

**REVIEW**

# The Need for Research on the Comparison of Sensory Characteristics between Cultured Meat Produced Using Scaffolds and Meat

Sol-Hee Lee and Jungseok Choi\*

Department of Animal Science, Chungbuk National University, Cheongju 28644, Korea

 OPEN ACCESS

**Received** November 7, 2023

**Revised** December 13, 2023

**Accepted** December 17, 2023

\***Corresponding author** : Jungseok Choi  
 Department of Animal Science, Chungbuk National University, Cheongju 28644, Korea  
 Tel: +82-43-261-2551  
 Fax: +82-43-261-2773  
 E-mail: [jchoi@chungbuk.ac.kr](mailto:jchoi@chungbuk.ac.kr)

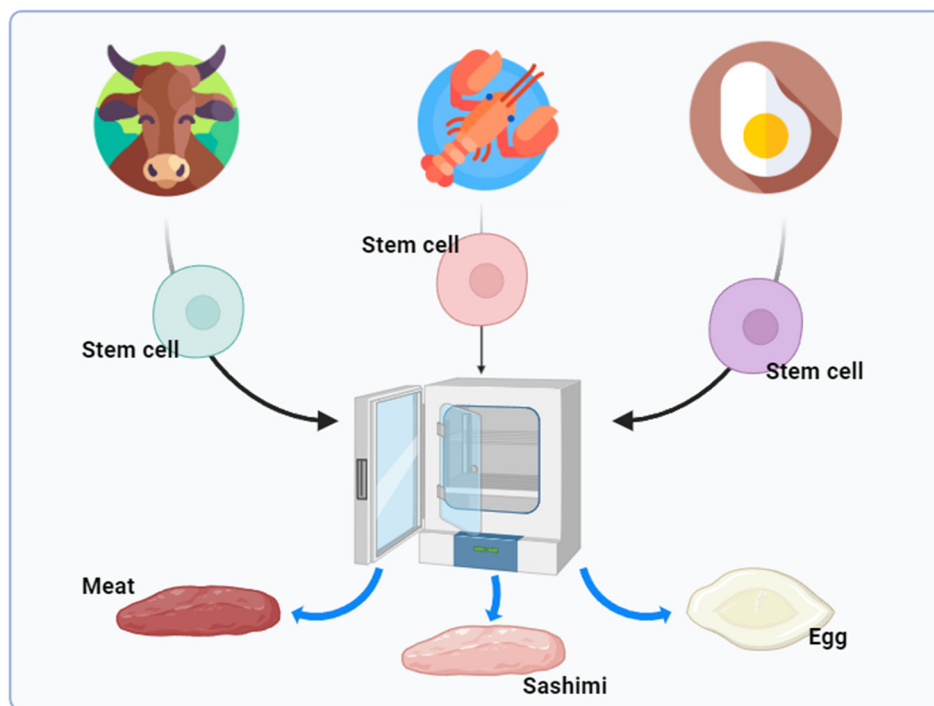
\***ORCID**  
 Sol-Hee Lee  
<https://orcid.org/0000-0003-1124-7095>  
 Jungseok Choi  
<https://orcid.org/0000-0001-8033-0410>

**Abstract** Cultured meat is one of the research areas currently in the spotlight in the agricultural and livestock industry, and refers to cells obtained from livestock that are proliferated and differentiated and processed into edible meat. These cell-cultured meats are mainly studied at the lab-scale by culturing them in flasks, and for commercial use, they are produced using scaffolds that mimic cell supports. Scaffolds are broadly divided into fiber scaffolds, hydrogels, and micro-carrier beads, and these are classified according to processing methods and materials. In particular, a scaffold is essential for mass production, which allows it to have appearance, texture, and flavor characteristics similar to meat. Because cultured meat is cultured in a state where oxygen is blocked, it may be lighter in color or produce less flavor substances than edible meat, but these can be compensated for by adding natural substances to the scaffolds or improving fat adhesion. In addition, it has the advantage of being able to express the texture characteristics of the scaffolds that make up the meat in various ways depending on the materials and manufacturing methods of the scaffolds. As a result, to increase consumers' preference for cultured meat and its similarity to edible meat, it is believed that manufacturing scaffolds taking into account the characteristics of edible meat will serve as an important factor. Therefore, continued research and interest in scaffolds is believed to be necessary.

**Keywords** cellular agriculture, scaffold, sensory evaluation

## Introduction

Cellular agriculture refers to the cells of agricultural products, such as meat, milk, eggs, and seafood, and is being studied as meat stem cell cultures (Fig. 1). The present outlook is that cell cultures of agricultural products will be needed not only in Korea but also worldwide for the following reasons. First, according to a United Nations (UN) report, the population will increase from 7.8 billion in 2021 to approximately 9.5 billion in 2050 (UN, 2015). As a result, there are concerns that a food shortage will occur, and in particular, the need for protein, which maintains and promotes the growth of the human body, will be approximately twice as high (Wikandari et al., 2021).



**Fig. 1.** Agricultural cell culture (cells from agricultural products, such as meat, milk, eggs, seafood, etc.).

Additionally, some vegetarians with animal welfare concerns may consider consuming cultured meat (Hopkins, 2015). Simply increasing the livestock population is not the clear answer to increases in protein needs, as this can cause many additional problems. As the number of livestock increases, the mass production of crops used as feed (corn, soybeans, etc.) follows, and environmental problems due to gases and excrement generated during livestock ingestion, the absorption of nutrients, and excretion arise (Herrero et al., 2016; Zhou, 2003). Culture meat may also be a way to solve two problems that may arise from livestock diseases: the loss of livestock due to highly infectious diseases, such as avian influenza, African swine fever, or foot-and-mouth disease, and instability in the price of livestock products (Chriki and Hocquette, 2020). Therefore, animal cell culture can be an important way to produce a certain amount of meat.

However, research on cellular agriculture needs to progress more actively and quickly. Until now, this research has mainly been conducted in medical fields such as medicine and pharmacy, especially histology. However, since the development of cultured meat by Professor Post of Mosa Meat in the Netherlands in 2013, cultured meat research has been conducted in various fields (Bodiou et al., 2020). Cultured meat still has various problems directly related to consumption and sales, such as being unfamiliar or causing feelings of distaste, and this is largely thought to be because consumers have not yet actually encountered cultured meat. In 2020, the Singapore Ministry of Food began selling cultured chicken breast developed by the American company Eat Just, and as of 2022, 156 cultured meat companies have been established around the world (Clare et al., 2022). Likewise, the mass production of cultured meat is being studied in every country except Singapore, and it is believed that the future of cultured meat can be accelerated if consumers are approached with a familiar image.

