg/g extract)	200 °C (mg/g extract)
25	115 °C	3.157±0.199	3.825±0.403	4.739±0.148
	121 °C	3.342±0.136	4.375±0.173	5.399±0.027
50	115 °C	2.916±0.041	6.535±0.448	4.834±0.049
	121 °C	3.918±0.033	8.336±0.207	7.574±0.260



Fig. 4 HPLC chromatograms of 115 °C/25 M/Raw (A), 115 °C/25 M/160 °C (B), 121 °C/25 M/Raw (C) and 121 °C/25 M/160 °C (D): steaming temperature/steaming time/roasting temperature treatments of BDC kernels

added to processed foods to intensify antioxidative activities. Using phenolic compounds in the dyeing of fabric has been proposed to improve environmental friendliness [23].

Previous studies reported correlations between roasting and chemical compositions within plants. Jannat et al. (2013) found that TPC and γ -tocopherol content in Iranian sesame seeds increased after roasting. The reason proposed was that γ -

tocopherol disconnected from membrane proteins or phospholipids during roasting [24].

On the other hand, we found that roasted samples had a lower FA content relative to unroasted samples (Fig. 4). BDC kernels that were steamed at $115 \,^{\circ}$ C for 25 min had 0.178 mg/g FA content before roasting, and levels decreased to 0.132 and 0.115 mg/g after roasting. Under different roasting conditions, FA

Steaming time (min)	Steaming temperature		Roasting temperature	
		Raw (mg/g extract)	160 °C (mg/g extract)	200 °C (mg/g extract)
25	115 °C 121 °C	0.178±0.001 0.222±0.001	0.132±0.000 0.120±0.001	0.115±0.001 0.118±0.000
50	115 °C 121 °C	0.222±0.000 0.232±0.000	$\begin{array}{c} 0.190{\pm}0.001 \\ 0.103{\pm}0.000 \end{array}$	0.111±0.001 0.118±0.000

Table 3 Content of FA in EtOH extracts of BDC kernels, according to steaming time and roasting temperature

content decreased 15 to 50% (Table 3). There is an inverse relationship between roasting temperature and FA content. However, this case wasn't true for the kernels steamed at 121 °C for 50 min as FA content decreased when the roasting temperature increased from 180 to 200 °C. Therefore, the authors concluded that it is hard to find a definite relationship between roasting temperature and FA content since the samples had minute amount of FA.

According to an analysis of free phenolic compounds by Samaras et al. (2005), FA was shown to be degraded in heavily roasted malts [25]. Previous studies mention that FA has antiinflammatory and anticancer properties, with low toxicity [8]. An experiment on rats by Wang et al. (2018) suggested that FA helps reduce obesity and the symptoms associated with obesity; they discovered that FA-treated mice fed high-fat meals gained less weight and had substantial blood sugar increases than control mice [26].

FA is used in multiple packaging and cosmetics applications because of its antimicrobial and other effects [27]. Sharma et al. (2020) studied the impacts of FA on food films, such as antibacterial efficiency and temperature equilibrium, and proposed that FA could be used in food packaging [28]. Peres et al. (2018) performed *ex vivo* antioxidant, *in vivo* sun protection factor, and *in vitro* UVA protection factor assays and proposed that FA has a synergistic effect with UV filters in sunscreens [29]. Park et al. (2018) investigated the skin-whitening and anti-wrinkle activity of FA as a food additive [30].

We observed no significant relationship between the two steaming conditions and FA content in BDC kernels; however, we found that roasting increased the TPC. We propose that the increase in TPC is not related to FA; rather, the TPC increase is likely due to better extractability of phenolic compounds as a result of roasting. We hypothesize that additional studies of FA might reveal more industrial applications for FA.

