# Occurrence of Gray Mold in Mango Caused by *Botrytis cinerea* in Korea

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During a disease survey conducted in April 2022, we observed severe gray mold symptoms on inflorescences of mango trees (*Mangifera indica*) grown in a vinyl greenhouse in Jeju, Korea. The symptoms occurred on the flowers and peduncles, on which a lot of gray molds were formed. The incidence of gray mold on the inflorescences in the vinyl greenhouse ranged from 10% to 40%. Three fungal isolates were obtained from the lesions and identified as *Botrytis cinerea* based on their morphological characteristics and phylogenetic analysis. All isolates of *B. cinerea* were tested for their pathogenicity to inflorescences of mango trees through artificial inoculation. The pathogenicity of the isolates was confirmed on the flowers and peduncles. The symptoms induced by the isolates were similar to those on the inflorescences of mango trees observed in the disease survey. This is the first report of *B. cinerea* causing gray mold in mango in Korea.

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Mango (*Mangifera indica*) is a tree belonging to the family Anacardiaceae, and its native is Assam, Belize, China South-Central, East Himalaya, Myanmar, and Thailand (Plants of the World Online, 2023). The tree grows primarily in the tropical and subtropical zones. In Korea, the tree has been mostly cultivated as a fruit tree in vinyl greenhouses in Jeju area. During a disease survey conducted in April 2022, we observed severe gray mold symptoms on inflorescences of mango trees grown in a vinyl greenhouse in Jeju, Korea. The symptoms occurred on the flowers and peduncles, on which a lot of gray molds were formed (Fig. 1A, B). The diseased flowers turned brown and blighted, and the diseased peduncles turned brown to black and rotted. As the disease progressed, black and long lesions were formed on the axes of inflorescences. In the vinyl greenhouse, five trees were observed, and 100 inflorescences of each tree were investi-

Research in Plant Disease eISSN 2233-9191 www.online-rpd.org gated for incidence of gray mold. The disease incidence on the inflorescences in the vinyl greenhouse ranged from 10% to 40%.

Mango inflorescences with gray mold symptoms were collected from the investigated vinyl greenhouse, and fungi were isolated from the diseased inflorescences. Conidial suspension was prepared from conidial masses on the gray mold symptoms using sterile distilled water and streaked on 2% water agar (WA) plate using a sterile loop. After incubation of the WA plate for 24 hr at 20°C, germinated conidia on WA were picked up under a dissecting microscope (Nikon SMZ 1780, Tokyo, Japan) and transferred to new WA plates. Three single-conidium isolates were obtained from the WA plate cultures after 5 days of incubation at 20°C. The isolates were cultured in potato dextrose agar (PDA) slants to use for identification and pathogenicity tests.

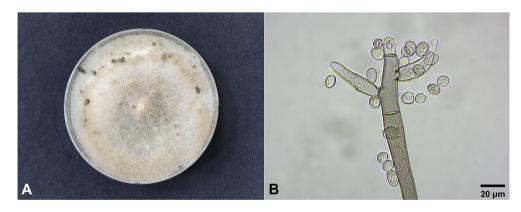
The three fungal isolates (MZBC-2201, MZBC-2202, and MZBC-2203) were cultured on PDA in 9 cm diameter Petri dishes at 20°C in the dark for 25 days to investigate their cultural and morphological characteristics. The colonies of the

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**Fig. 1.** Gray mold symptoms of mango trees. (A, B) Symptoms on the flowers and peduncles observed in the vinyl greenhouse. (C) Symptoms on the flowers and peduncles induced by artificial inoculation test with *Botrytis cinerea* isolate (MZBC-2202). (D) A non-inoculated tree (control).