Meat was initially cultured by adhering cells to a flat flask. However, when the cells covered the flask, abnormal proliferation and differentiation occurred, or they died, making mass production difficult (Choi et al., 2020). This occurred because cells grown in a single layer have difficulties excreting waste products, and problems arise in the uptake of nutrients contained in the culture medium (Hubalek et al., 2022). Therefore, the need for cell culture using scaffolds that can replace

blood vessels has emerged (Kim et al., 2018). Additionally, because the scaffold has a three-dimensional (3D) shape, it has the additional advantage of being able to be manufactured in larger quantities compared to flask cultures. Research on different scaffold materials and methods is actively underway, and as the quality and quantity of cultured meat are produced differently, research is underway for mass cultivation. Additionally, because cultured meat should not only focus on mass production but also resemble edible meat, research focusing on the quality characteristics of cultured meat such as flavor, appearance, and texture, is necessary.

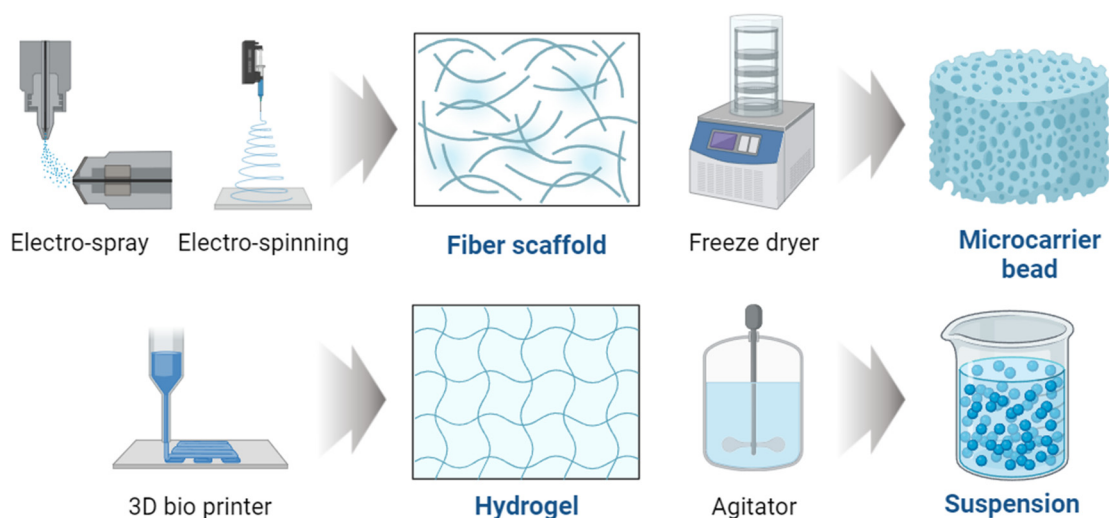
Therefore, in this review, we investigated the current manufacturing technology of scaffolds to develop cultured meat similar to edible meat, and previous studies on the appearance, flavor, and texture of cultured meat when scaffolds were used in cultured meat.

## Classification and Introduction of 3D Scaffolds

The scaffolds mainly used in cultured meat-cell culture include fiber scaffolds, hydrogels, and micro-carrier beads (MC) to maximize the adhesion ability of cells (Fig. 2). A suspension method (floating culture method), which suppresses the tendency of cells to attach and does not use a scaffold, can also be used. These scaffolds are all manufactured by imitating living tissue to directly deliver oxygen and nutrients to cells or remove waste (Bružauskaitė et al., 2016).

A fiber scaffold is a support in which thin and long fibers produced by electrospinning (ESI) form a matrix for cells to attach to, proliferate, and differentiate (Badami et al., 2006). ESI combines a variety of natural and synthetic polymers into fibers that are randomly or aligned according to the fiber diameter ( $\mu\text{m}$ ) or the setter's purpose (Bai et al., 2022). Fiber scaffolds also appear to have an excellent ability to form shapes because they can easily form muscle bundles, and their ability to form aligned fibers facilitates mass production (Feng et al., 2021). Since the proliferation and differentiation ability of cells varies as the fibers are arranged, research continues to determine which shape is most effective.

The second scaffold to be introduced is hydrogels, which are hydrophilic scaffolds made of one or more polymers. Like fiber scaffolds, they are supports that form a network structure to allow cells to adhere better to a planar substrate (Chimene et al., 2020). Hydrogel has the advantage of responding to cell secretion signals by forming bioactive ligands and requires a



**Fig. 2.** Introduction to the main scaffold manufacturing technologies and scaffolds produced.

high moisture content to maintain this ability (Rosales and Anseth, 2016). However, since hydrogel is mainly composed of animal/plant proteins, it can be degraded by the precursor proteins of extracellular proteolytic enzymes, such as matrix metalloproteinase proteins secreted from its own proteins (Samorezov and Alsberg, 2015). Therefore, an accurate analysis of manufacturing materials is required, and efforts are being made to improve chemical and structural aspects, such as porosity and elasticity (Myung et al., 2007). In addition, because hydrogel can retain a large amount of moisture, it is possible to mix substances such as water-soluble growth factors and hormones during design. Thus, research is underway to determine the mixing ratio.

MC have been developed from materials such as cellulose, gelatin, alginate, chitosan, and polystyrene, and most of the materials are used after being molded into a sphere, which induces electrostatic interaction with negatively charged cells through a positive charge coating on the cell attachment surface (Chang and Wang, 2011). In addition to electrostatic coating, the adhesion ability of cells is also improved through coating with proteins such as extracellular matrix. MC culture, similar to a suspension, requires additional mechanical functions, such as a stirrer, to help prevent cells from sticking to the wall. Although it may require more elements than other technologies, it is widely used in industry because it allows for the production of cells in large quantities compared to other culture techniques.

The suspension technique is the method most similar to 2D and has been used for the longest time in 3D cell culture. Like MC, suspended cells grow while floating in the culture medium and require machinery such as a stirrer or impeller (Fenge and Lüllau, 2005). In this method, cells are dispensed into a culture medium and allowed to grow on their own by supplying them with nutrients contained in the suspension or facilitating the excretion of waste products. Although it is easy to study the benefits of additives in suspension cultures, such as fetal bovine serum, and the ability of additives to participate in proliferation and differentiation, it has the disadvantage of being difficult to separate cells from the culture medium, thereby consuming the researcher's labor and time. Research is underway to easily distinguish cells from the culture medium when changing the culture medium and develop technologies, such as coatings or micro-wells, that can prevent adherent cells from sticking to the wall of the culture plates (Dang et al., 2002; Silk et al., 2010). Although this manuscript does not cover everything, various scaffolds can be used, depending on the cultured meat production technology and purpose.

## Scaffold Production Methods

The scaffolds described above are a type of support, and various machines and technologies exist to manufacture them, including ESI/spraying, 3D printing, molding, freeze drying, and decellularization, depending on the composition of the material and the researcher's design (Table 1).