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References

 Rosas-Castor JM, Guzmán-Mar JL, Hernández-Ramírez A, Garza-González MT, Hinojosa-Reyes L (2014) Arsenic accumulation in maize crop (Zea mays): a review. Sci Total Environ 488-489: 176-187. doi: 10.1016/j.scitotenv.2014.04.075

- Ghaffari A, Ali A, Tahir M, Waseem M, Ayub M, Iqbal A, Ullah Mohsin A (2011) Influence of integrated nutrients on growth, yield and quality of maize (*Zea mays* L.). Am J Plant Sci 2: 63–69. doi: 10.4236/AJPS. 2011.21009
- Tandzi LN, Mutengwa CS (2019) Estimation of maize (Zea mays L.) yield per harvest area: appropriate methods. Agron 10: 29. doi: 10.3390/ agronomy10010029
- Leandro da Cruz L, Moreno Bernardo Gonçalves G, de Lima Glória L, Menezes de Faria Pereira S, de Almeida Carlos L, Vivas M, Gonzaga Pereira M, Barros de Oliveira D (2022) Phenolic compounds, carotenoids, and antioxidant activity in a super-sweet corn hybrid. Pesqui Agropecu Bras 57: e02663. doi: 10.1590/S1678-3921.pab2022.v57. 02663
- Chudhangkura A, Teangpook C, Sikkhamondhol C, Jariyavattanavijit C (2018) Effects of ultraviolet C, controlled atmosphere, and ultrasound pretreatment on free ferulic acid in canned sweet corn kernels. J Food Sci Technol 55: 4167–4173. doi: 10.1007/s13197-018-3346-0
- Srinivasan M, Sudheer AR, Menon VP (2007) Ferulic acid: therapeutic potential through its antioxidant property. J Clin Biochem Nutr 40: 92– 100. doi: 10.3164/jcbn.40.92
- Kumar N, Pruthi V (2014) Potential applications of ferulic acid from natural sources. Biotechnol Rep 4: 86–93. doi: 10.1016/J.BTRE.2014. 09.002
- Mancuso C, Santangelo R (2014) Ferulic acid: pharmacological and toxicological aspects. Food Chem Toxicol 65: 185–195. doi: 10.1016/ J.FCT.2013.12.024
- Ou S, Kwok KC (2004) Ferulic acid: pharmaceutical functions, preparation and applications in foods. J Sci Food Agric 84: 1261–1269. doi: 10.1002/JSFA.1873
- Ghaisas MM, Kshirsagar SB, Sahane RS (2014) Evaluation of wound healing activity of ferulic acid in diabetic rats. Int Wound J 11: 523–532. doi: 10.1111/J.1742-481X.2012.01119.X
- Adom KK, Liu RH (2002) Antioxidant activity of grains. J Agric Food Chem 50: 6182–6187. doi: 10.1021/JF0205099
- Tracy WF (1997) History, genetics, and breeding of supersweet (shrunken2) sweet corn. In: Janick J (ed) Plant Breeding Reviews 14, John Wiley and Sons, Inc., pp 189–236. doi:10.1002/9780470650073. ch7
- Song EM, Kim HY, Lee SH, Woo SH, Kim HS, Kyung KS, Lee JS, Jeong HS (2011) Chemical components and quality characteristics of waxy corns cultured by conventional and environmentally-friendly methods. J Korean Soc Food Sci Nutr 40: 962–968. doi: 10.3746/jkfn. 2011.40.7.962
- Broberg M, Simaan H, Shmoish M, Rabner A, Karlsson M, Horwitz BA (2021) Ferulic acid, an abundant maize phenolic, regulates ABC and MFS transporter gene expression in the maize pathogen *Cochliobolus heterostrophus*. J Plant Dis Prot 128: 1383–1391. doi: 10.1007/S41348-021-00451-0
- Nuss ET, Tanumihardjo SA (2010) Maize: a paramount staple crop in the context of global nutrition. Compr Rev Food Sci Food Saf 9: 417–436. doi: 10.1111/J.1541-4337.2010.00117.X