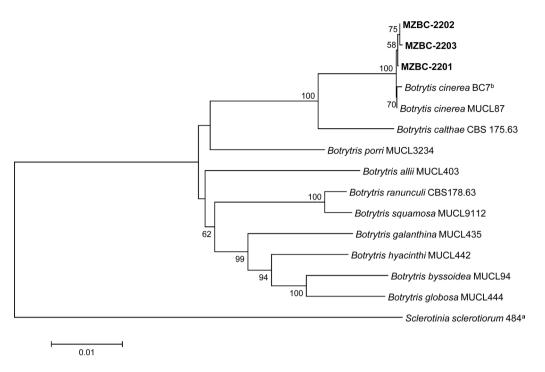


**Fig. 2.** Cultural and morphological features of *Botrytis cinerea* isolates from diseased mango trees. (A) A colony of an isolate (MZBC-2202) grown on potato dextrose agar at 20°C for 25 days. (B) A conidiophore and conidia.

isolates cultured on PDA consisted of white to gray mycelia and black, spherical or irregular sclerotia of which diameter was 1–7 mm (Fig. 2A). Fifty conidia and 25 conidiophores of each isolate were examined for their morphology. Conidia were hyaline to pale brown, globose or ellipsoidal, and measured 7–14×5–11 µm (av. 10.6×8.4 µm) (Fig. 2B). Conidiophores were erect, long more than 1 mm, 13–18 µm thick, branched, brown below, and paler near the apex. The mor-

phological characteristics of the isolates were similar to those of *Botrytis cinerea* described in a previous study (Ellis, 1971).

Phylogenetic analysis was conducted to confirm the identification of *B. cinerea* isolates based on their morphological characteristics. Genomic DNA of the isolates was extracted using the protocol in a previous study (Dong et al., 2022), with slight modifications. Polymerase chain reaction (PCR) products of glyceraldehyde-3-phosphate dehydrogenase



**Fig. 3.** Phylogenetic tree based on glyceraldehyde-3-phosphate dehydrogenase, heat-shock protein 60, and DNA-dependent RNA polymerase subunit II gene regions of the three isolates (MZBC-2201, MZBC-2202, and MZBC-2203) from mango and reference strains of *Botrytis* species. Sequence data of the reference species were obtained from the NCBI GenBank database. The phylogenetic tree was generated using the neighbor-joining method with a maximum composite likelihood model. The bootstrap support values more than 50 are shown at the nodes. The scale bar represents the number of nucleotide substitutions per site.

(G3PDH), heat-shock protein 60 (HSP60), and DNA-dependent RNA polymerase subunit II (RPB2) gene regions were obtained using the primer sets with the amplification conditions described in a previous study (Staats et al., 2005) and DNA Free-Multiplex Master Mix (Cellsafe, Yongin, Korea). Purification of the PCR products was done using the universal DNA purification kit (Tiangen, Beijing, China). Sequencing of the PCR products was conducted at Bionics Co., Ltd. (Seoul, Korea) with the same primers. The sequences were adjusted by SeqMan II (DNASTAR Inc., Madison, WI, USA) if necessary. Alignment of the sequences of the isolates from mango and other Botrytis spp. (Staats et al., 2005) was done using MUS-CLE (Edgar, 2004). Sclerotinia sclerotiorum 484 (Van Der Vlugt-Bergmans, 1993) was selected as an outgroup taxon. MEGA version 7 software (Kumar et al., 2016) was used to process and enhance the multiple sequence alignments, if necessary. Neighbor-joining analysis for concatenated alignments was conducted with maximum composite likelihood model performing 1,000 bootstrap replicates by MEGA version 7 software (Kumar et al., 2016). The phylogenetic analysis revealed that all isolates were clustered with the two strains (BC7<sup>b</sup> and MUCL87) of *B. cinerea* (Fig. 3). The sequence data

of G3PDH, HSP60, and RPB2 genes obtained from the three isolates were deposited in GenBank with accession numbers of OR469305–OR469307, OR469308–OR469310, and OR469311–OR469313, respectively.