ESI, which is mainly used to manufacture fiber scaffolds, can extract fibers with a diameter of 1 to several tens of  $\mu\text{m}$ s, and thus, can be used to create a fiber matrix, mold it to make MCs or manufacture various scaffolds (Pu et al., 2015). These nanofibers are produced by dry-spinning, which utilizes air or inert gas to evaporate residual solvents, and dry-jet wet spinning, which allows the polymer to orient and coalesce on the external surface before it hardens into shape. Wet-spinning, which solidifies, and melt-spinning, in which molten polymers are manufactured by exposing them to cooling air, are classified into various types depending on the material (Luo et al., 2012). The similar electro-spraying (E-SR) uses the same material as ESI. However, the material is emitted from the capillary itself rather than a nozzle during the spraying process. These two methods are similar in that they involve adding material to a capillary tube, a metal needle or spinneret, and a fiber collection device, but they are differentiated depending on the type of material.

**Table 1. Introduction of materials according to scaffold production method**

Production method	Materials	Cells	Introduction	References
Electrospinning	Textured soy protein	Bovine satellite cell	A step forward for the production of cell-based meat as food	Ben-Arye et al., 2020
Electrospinning	Polycaprolactone	Endothelial cell	Developing new scaffolds by investigating the interactions between endothelial cells, starch, and polyfibers	Santos et al., 2007
Electrospinning	Cellulose fibers	-	Systematic study of electrospinning conditions and application as reinforcing fiber for biocomposites	Han et al., 2008
Molding	Soybean powder and gelatin	C2C12 and 3T3-L1 cells	Manufacture cultured meat with a muscle-like texture by adding pre-fat cells that produce mass-produced cultured meat	Li et al., 2022
Molding	Polydimethylsiloxane and naringenin	Porcine satellite cells	Development of an efficient and innovative cultured meat production system through upregulation of signal transduction	Yan et al., 2022
Molding	Sodium alginate	Murine myoblast C2C12 cell line	Utility of a dual cross-linked alginate hydrogel system to support <i>in vitro</i> meat growth	Tahir and Floreani, 2022
3D printing	Salecan and $\kappa$ -carrageenan	Mouse fibroblast cells	A new strategy for fabricating and optimizing polysaccharide-based hydrogel scaffolds	Qi et al., 2020
3D printing	Sol-gel transition and ionic gelation	Mouse C2C12	Analyzing the adhesion of gellan gum and developing a new support accordingly	Koivisto et al., 2019
3D printing	Soy protein isolate	Primary bovine satellite cell	Great potential for research on cultivated meat through the use of peas, which have low allergenicity	Ianovici et al., 2022
Decellularization	Spinach	Primary bovine satellite cell	A cost-effective and environmentally friendly scaffold, potentially accelerating the development of laboratory-grown meat	Jones et al., 2021
Decellularization	Fresh whole jackfruit	Primary bovine satellite cell	Proposing a new closed bioreactor system for cellular agriculture products	Perreault et al., 2023
Decellularization	Grass blade	Murine C2C12 myoblasts	Presents the need for the development of inexpensive and sustainable support materials and structures	Allan et al., 2021
Freeze-drying	Gellan gum and guar gum	Mouse fibroblast cell (L929)	Development of a new scaffold that confirmed the cytotoxicity and cell adhesion of the scaffold	Anandan et al., 2019
Freeze-drying	Hydrolyzed collagen	-	Focus on using food-grade materials to ensure commercial availability of developed collagen ink formulations	Koranne et al., 2022
Freeze-drying	Collagen from turkey tendons	Skeletal muscle satellite cells	A major challenge for successful cultured meat production is the need for large quantities of skeletal muscle satellite cells	Andreassen et al., 2022

These methods are also similar to the 3D printing method, which is used in two distinct ways: fused deposition modeling (FDM) and extrusion modeling (EM; Placone and Engler, 2018; Pu'ad et al., 2020). FDM uses two nozzles, one for the material used in manufacturing and the other for the material used to maintain the shape of the material. Therefore, two nozzles

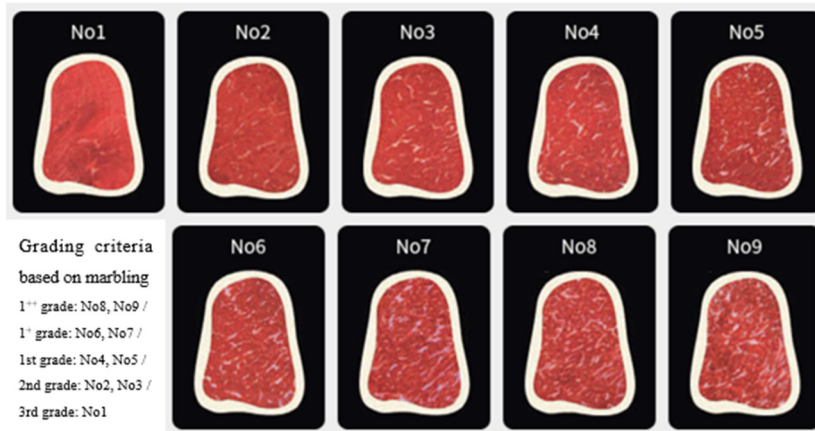
are used during production, and the shape-retaining material is later removed using a dedicated liquid (Ceretti et al., 2017). In contrast, EM uses a single material and is a method of developing a support by applying physical pressure. EM is divided into syringe-based extrusion, screw-based extrusion, and pneumatic extrusion (Guo et al., 2019), depending on the material it is manufactured from. The shape of the support produced differs depending on the density and physical properties, so it can be manufactured to reflect the researcher's exact purpose. In particular, the difference between the two is that FDM is mainly used to develop a scaffold, and EM uses the cells themselves as ink, enabling the direct production of cultured meat.

Molding refers to a method of molding a scaffold polymer to which cells are attached into a mold designed according to the researcher's intention. This includes molds formed by 3D printing (not a bioprinter), decellularization, and freeze-drying, in the same sense, MC is also used for this belongs (Ogawa et al., 2022). This scaffold has a relatively simple manufacturing method compared to other methods and is inexpensive because it can be used semi-permanently. Decellularization is a method in which cellulose-based cell walls are obtained using a chemical reaction, and nuclear material is removed from plant tissue (Toker-Bayraktar et al., 2023). This scaffold has a structure favorable for cell attachment and, like animal blood vessels, can promote cell growth because it is thin and has a large surface area (Walawalkar and Almelkar, 2021). In contrast, the freeze-drying method vaporizes water molecules to create a scaffold. Briefly, it is a method of mixing water and the solution used in the scaffold, molding it into the shape intended by the manufacturer, and then freeze-drying, which is a relatively simple process compared to other scaffolds (Chen et al., 2024). Cells attach and grow in the space created by vaporization and to freeze-drying is a widely used scaffold manufacturing method because mass production is possible.