- Sivam AS, Sun-Waterhouse D, Quek SY, Perera CO (2010) Properties of bread dough with added fiber polysaccharides and phenolic antioxidants: a review. J Food Sci 75: R163–R174. doi: 10.1111/J.1750-3841.2010. 01815.X
- Boateng J, Verghese M, Walker LT, Ogutu S (2008) Effect of processing on antioxidant contents in selected dry beans (*Phaseolus* spp. L.). LWT-Food Sci Technol 41: 1541–1547. doi: 10.1016/J.LWT.2007.11.025
- Lee JH (2009) Effects of steam-thermal processing on the food cooking quality. MS Thesis. Ewha Womans University, Seoul, Korea
- Kim KJ, Hwang IG, Yoo SM, Min SG, Choi MJ (2014) Effects of various pretreatment methods on physiochemical and nutritional properties of carrot. J Korean Soc Food Sci Nutri 43: 1881–1888. doi: 10.3746/jkfn.2014.43.12.1881
- Yadev DN, Kaur J, Anand T, Singh AK (2012) Storage stability and pasting properties of hydrothermally treated pearl millet flour. Int J Food Sci Technol 47: 2532–2537. doi: 10.1111/J.1365-2621.2012.03131.X
- Oksana S, Marian B, Mahendra R, Bo SH (2012) Plant phenolic compounds for food, pharmaceutical and cosmetics production. J Med Plants Res 6: 2526–2539. doi: 10.5897/JMPR11.1695
- 22. Minatel IO, Borges CV, Ferreira MI, Gomez HAG, Chen CYO, Lima GPP (2017) Phenolic compounds: functional properties, impact of processing and bioavailability. In: Hernadez MS, Tenango MP, Mateos MRG (eds) Phenolic Compounds: Biological Activity. Intech, pp 1–24. doi: 10.5772/66368
- Albuquerque BR, Heleno SA, Oliveira MBPP, Barros L, Ferreira ICFR (2021) Phenolic compounds: current industrial applications, limitations and future challenges. Food Funct 12: 14–29. doi: 10.1039/D0FO02324H

- 24. Jannat B, Reza Oveisi M, Sadeghi N, Hajimahmoodi M, Behzad M, Nahavandi B, Tehrani S, Sadeghi F, Oveisi M (2013) Effect of roasting process on total phenolic compounds and γ-tocopherol contents of Iranian sesame seeds (*Sesamum indicum*). Iran J Pharm Res 12: 751–758
- Samaras TS, Camburn PA, Chandra SX, Gordon MH, Ames JM (2005) Antioxidant properties of kilned and roasted malts. J Agric Food Chem 53: 8068–8074. doi: 10.1021/JF051410F
- Wang W, Pan Y, Zhou H, Wang L, Chen X, Song G, Liu J, Li A (2018) Ferulic acid suppresses obesity and obesity-related metabolic syndromes in high fat diet-induced obese C57BL/6J mice. Food Agric Immunol 29: 1116–1125. doi: 10.1080/09540105.2018.1516739
- Batista R (2014) Uses and potential applications of ferulic acid. In: Warren B (ed) Ferulic acid: antioxidant properties, uses and potential health benefits. Nova Science Publishers, pp. 39–70
- Shubham S, Amit KJ, Brendan D, Swarna J (2020) Ferulic acid incorporated active films based on poly(lactide)/poly(butylene adipateco-terephthalate) blend for food packaging. Food Packag Shelf Life 24: 100491. doi: 10.1016/j.fpsl.2020.100491
- Peres DDA, Sarruf FD, de Oliveira CA, Velasco MVR, Baby AR (2018) Ferulic acid photoprotective properties in association with UV filters: multifunctional sunscreen with improved SPF and UVA-PF. J Photochem Photobiol B Biol 185: 46–49. doi: 10.1016/j.jphotobiol.2018. 05.026
- Park HJ, Cho JH, Hong SH, Kim DH, Jung HY, Kang IK, Cho YJ (2018) Whitening and anti-wrinkle activities of ferulic acid isolated from *Tetragonia tetragonioides* in B16F10 melanoma and CCD-986sk fibroblast cells. J Nat Med 72: 127–135. doi: 10.1007/s11418-017-1120-7