The three isolates of *B. cinerea* were tested for their pathogenicity to inflorescences of mango trees through artificial inoculation. A conidial suspension (1–2×10<sup>6</sup> conidia/ml) of each isolate was prepared from the 30-day-old PDA cultures. A 40 ml of the conidial suspension of each isolate was sprayed onto inflorescences of a three-year-old mango tree grown in a circular plastic pot (height: 30 cm; diameter: 32 cm) in a vinyl greenhouse. A control mango tree was treated with 40 ml of sterile distilled water. The inoculated trees were covered with plastic bags of which inside was wetted with sterile distilled water and placed in a cultivation room at 18–22°C for 7 days. Thereafter, the plastic bags were removed from the inoculated trees, and the inoculated trees were placed in the cultivation room. Pathogenicity of the isolates was rated based on the degree of gray mold symptoms 10 days after inoculation. The inoculation test was conducted in three replicates. All the tested isolates caused gray mold symptoms on the flowers and peduncles of the inoculated trees (Fig. 1C), but no symptoms were observed on those of the control trees (Fig. 1D). The symptoms induced by the isolates were similar to those on the inflorescences of mango trees observed in the disease survey. Re-isolation of the inoculated isolates from the lesions was confirmed.

*B. cinerea* is known as a cosmopolitan gray mold and damages flowers, leaves, stems, fruits, etc. of many plants (Ellis, 1971). It has been reported that *B. cinerea* is the causal agent of gray mold on floral tissue of mango in Japan (Ajitomi et al., 2022) and causes stem end rot of mango fruit in Pakistan (Alam et al., 2017). The disease occurrence in many plants was reported in Korea (Korean Society of Plant Pathology, 2023). However, there has been no report on the disease occurrence in mango. This is the first report of *B. cinerea* causing gray mold in mango in Korea.

# **Conflicts of Interest**

No potential conflict of interest relevant to this article was reported.

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### References

- Ajitomi, A., Yamashiro, M., Takushi, T. and Ooshiro, A. 2022. *Botrytis cinerea* as the causal agent of grey mould on floral tissue of mango in Japan. *Aust. Plant Dis. Notes* 17: 18.
- Alam, M. W., Gleason, M. L., Aslam, S., Riaz, K. and Rehman, A. 2017. First report of *Botrytis cinerea* causing stem end rot of mango fruit in Pakistan. *Plant Dis.* 101: 255.
- Dong, L., Liu, S., Li, J., Tharreau, D., Liu, P., Tao, D. et al. 2022. A rapid and simple method for DNA preparation of *Magnaporthe oryzae* from single rice blast lesions for PCR-based molecular analysis. *Plant Pathol. J.* 38: 679-684.
- Edgar, R. C. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 32: 1792-1797.
- Ellis, M. B. 1971. Dematiaceous Hyphomycetes. Commonwealth Mycological Institute, Kew, Surrey, UK. 608 pp.
- Korean Society of Plant Pathology. 2023. List of Plant Diseases in Korea. URL http://genebank.rda.go.kr/kplantdisease.do [22 August 2023].
- Kumar, S., Stecher, G. and Tamura, K. 2016. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol. Biol. Evol.* 33: 1870-1874.
- Plants of the World Online. 2023. *Mangifera indica* L. Royal Botanic Garden, Kew. URL https://powo.science. kew.org/ [22 August 2023].
- Staats, M., Van Baarlen, P. and Van Kan, J. A. L. 2005. Molecular phylogeny of the plant pathogenic genus *Botrytis* and the evolution of host specificity. *Mol. Biol. Evol.* 22: 333-346.
- Van Der Vlugt-Bergmans, C. J. B., Brandwagt, B. F., Van't Klooster, J. W., Wagemakers, C. A. M. and Van Kan, J. A. L. 1993. Genetic variation and segregation of DNA polymorphisms in *Botrytis cinerea*. *Mycol. Res.* 97: 1193-1200.