## Appearance Characteristics of Cultured Meat

The production of excellent scaffolds is a basic step in cultured meat, and further analysis is required to determine how similar cultured meat and edible meat actually are. When purchasing meat, the first factor consumers consider is the appearance of the meat itself, such as red color, bright color, and a harmonious proportion of fat (Lee et al., 2020; Fig. 3). However, unlike meat, cultured meat grows in a culture medium and is cultured in oxygen-blocked conditions, so it has a light color (Fraeye et al., 2020). Therefore, it is important to form myoglobin protein, which is a factor in the red color of meat, because it is not present in cultured meat (Suman and Joseph, 2013). Color changes are induced by manipulating the culture conditions to replace myoglobin, such as adding extracellular heme protein or adding additional iron to the culture medium (Post and Hocquette, 2017; Siddiqui et al., 2022). Research is also being conducted to replace the red color using additives that are natural colorants, such as beets, carrots, tomatoes, and paprika (Bohrer, 2019; Grispoldi et al., 2022). Other studies are underway to increase the expression of myoglobin by culturing muscle fibers under hypoxic conditions. However, further research is needed (Moritz et al., 2015). Scaffolds are essential for cultured meat, and since the amount of scaffold is greater than that of the edible meat produced, studies are also being conducted to dye and use the scaffold itself. Since the materials of most scaffolds are transparent or white, they are easy to stain with dyes, colorants, such as hematoxylin, rhodamine, beet extract, and natural polyphenols (Bezjak et al., 2023; Xiang et al., 2022), and have visual dispersion effects when cells attach, proliferate, and differentiate. Marbling, another exterior characteristic, is another important consideration of consumers when purchasing a product. According to research by Killinger et al. (2004), when purchasing beef, selecting the degree of marbling differed depending on the fat content that buyers wanted. Therefore, it seems worth researching the proliferation and differentiation of fat cells, as well as the production technology of cultured meat with a variety of fat contents rather than a single fat content. Since the fat content in cultured meat can be selected, the development of a scaffold

Degree of fat distribution shown in the cross section of the *longissimus dorsi* muscle area



**Fig. 3.** Resources that may help create appearances similar to meat. Adapted from MAFRA (2023) with public domain.

that fat cells can easily attach to, such as polyunsaturated fat or omega-3, is also an area that needs to be continuously researched.

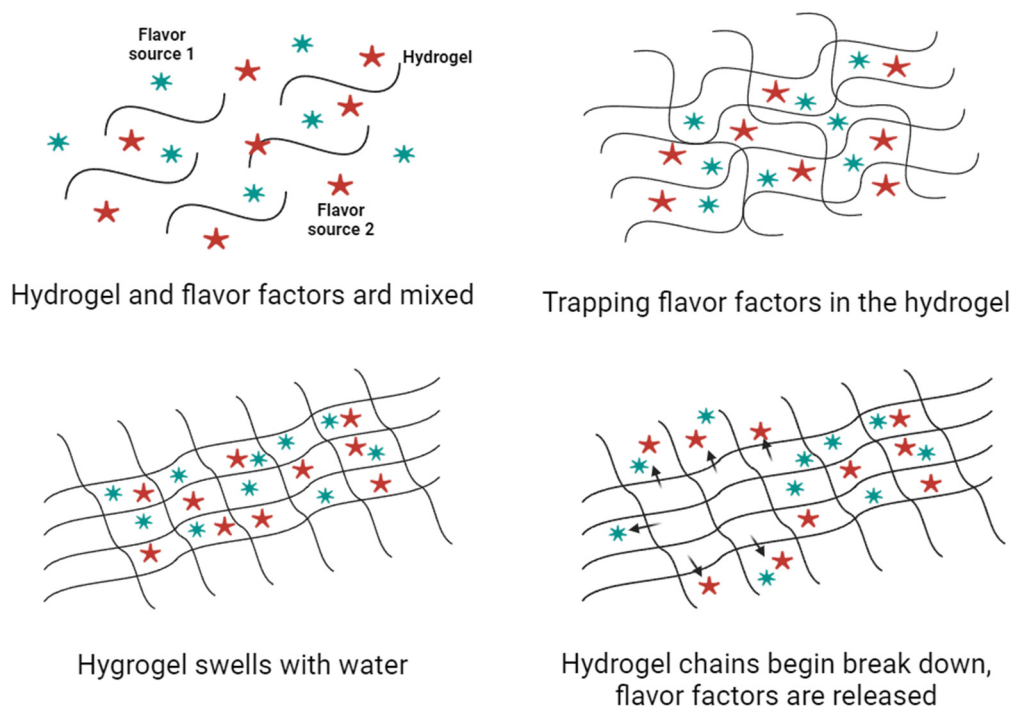
## Flavor Characteristics of Cultured Meat

The factor that meat buyers consider most important next to appearance is flavor. Therefore, flavor resulting from volatile compounds generated from intramuscular fat is important. In general, most consumers have aversions to new things and tend to look for characteristics they are familiar with (Stallberg-White and Pliner, 1999). The aroma and taste of meat are generated by volatile substances produced by the reactions of non-volatile components induced by heat. Water-soluble compounds with low molecular weight and meat lipids have been reported to be important factors in the taste of cooked meat (Khan et al., 2015). Additionally, since amino acids, such as methionine and cysteine, are important factors in meat flavor, their inclusion means that the flavor can be similar to that of regular meat (Yang et al., 2022). Since the flavors generated by these compounds or lipid states are different, consumers' preferences may also differ accordingly (Garmyn, 2020). Therefore,

cells and scaffolds should be developed in the cultured meat market with flavors similar to or superior to those of meat (Lee et al., 2022). Since scent is mainly expressed by fat cells, it is also important to have fat cells with an attractive scent. Song et al. (2022) and others conducted a study aimed at improving flavor due to the proliferation of fat cells, reporting that a scaffold made from peanut protein promoted the proliferation of mesenchymal stem cells derived from pig fat. In addition, as previously mentioned, many different scaffolds can be used to culture meat, so it is important to select the appropriate scaffold components well (Post et al., 2020). According to reports that binding some flavor precursors of fat cells to a scaffold could promote cell differentiation and improve the taste of the final product, scaffolds are being developed using mushrooms that produce meat flavor or microbial flora (Yalman et al., 2023; Zhang et al., 2022). For cultured meat to have a flavor similar to that of edible meat, the combination of aromatic substances with a support and the development of a scaffold to which fat cells with the main flavor of meat adhere well must be continuously pursued (Fig. 4).

## Textural Characteristics of Cultured Meat

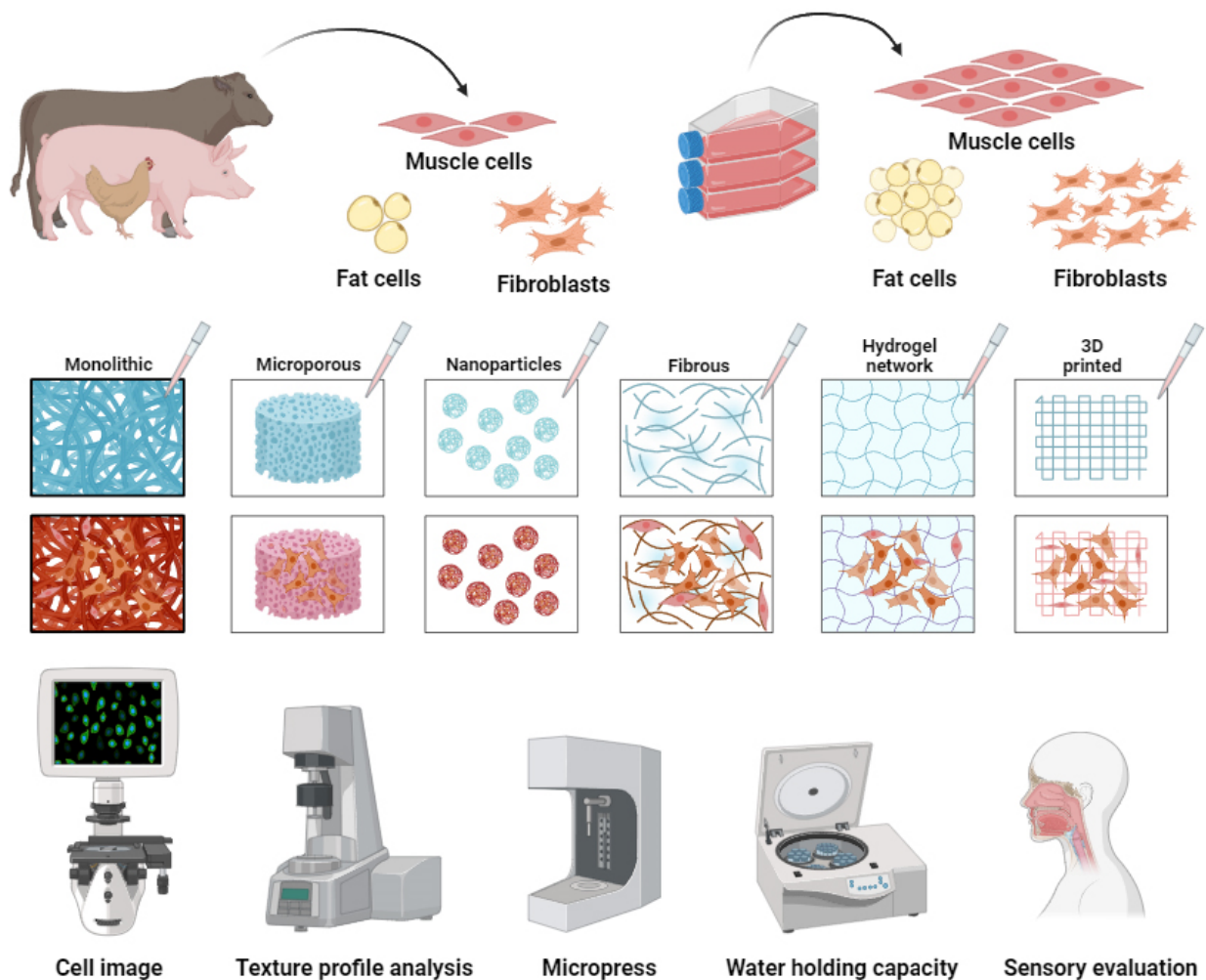
The most influential factor in the production of cultured meat using scaffolds is the textural characteristics of tissues such as cells, scaffolds, and fat. In addition, the composition and shape of the tissue are important because these affect sensory factors, such as the texture and juiciness of meat (Martinez et al., 2023). The factor that has the most practical influence on the scaffold of cultured meat is the tissue, and as shown in Chapters 1 and 2, the method of manufacturing the scaffold varies depending on the material (Szymczyk-Ziółkowska et al., 2020). Most 3D scaffolds are manufactured using natural materials, such as animal collagen, chitosan, and vegetable cellulose, as they are often consumed as-is after producing cultured meat (Moslemy et al., 2023). In most cases, collagen is made into hydrogel or MC, and because it is already extracted from animal protein, this scaffold has a texture similar to meat (Chen et al., 2023). However, when used as a cultured meat scaffold, if the



**Fig. 4.** Description of how to influence cells by conjugating flavor substances to the scaffold. Adapted from Blackwood et al. (2012) with CC-BY.



concentration is too high or the amount is too large, hardness increases, and the textural preference is lower than that of meat (Grønlien et al., 2019). de la Cruz Bosques et al. (2023) stated that when bovine pericardium is decellularized and used as a scaffold, large amounts of cultured meat can be produced at a low cost. Soybean, a vegetable protein, has been used as a textured vegetable protein to mimic meat, and many recent studies have shown that it can be used as a scaffold (Ben-Arye et al., 2020; Guan et al., 2023). However, in the case of plant proteins, there is no arginine-glycine-aspartic acid sequence that allows cells to attach, so coating with animal protein is necessary to enable the cells to attach (Lee et al., 2022). In addition, decellularized cell tissues were reported to require animal proteins, such as collagen, or plant proteins, such as alginate, to attach to cells (Brown et al., 2017). Vegetable protein itself has a lower hardness than regular meat and may have a slightly heterogeneous feeling, so additional research is needed. Alternatively, cells may be attached through electrostatic biocompatibility. This production method is known to have a texture more similar to real meat than the method described above (Ravishankar et al., 2019). Yen et al. (2023) and others reported that cultured meat produced with MC made of 2% chitosan and 0.2%–0.3% collagen showed low hardness, elasticity, and cohesion in the raw state but that these factors could be improved in the cooked state. Therefore, cultured meat should be produced by manufacturing scaffolds using appropriate concentrations and processing methods to improve appearance, flavor, and texture (Fig. 5).



**Fig. 5.** Textural characteristics analysis method according to cultured meat produced using different scaffold manufacturing techniques.

## Conclusion

Cultured meat appears to be valuable as a future protein because it can solve various problems, such as food and environmental problems, not only in Korea but also around the world. Because the impact on the future protein market varies depending on the level and extent of current development, various efforts, such as symposiums and seminars on future proteins, are currently ongoing. Globally, the research and development of cultured meat continues to increase. Accordingly, Korea has set a goal to mass produce cultured meat, and many companies are starting up or collaborating to produce cultured meat. Among the various methods for mass production, scaffold technology, mostly manufactured from edible materials, is described in this manuscript. When culturing muscle cells using a scaffold, more cells can be attached and mass-produced compared to 2D. These scaffolds have various manufacturing methods (e.g., ESI, E-SR, 3D printing, molding, decellularization, and freeze drying) using animal, vegetable, and chemical materials. Fiber scaffolds, hydrogels, MC, and suspensions made through these manufacturing methods utilize the adhesion of cells to enable mass culture. However, if the cultured meat produced in this way is different from actual meat, it may not be preferred by consumers, so additional research is needed. First, when muscle cells proliferate and differentiate, they are submerged in a culture medium and grow in an anaerobic state, resulting in a lack of myoglobin, a pigment-protein. Therefore, the appearance (red, light, dark, etc.) can be supplemented using natural colorants or adding heme protein or iron to the scaffold. In addition, research on flavor and taste, such as producing scaffolds with strains that exhibit the flavor of meat or attaching precursors related to the flavor of the scaffold, is ongoing. Lastly, cultured meat manufactured using scaffolds is also related to texture because, like regular meat, the scaffold (tendon, etc.) affects the texture. Therefore, it is important to control the concentration and strength of the material and the thickness and hardness of the scaffold to achieve a texture similar to that of meat. Thus, since the use of a scaffold is inevitable for the mass production of cultured meat, research on scaffolds, as well as research to produce products that are similar to meat in external factors such as appearance, flavor, and texture while using the scaffold, need to continue.

## Conflicts of Interest

The authors declare no potential conflicts of interest.

## Acknowledgements

This work was supported by the Korea Institute of Planning and Evaluation for Technology in Food, Agriculture and Forestry (IPET) through the High Value-added Food Technology Development Project, funded by the Ministry of Agriculture, Food, and Rural Affairs (MAFRA) (321028-5). This work was also supported by the “Regional Innovation Strategy (RIS)” through the National Research Foundation of Korea (NRF), funded by the Ministry of Education (MOE) (2021RIS-001).

## Author Contributions

Conceptualization: Lee SH, Choi J. Data curation: Lee SH, Choi J. Formal analysis: Lee SH. Methodology: Lee SH, Choi J. Software: Lee SH. Validation: Lee SH, Choi J. Investigation: Lee SH, Choi J. Writing - original draft: Lee SH. Writing - review & editing: Lee SH, Choi J.

## Ethics Approval

This article does not require IRB/IACUC approval because there are no human and animal participants.

## References

- Allan SJ, Ellis MJ, De Bank PA. 2021. Decellularized grass as a sustainable scaffold for skeletal muscle tissue engineering. *J Biomed Mater Res A* 109:2471-2482.
- Anandan D, Madhumathi G, Nambiraj NA, Jaiswal AK. 2019. Gum based 3D composite scaffolds for bone tissue engineering applications. *Carbohydr Polym* 214:62-70.
- Andreassen RC, Rønning SB, Solberg NT, Grønlien KG, Kristoffersen KA, Høst V, Kolset SO, Pedersen ME. 2022. Production of food-grade microcarriers based on by-products from the food industry to facilitate the expansion of bovine skeletal muscle satellite cells for cultured meat production. *Biomater* 286:121602.
- Badami AS, Kreke MR, Thompson MS, Riffle JS, Goldstein AS. 2006. Effect of fiber diameter on spreading, proliferation, and differentiation of osteoblastic cells on electrospun poly (lactic acid) substrates. *Biomaterials* 27:596-606.
- Bai Y, Liu Y, Lv H, Shi H, Zhou W, Liu Y, Yu DG. 2022. Processes of electrospun polyvinylidene fluoride-based nanofibers, their piezoelectric properties, and several fantastic applications. *Polymers* 14:4311.
- Ben-Arye T, Shandalov Y, Ben-Shaul S, Landau S, Zagury Y, Ianovici I, Lavon N, Levenberg S. 2020. Textured soy protein scaffolds enable the generation of three-dimensional bovine skeletal muscle tissue for cell-based meat. *Nat Food* 1:210-220.
- Bezjak D, Orellana N, Valdin scaffolds enable the do CA. 2023. Towards understanding the role of microstructured edible scaffolds for cultured meat production. *Food Bioprocess Technol* (in press). doi: 10.1007/s11947-023-03166-2.
- Blackwood KA, Bock N, Dargaville TR, Ann Woodruff M. 2012. Scaffolds for growth factor delivery as applied to bone tissue engineering. *Int J Polym Sci* 2012:174942.
- Bodiou V, Moutsatsou P, Post MJ. 2020. Microcarriers for upscaling cultured meat production. *Front Nutr* 7:10.
- Bohrer BM. 2019. An investigation of the formulation and nutritional composition of modern meat analogue products. *Food Sci Hum Wellness* 8:320-329.
- Brown BN, Buckenmeyer MJ, Prest TA. 2017. Preparation of decellularized biological scaffolds for 3D cell culture. *Methods Mol Biol* 1612:15-27.
- Bružauskaitė I, Bironaitė D, Bagdonas E, Bernotienė E. 2016. Scaffolds and cells for tissue regeneration: different scaffold pore sizes—different cell effects. *Cytotechnology* 68:355-369.
- Ceretti E, Ginestra P, Neto PI, Fiorentino A, Da Silva JVL. 2017. Multi-layered scaffolds production via fused deposition modeling (FDM) using an open source 3D printer: Process parameters optimization for dimensional accuracy and design reproducibility. *Procedia CIRP* 65:13-18.
- Chang HI, Wang Y. 2011. Cell responses to surface and architecture of tissue engineering scaffolds. In *Regenerative medicine and tissue engineering: Cells and biomaterials*. Eberli D (ed). InTechOpen, London, UK. pp 570-572.
- Chen Y, Li L, Chen L, Shao W, Chen X, Fan X, Liu Y, Ding S, Xu X, Zhou G, Feng X. 2023. Gellan gum-gelatin scaffolds with Ca<sup>2+</sup> crosslinking for constructing a structured cell cultured meat model. *Biomaterials* 299:122176.
- Chen Y, Zhang W, Ding X, Ding S, Tang C, Zeng X, Wang J, Zhou G. 2024. Programmable scaffolds with aligned porous structures for cell cultured meat. *Food Chem* 430:137098.

- Chimene D, Kaunas R, Gaharwar AK. 2020. Hydrogel bioink reinforcement for additive manufacturing: A focused review of emerging strategies. *Adv Mater* 32:1902026.
- Choi KH, Yoon JW, Kim M, Lee HJ, Jeong J, Ryu M, Jo C, Lee CK. 2021. Muscle stem cell isolation and *in vitro* culture for meat production: A methodological review. *Compr Rev Food Sci Food Saf* 20:429-457.
- Chriki S, Hocquette JF. 2020. The myth of cultured meat: A review. *Front Nutr* 7:7.
- Clare K, Maani N, Milner J. 2022. Meat, money and messaging: How the environmental and health harms of red and processed meat consumption are framed by the meat industry. *Food Policy* 109:102234.
- Dang SM, Kyba M, Perlingeiro R, Daley GQ, Zandstra PW. 2002. Efficiency of embryoid body formation and hematopoietic development from embryonic stem cells in different culture systems. *Biotechnol Bioeng* 78:442-453.
- de la Cruz Bosques JA, Ibarra Ses JArliJ, Mendoza-Novelo B, Segovia-Hernandez JG, Molina-Guerrero CE. 2023. Profitability of chemically cross-linked collagen scaffold production using bovine pericardium: Revaluing waste from the meat industry for biomedical applications. *Polymers* 15:2797.
- Feng W, Zhang Y, Shao Y, Huang T, Zhang N, Yang J, Qi X, Wang Y. 2021. Coaxial electrospun membranes with thermal energy storage and shape memory functions for simultaneous thermal/moisture management in personal cooling textiles. *Eur Polym J* 145:110245.
- Fenge C, LüC, L E. 2005. Cell culture bioreactors. In *Cell culture technology for pharmaceutical and cell-based therapies*. 1<sup>st</sup> ed. Ozturk S, Hu WS (ed). CRC Press, Boca Raton, FL, USA. pp 173-242.
- Fraeye I, Kratka M, Vandemburgh H, Thorrez L. 2020. Sensorial and nutritional aspects of cultured meat in comparison to traditional meat: Much to be inferred. *Front Nutr* 7:35.
- Garmyn A. 2020. Consumer preferences and acceptance of meat products. *Foods* 9:708.
- Grispoldi L, Karama M, El-Ashram S, Saraiva C, García-Díez J, Chalias A, De Gennis M, Vannuccini A, Poerio G, Torlai P, Chianese G, Fermani AG, Barbera S, Cenci-Goga BT. 2022. A study on the application of natural extracts as alternatives to sodium nitrite in processed meat. *J Food Process Preserv* 46:e16351.
- Grønlien KG, Pedersen ME, Sanden KW, Høst V, Karlsen J, Tønnesen HH. 2019. Collagen from Turkey (*Meleagris gallopavo*) tendon: A promising sustainable biomaterial for pharmaceutical use. *Sustain Chem Pharm* 13:100166.
- Guan X, Yan Q, Ma Z, Zhou J. 2023. Production of mature myotubes *in vitro* improves the texture and protein quality of cultured pork. *Food Funct* 14:3576-3587.
- Guo CF, Zhang M, Bhandari B. 2019. A comparative study between syringe-based and screw-based 3D food printers by computational simulation. *Comput Electron Agric* 162:397-404.
- Han SO, Son WK, Youk JH, Park WH. 2008. Electrospinning of ultrafine cellulose fibers and fabrication of poly(butylene succinate) biocomposites reinforced by them. *J Appl Polym Sci* 107:1954-1959.
- Herrero M, Henderson B, Havlík P, Thornton PK, Conant RT, Smith P, Wiersenius S, Hristov AN, Gerber P, Gill M, Butterbach-Bahl K, Valin H, Garnett T, Stehfest E. 2016. Greenhouse gas mitigation potentials in the livestock sector. *Nat Clim Change* 6:452-461.
- Hopkins PD. 2015. Cultured meat in western media: The disproportionate coverage of vegetarian reactions, demographic realities, and implications for cultured meat marketing. *J Integr Agric* 14:264-272.
- Hubalek S, Post MJ, Moutsatsou P. 2022. Towards resource-efficient and cost-efficient cultured meat. *Curr Opin Food Sci* 47:100885.
- Ianovici I, Zagury Y, Redenski I, Lavon N, Levenberg S. 2022. 3D-printable plant protein-enriched scaffolds for cultivated

- meat development. *Biomaterials* 284:121487.
- Jones JD, Rebello AS, Gaudette GR. 2021. Decellularized spinach: An edible scaffold for laboratory-grown meat. *Food Biosci* 41:100986.
- Khan MI, Jo C, Tariq MR. 2015. Meat flavor precursors and factors influencing flavor precursors: A systematic review. *Meat Sci* 110:278-284.
- Killinger KM, Calkins CR, Umberger WJ, Feuz DM, Eskridge KM. 2004. Consumer visual preference and value for beef steaks differing in marbling level and color. *J Anim Sci* 82:3288-3293.
- Kim W, Kim M, Kim GH. 2018. 3D-printed biomimetic scaffold simulating microfibril muscle structure. *Adv Funct Mater* 28:1800405.
- Koivisto JT, Gering C, Karvinen J, Cherian RM, Belay B, Hyttinen J, Aalto-Setälä K, Kellomäki M, Parraga, J. 2019. Mechanically biomimetic gelatin–gellan gum hydrogels for 3D culture of beating human cardiomyocytes. *ACS Appl Mater Interfaces* 11:20589-20602.
- Koranne V, Jonas OLC, Mitra H, Bapat S, Ardekani AM, Sealy MP, Rajurkar K, Malshe AP. 2022. Exploring properties of edible hydrolyzed collagen for 3D food printing of scaffold for biomanufacturing cultivated meat. *Procedia CIRP* 110:186-191.
- Lee HJ, Yong HI, Kim M, Choi YS, Jo C. 2020. Status of meat alternatives and their potential role in the future meat market: A review. *Asian-Australas J Anim Sci* 33:1533-1543.
- Lee M, Park S, Choi B, Kim J, Choi W, Jeong I, Han D, Koh WG, Hong J. 2022. Tailoring a gelatin/agar matrix for the synergistic effect with cells to produce high-quality cultured meat. *ACS Appl Mater Interfaces* 14:38235-38245.
- Li CH, Yang IH, Ke CJ, Chi CY, Matahum J, Kuan CY, Celikkin N, Swieszkowski W, Lin FH. 2022. The production of fat-containing cultured meat by stacking aligned muscle layers and adipose layers formed from gelatin-soymilk scaffold. *Front Bioeng Biotechnol* 10:875069.
- Luo CJ, Stoyanov SD, Stride E, Pelan E, Edirisinghe M. 2012. Electrospinning *versus* fibre production methods: From specifics to technological convergence. *Chem Soc Rev* 41:4708-4735.
- Martinez HA, Miller RK, Kerth C, Wasser BE. 2023. Prediction of beef tenderness and juiciness using consumer and descriptive sensory attributes. *Meat Sci* 205:109292.
- Ministry of Agriculture, Food and Rural Affairs. [MAFRA]. 2023. Detailed standards for grading livestock products. Available from: <https://www.law.go.kr/%ED%96%89%EC%A0%95%EA%B7%9C%EC%B9%99/%EC%B6%95%EC%82%B0%EB%AC%BC%20%EB%93%B1%EA%B8%89%ED%8C%90%EC%A0%95%20%EC%84%B8%EB%B6%80%EA%B8%B0%EC%A4%80>. Accessed at Sep 30, 2023.
- Moritz MSM, Verbruggen SEL, Post MJ. 2015. Alternatives for large-scale production of cultured beef: A review. *J Integr Agric* 14:208-216.
- Moslemy N, Sharifi E, Asadi-Eydivand M, Abolfathi N. 2023. Review in edible materials for sustainable cultured meat: Scaffolds and microcarriers production. *Int J Food Sci Technol* 58:6182-6191.
- Myung D, Koh W, Bakri A, Zhang F, Marshall A, Ko J, Noolandi J, Carrasco M, Cochran JR, Frank CW, Ta CN. 2007. Design and fabrication of an artificial cornea based on a photolithographically patterned hydrogel construct. *Biomed Microdevices* 9:911-922.
- Ogawa M, Moreno García J, Nitin N, Baar K, Block DE. 2022. Assessing edible filamentous fungal carriers as cell supports for growth of yeast and cultivated meat. *Foods* 11:3142.

- Perreault LR, Thyden R, Kloster J, Jones JD, Nunes J, Patmanidis AA, Reddig D, Dominko T, Gaudette GR. 2023. Repurposing agricultural waste as low-cost cultured meat scaffolds. *Front Food Sci Technol* 3:1208298.
- Placone JK, Engler AJ. 2018. Recent advances in extrusion-based 3D printing for biomedical applications. *Adv Healthc Mater* 7:1701161.
- Post MJ, Hocquette JF. 2017. New sources of animal proteins: Cultured meat. In *New aspects of meat quality*. Purslow PP (ed). Woodhead, Sawston, UK. pp 425-441.
- Post MJ, Levenberg S, Kaplan DL, Genovese N, Fu J, Bryant CJ, Negowetti N, Verzijden K, Moutsatsou P. 2020. Scientific, sustainability and regulatory challenges of cultured meat. *Nat Food* 1:403-415.
- Pu J, Yuan F, Li S, Komvopoulos K. 2015. Electrospun bilayer fibrous scaffolds for enhanced cell infiltration and vascularization *in vivo*. *Acta Biomater* 13:131-141.
- Pu'ad NASM, Haq RHA, Noh HM, Abdullah HZ, Idris MI, Lee TC. 2020. Review on the fabrication of fused deposition modelling (FDM) composite filament for biomedical applications. *Mater Today Proc* 29:228-232.
- Qi X, Su T, Zhang M, Tong X, Pan W, Zeng Q, Zhou Z, Shen L, He X, Shen J. 2020. Macroporous hydrogel scaffolds with tunable physicochemical properties for tissue engineering constructed using renewable polysaccharides. *ACS Appl Mater Interfaces* 12:13256-13264.
- Ravishankar K, Venkatesan M, Desingh RP, Mahalingam A, Sadhasivam B, Subramaniam R, Dhamodharan R. 2019. Biocompatible hydrogels of chitosan-alkali lignin for potential wound healing applications. *Mater Sci Eng C* 102:447-457.
- Rosales AM, Anseth KS. 2016. The design of reversible hydrogels to capture extracellular matrix dynamics. *Nat Rev Mater* 1:15012.
- Samorezov JE, Alsberg E. 2015. Spatial regulation of controlled bioactive factor delivery for bone tissue engineering. *Adv Drug Deliv Rev* 84:45-67.
- Santos MI, Fuchs S, Gomes ME, Unger RE, Reis RL, Kirkpatrick CJ. 2007. Response of micro- and macrovascular endothelial cells to starch-based fiber meshes for bone tissue engineering. *Biomaterials* 28:240-248.
- Siddiqui SA, Bahmid NA, Karim I, Mehany T, Gvozdenko AA, Blinov AV, Nagdalian AA, Arsyad M, Lorenzo JM. 2022. Cultured meat: Processing, packaging, shelf life, and consumer acceptance. *LWT-Food Sci Technol* 172:114192.
- Silk NJ, Denby S, Lewis G, Kuiper M, Hatton D, Field R, Baganz F, Lye GJ. 2010. Fed-batch operation of an industrial cell culture process in shaken microwells. *Biotechnol Lett* 32:73-78.
- Song WJ, Liu PP, Zheng YY, Meng ZQ, Zhu HZ, Tang CB, Li HX, Ding SJ, Zhou GH. 2022. Production of cultured fat with peanut wire-drawing protein scaffold and quality evaluation based on texture and volatile compounds analysis. *Food Res Int* 160:111636.
- Stallberg-White C, Pliner P. 1999. The effect of flavor principles on willingness to taste novel foods. *Appetite* 33:209-221.
- Suman SP, Joseph P. 2013. Myoglobin chemistry and meat color. *Annu Rev Food Sci Technol* 4:79-99.
- Szymczyk-Ziółkowska P, Łabowska MB, Detyna J, Michalak I, Gruber P. 2020. A review of fabrication polymer scaffolds for biomedical applications using additive manufacturing techniques. *Biocybern Biomed Eng* 40:624-638.
- Tahir I, Floreani R. 2022. Dual-crosslinked alginate-based hydrogels with tunable mechanical properties for cultured meat. *Foods* 11:2829.
- Toker-Bayraktar M, Erenay B, Altun B, Odabaş S, Garipcan B. 2023. Plant-derived biomaterials and scaffolds. *Cellulose* 30:2731-2751.

- United Nations [UN]. 2015. World population prospects: The 2015 revision. Available from: [https://population.un.org/wpp/publications/files/key\\_findings\\_wpp\\_2015.pdf](https://population.un.org/wpp/publications/files/key_findings_wpp_2015.pdf). Accessed at Sep 25, 2023.
- Walawalkar S, Almelkar S. 2021. Fabricating a pre-vascularized large-sized metabolically-supportive scaffold using *Brassica oleracea* leaf. *J Biomater Appl* 36:165-178.
- Wikandari R, Manikharda, Baldermann S, Ningrum A, Taherzadeh MJ. 2021. Application of cell culture technology and genetic engineering for production of future foods and crop improvement to strengthen food security. *Bioengineered* 12:11305-11330.
- Xiang N, Yuen JSK Jr, Stout AJ, Rubio NR, Chen Y, Kaplan DL. 2022. 3D porous scaffolds from wheat glutenin for cultured meat applications. *Biomaterials* 285:121543.
- Yalman S, Trapp T, Vetter C, Popa F, Fraatz MA, Zorn H. 2023. Formation of a meat-like flavor by submerged cultivated *Laetiporus montanus*. *J Agric Food Chem* 71:8083-8092.
- Yan Q, Fei Z, Li M, Zhou J, Du G, Guan X. 2022. Naringenin promotes myotube formation and maturation for cultured meat production. *Foods* 11:3755.
- Yang Y, Li J, Jia X, Zhao Q, Ma Q, Yu Y, Tang C, Zhang J. 2022. Characterization of the flavor precursors and flavor fingerprints in grazing lambs by foodomics. *Foods* 11:191.
- Yen FC, Glusac J, Levi S, Zernov A, Baruch L, Davidovich-Pinhas M, Fishman A, Machluf M. 2023. Cultured meat platform developed through the structuring of edible microcarrier-derived microtissues with oleogel-based fat substitute. *Nat Commun* 14:2942.
- Zhang C, Shi L, Dai J. 2022. Cultured meat from biomaterials: Challenges and prospects. *Synth Biol J* 3:676-689.
- Zhou ZY. 2003. Feed versus food: The future challenge and balance for farming. 1<sup>st</sup> ed. Parliament House, Canberra, Australia. pp 40-42.